

Novel 3-D In Vitro Models for Studying Pancreatic Cancer Drug Response and Resistance

Naomi Walsh, MPH, PhD Associate Professor, Life Sciences Institute, School of Biotechnology, Dublin City University

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Utilization of novel 3D *in vitro* models for studying pancreatic cancer drug response and resistance

Naomi Walsh PhD MPH Associate Professor, Life Sciences Institute, School of Biotechnology, Dublin City University Ireland



Objectives/Key Points



Describe the role of preclinical systems focusing on organoids in modelling pancreatic cancer



Present a method for the establishment of isogenic primary cell lines from patient derived organoids (PDOs) and the recapitulation to 3D cell line organoids (CLOs)



Demonstrate that 3D CLO culture method can be used as an expandable, easy scale-up, affordable, and less time-consuming research model



Highlight a methodology for the development and characterization of drug resistance using pancreatic cancer organoids

Pancreatic cancer



- Pancreatic ductal adenocarcinoma (PDAC) is the most common type
- Aggressive, poor prognosis with 5-year survival rate 13%, distant metastases 3%*.
- Diagnosed at late state
 - Silent progression
 - Nonspecific symptoms
 - More than half of patients diagnosed at an advanced stage
- Treatment
 - Surgery offers curative intent however, approximately 20% patients are operable
 - Chemotherapy and/or radiotherapy as the standard of care for most patients.
 - Inherent or acquired resistance
- 7th leading cause of cancer mortality in the World
 - Increasing trends in incidence and mortality of PDAC across the World



* SEER = Surveillance, Epidemiology, and End Results (2013-2019)

Greatest challenge in cancer treatment

- Predict response because cancer and its treatment are patient-specific
- Personalised medicine, individual prevention, and treatment strategies based on unique patient-specific variables



Landscape of PDAC

- Genomic and transcriptomic subtypes can enrich for therapeutic vulnerabilities
 - > 90% cases present with KRAS mutation
 - *TP53*, *SMAD4* and *CDKN2A* inactivated in > 50%
 - Limited success in targeted therapy strategies
- Familial predisposition
 - Genetic syndromes account for 5-10% of PDAC
 - Germline mutations in DNA damage repair pathway (BRCA1/BRCA2 etc)

Cancer Genome Atlas Research Network. *Cancer Cell*. 2017;32(2):185-203.e13. Stoof et al. *Frontiers in cell and developmental biology*. 2021;9, 749490.





Preclinical models of pancreatic cancer



Cancer organoids as models of disease

- Miniaturized and simplified 3D structures
- Patient specific as derived from patient tissue
- Self-organizing, resembling the original tumor architecture
- Show heterogeneity of tumors
- Long-term growth potential biobanking
- Alternative to animal models



PDOs in personalized medicine





Personalized therapy development – providing accurate and reliable drug screening systems

Uncover **underlying mechanisms driving cancer progression** – genetic mutations, signalling pathways etc.

Can be developed into **more complex models to mimic tumor microenvironment**

Platform to study early and late stages of tumor development

Developing 3D organoid systems to model pancreatic cancer

Development of 3D organoid models | 37 °C Removal of infiltrating xenograft mouse cells

Schematic: Sara Noorani (Biorender) Nelson et al., *Scientific reports*, 2020;*10*(1), 2778

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Development of primary 2D cell lines from organoids



Generation of organoid-derived primary cell lines and 3D cell line organoids (CLOs)



- Developed three new primary 2D cell lines derived from 3D organoids purchased from ATCC
 - HCM-CSHL-0090-C25 (ATCC® PDM-37[™])
 - HCM-CSHL-0094-C25 (ATCC® PDM-41[™])
 - HCM-BROD-0008-C25 (ATCC[®] PDM-106[™])
- 2D cell lines were expanded over 2 passages and recapitulated to cell line organoids (CLOs) using 3D organoid culture conditions

CLOs maintain the phenotypic and growth characteristics of similar to organoids



Differential proliferation rate observed between the three cell lines

Comparison of proliferation between CLOs and respective isogenic matched organoid revealed similar rates of proliferation between PDM37 and PDM106 models;

PDM41-CLO proliferated faster compared to PDM41-organoid

Noorani et al., Organoids. 2022; 1(2):168-183.

CLOs retain therapeutic drug response comparable to derived organoids



Noorani et al., Organoids. 2022; 1(2):168-183.

Stem cell marker expression in 2D cell line, CLOs and organoids



Overexpression of Cancer Stem Cell markers in 3D models compared to 2D cell lines ALDH1A1 CXCR4 HCAM (CD44) **EpCAM (ESA)** В. Α. C. D. PDM41 **PDM106** PDM41 **PDM106** PDM41 **PDM106** PDM41 PDM106 Cell line Cell line Cell line Cell line CLO CLO CLO CLO Organoid Organoid Organoid Organoid ** **** **** rescent Units (RFU) - 05 - 05 (RFU) 80 (RFU) 80. **** (RFU) 20 Relative Fluorescent Units (RFU) (RFU) 20-(RFU) **** (RFU) 30-**** *** **** 100 the second Units (scent Units (-07 (10115 (ent Units 25-20-15-10-40. 5-CLO CLO 00°.

Noorani et al., Organoids. 2022; 1(2):168-183.

RNA-Seq transcriptomic analysis identifies similar CLO and organoid signature





1.5

2.5

3.5

 $Log_{10}(CL - Expression)$

4.5

5.5

Nelson et al., Scientific reports, 2020;10(1), 2778

CLOs accurately reflect the cellular architecture and heterogeneity of organoids *in vivo*

Α. Β. Grouped relative weight change Grouped tumour volume 1500-1.3 Organoid Organoid Relative weight change Tumour volume (mm³) CLO CLO 1000-1.2 Cell line Cell line **Original PDX** 500. 100 80 **Days Post Implant** 0.9 -500-80 0 20 60 **Days Post Implant**

(C) ALDH1A1, (D) CXCR4, (E) ESA/EpCAM, (F) CD44, (G) MASPIN, (H) PDX1, (I) Ki67, and (J) negative control.

Noorani et al., Organoids. 2022; 1(2):168-183.

Cancer Stem Cell expression altered in long-term cultured 2D cell lines







Long-term 2D culture expression of CD44/CD24/ESA is cell line dependent



Stem cell plasticity is reconciled by culture conditions



CSC expression altered between tumour and normal



CD44 associated with overall survival



Modelling drug resistance in vitro:

 Poor long-term survival rates of pancreatic cancer are the consequences of rapidly acquired chemoresistance and represent a major therapeutic challenge

 Studying the emergence of resistance to therapeutics would allow us to identify key markers to guide treatment strategies



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Long-term drug selection derived organoids

Advanced in vitro organoid models



Establishment of 5-FU resistant PDAC organoids



Transcriptomic identification of differentially regulated genes involved in 5-FU resistance

59

PDM41-5FUR

195

PDM41-5FUR

136







Conclusions

- Organoids can be used as an emerging technology to advance of personalized medicine
- Model tumorigenesis, and recapitulate critical features of original cancer tissue
- CLOs are flexible, expandable, traceable models used for high-throughput screening of sensitive drugs to provide individualized treatment options
- Tools for understanding the mechanisms of drug resistance
 - Identify markers of resistance
 - Develop novel therapeutics to overcome drug resistance





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