

### Bacteriology Fundamentals

ATCC's best practices for optimal growth and propagation of bacteria and bacteriophages

Jeanette Rimbey, MSc Lead Biologist, Bacteriology ATCC

### Credible Leads to Incredible®



### Speaker information



### Jeanette Rimbey, MSc Lead Biologist, Bacteriology ATCC

Jeanette Rimbey is a team lead within ATCC with 15 years' experience as a microbiology scientist in the fields of bacteriology, genetics, veterinary, and virology. Ms. Rimbey achieved a Master of Science from Texas Tech University and began her scientific journey at the University of Missouri, where she explored bacteriophage applications to develop innovative methods for combating pathogens that threaten global food security. On a related note, her mentor was awarded the 2018 Nobel Prize in Chemistry for his work in developing phage display technology. She has contributed significantly to microbiology research, including work on temperature-specific biofilm adaptations in antibiotic-resistant microorganisms. During the COVID-19 pandemic, she served on the COVID-19 diagnostic task force as a molecular biologist, supporting critical front-line efforts in Colorado. Her passion for science drives her commitment to advancing microbiology and giving back to the scientific community.



# About ATCC®

Founded in 1925, ATCC<sup>®</sup> is a non-profit organization with HQ in Manassas, VA, and an R&D and Services center in Gaithersburg, MD

 World's premier biological materials resource and standards development organization

- 5,000 cell lines
- 80,000 microorganisms
- Genomic & synthetic nucleic acids
- Media/reagents

ATCC<sup>®</sup> collaborates with and supports the scientific community with industry-standard biological products and innovative solutions

- Growing portfolio of products and services
- Sales and distribution in 150 countries, 20 international distributors
- Talented team of 600+ employees, over onethird with advanced degrees











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### Agenda



- 1 ATCC<sup>®</sup> Bioproduction
- 2 Microbial cultivation and propagation
- 3 Troubleshooting
- 4 Quality assurance
- 5 Depositing at ATCC<sup>®</sup>
- 6 Resources





# ATCC<sup>®</sup> Bioproduction



# Comprehensive microbial collection

- Comprehensive microbial collection with enhanced authentication
  - 70,000+ bacteria, fungi, viruses, and protozoa
  - 8,000+ microbial type strains
- Brand recognition
  - Organizations and regulatory agencies specify ATCC<sup>®</sup> cultures in their standards and guidelines
  - USP, ISO, FDA, CLSI, USDA, ASTM, AOAC, WHO
  - 475+ strains recommended for use in quality control
- ATCC<sup>®</sup> has live microbes and derivatives, including inactivated materials and nucleic acids
- Variety of advanced techniques used to characterize and authenticate biomaterials—no single method of identification is sufficient





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# Types of vials to store and ship microbes

Batch vials: lyophilized format with flame-sealed external glass shell (glass ampoule)





"Mini" vials: 0.2 mL of frozen material sold in packs of 6

Serum/preceptrol vials: lyophilized material sealed with rubber plug and metal crimp





**1.0 mL Frozen/cryovials:** 0.5 mL of frozen material



# How to open a glass ampoule

- 1. Disinfect the outside of the ampoule with freshly prepared 70% ethanol.
- 2. Wipe off the ampoule with a sterile towel or gauze to dry residual ethanol.
- 3. Score the neck of the ampoule with a vial scorer until a fracture line is made. Apply gentle pressure when rotating.
- 4. Hold the vial upright and lightly tap open the vial using a metal file along the fracture line.
- 5. Immediately, relocate vial to a Biosafety cabinet (BSC) prior to rehydrating the material.







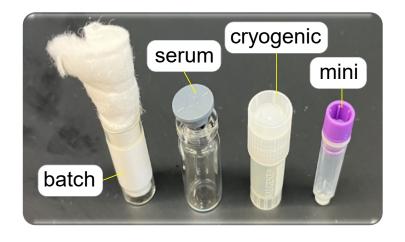
# Initiating frozen and lyophilized cultures

### **Rehydrating pellets**

- 1. Lyophilized vials should be stored at 4°C.
- 2. Open the vial according the enclosed instructions.
- 3. Working in a Biosafety Cabinet (BSC), use a single tube containing 5-6 mL of broth and rehydrate the entire pellet.
- 4. Aseptically transfer the aliquot back into the broth tube and mix well.
- 5. Use several drops of the suspension to inoculate the recommended agar plate.
- 6. Incubate at the suggested temperature and atmospheric conditions.

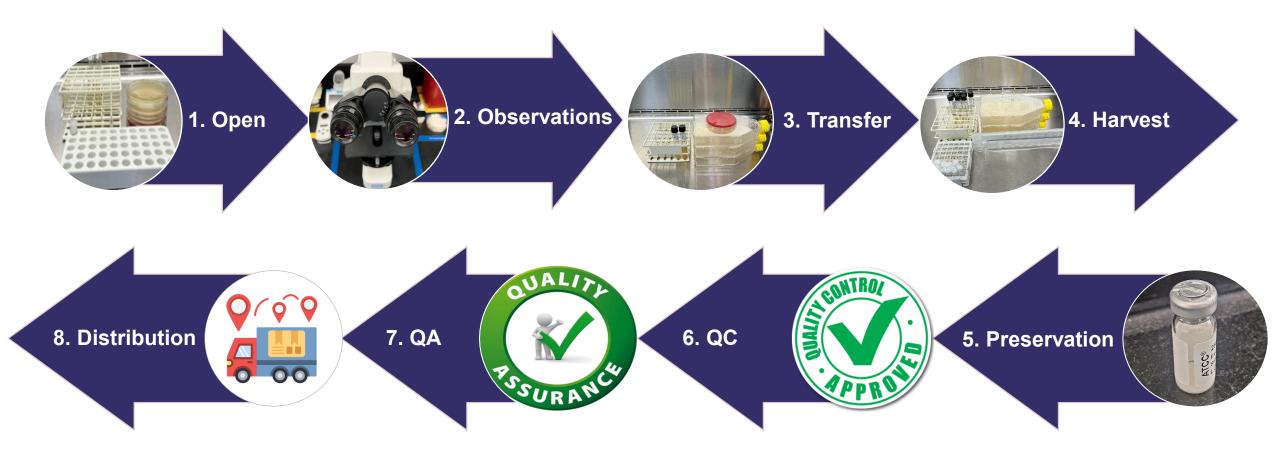
### **Thawing Frozen Vials**

- **1**. Frozen vials should be stored at -80°C.
- 2. Open the vial according the enclosed instructions.
- 3. Thaw vial in a 37°C water bath for ~3 mins.
- 4. Working in a BSC, unscrew the vial and transfer the contents to a sterile test tube containing the appropriate growth medium. Proceed as in previous steps 5 & 6.



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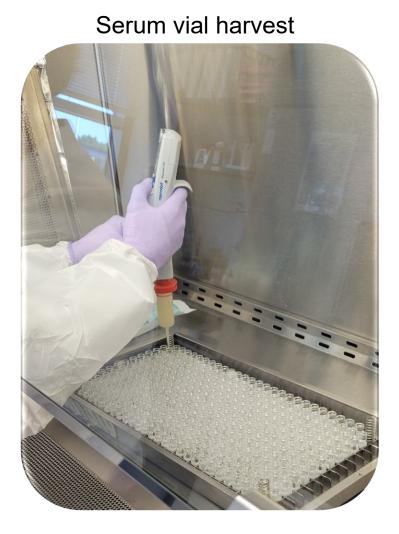
### Bioproduction process of BSL-1 and BSL-2 microbes





# Vialing on harvest day

Batch vial harvest



Cryovial harvest







# Microbial cultivation and propagation



# Microbe-specific atmospheric conditions

### Aerobic

- Pathogenic or commensal bacterial strains grow well at body temperature of 37°C
- Environmental strains thrive at lower temperatures, often 25°C to 30°C

### Microaerophilic

- May require 5% CO<sub>2</sub> incubation
- Most rely on 3-5% O<sub>2</sub> and 10% CO<sub>2</sub>

### Anaerobic

Establishing a great anaerobic cultivation practice is key to maintain good growth.

### Anaerobic chamber



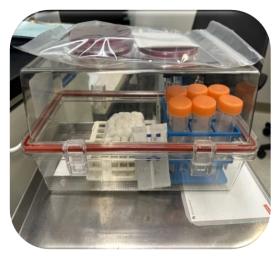
Needle & gas exchange





Anaerobic jars

Anaerobic box with gas generating sachets



**ATCC**<sup>°</sup>

# Aerobic bacteria and its many applications

# Escherichia coli, Salmonella enterica, and Listeria monocytogenes

- Reference strains needed for the testing of food production.
- Wide assortment of food testing solutions support QC testing, process validation, and development of new methods for testing food products.

### Staphylococcus aureus

 Antimicrobial resistant strains for research and drug development for clinical medicine.

### Pseudomonas aeruginosa

 Specific cystic fibrosis strains from phase III clinical trial studies that provide a unique model system for better understanding of the molecular pathology associated with cystic fibrosis.





# Common aerobic media

General purpose, non-selective, and nutrient-rich

### General purpose

- Nutrient Agar/Broth
- Tryptic Soy Agar/Broth (TSA/TSB)
- Luria-Bertani broth and agar (LB)



### **Nutrient-Rich Media**

- Brain-Heart Infusion (BHI)
- Chocolate Agar Base (GC medium)
- TSA w/ 5% Sheep's Blood (Blood agar)



# Specialized group-specific media

- Charcoal Yeast Extract
  - Legionella and related genera
- Marine Media
  - Vibrio and related marine genera
- Group-specific supplemented media
  - Haemophilus Test Media
  - Azotobacter Supplement Media
  - Lactobacilli MRS Media
- Bordet-Gengou used to isolate Bordetella pertussis
- Additional supplements
  - Fetal Bovine Serum (FBS)
    - Mycoplasma growth
  - Horse Serum
    - o Ureaplasma growth

### Lactobacilli MRS Media





- Facultative anaerobes
  - Grows with or without the presence of oxygen.
- Aerotolerant anaerobes
  - Uninhibited by oxygen but can generate energy without using oxygen via fermentation.
- Strict anaerobes (obligate)
  - Only grows in the absence of oxygen. Oxygen is toxic to this organisms. Picture to the right is an example of a strict anaerobe.
- Methanogens
  - Class of strict anaerobes that require nitrogen or ammonia sources in their media.
  - Methane produced as a byproduct of their metabolism.

### Coprococcus catus (ATCC<sup>®</sup> 27761<sup>™</sup>)





Nutritional requirements is key to establishing healthy growth

- The superior choice is using Pre-Reduced Anaerobically Sterilized (PRAS) media.
- At ATCC<sup>®</sup>, we recommend using pre-reduced media that was either freshly prepared or previously prepared and stored under anaerobic conditions.
- Common broth medias are chopped meat, reinforced clostridial, and peptone yeast extract broth with glucose (PYG).
- Media should be prepared with reducing agents and stored in anaerobic environments.
- Resazurin, an indicator compound, is added to media that has been reduced of oxygen.

### Bottled broth + TSA blood plates



Broth media: chopped meat



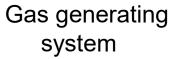
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# Reducing anaerobic media

- Standard anaerobic gas mixture is 80% N<sub>2</sub>, 10% CO<sub>2</sub>, and 10% H<sub>2</sub> for most anaerobes.
- Media should be prepped with reducing agents and stored in an anaerobic environment.
- Common reducing agents are 3% cysteine HCI, 5% coenzyme M, or 15% sodium sulfide.

Gas exchange through a needle and syringe system

Pre-reduce media bottles



Anaerobic chamber











### Learn more about anaerobes



Overcoming the Challenges of Growing Anaerobic Bacteria

June 12, 2024 Jeanette Rimbey, MS

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Explore the best approaches for achieving successful growth of anaerobic bacteria.

Read our blog post to learn more about our anaerobic laboratory practices

- Get more details on anaerobes
- Learn how to select the right media
- Find tips on achieving the best growth conditions



Scan to get the blog

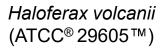
# Extremophiles

### Temperature and pH

- Thermophile 45°C and up
- Psychrophile 15°C and below
- Halophile prefer high salt concentration
- Acidophile pH 3.0 or below
- Alkaliphile pH 9.0 or above

### **Unique growth requirements**

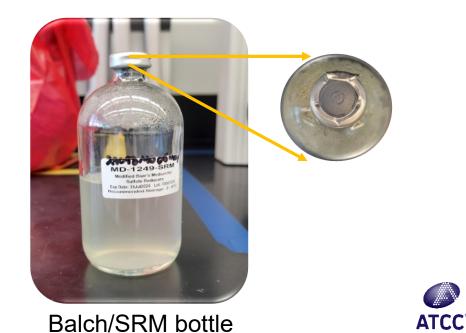
- Environmental anaerobes that grow in niche environments
  Acidic, excessively hot, or irregular in air pressure
- Growth may require Balch tubes/SRM: crimped aluminum seal for culturing
  - Maintains additional pressure
- May also require gas mixtures free of carbon dioxide (CO<sub>2</sub>)







Yellowstone National Park



# Microaerophiles

### Bacteria requiring lower levels of oxygen vs those in normal air

- Examples:
  - Helicobacter
  - Neisseria
  - Campylobacter
- Requires nutrient rich media blood based media or serum
- Beneficial to use a reliable gas generating system
- Atmospheric conditions is between 5-10% O<sub>2</sub> and 8-10% CO<sub>2</sub>
- May grow best in a biphasic environment
- Growth tip
  - Make sure media is pre warmed before inoculation to avoid growth issues
- Fun history fact
  - Microaerophiles were historically cultivated in candle jars!

### **Biphasic growth**





# Bacteriophages - "Bacteria eater"

- ATCC uses Broth propagation methods routinely, but Adams Agar Overlay Method is acceptable as well.
- Propagation Steps:

Day 1	Day 2	Day 3	Day 4		
Propagate host following basic cultivation practices	Amplification of host + bacteriophage using the respective bacterial host strain. Host should be in early log phase prior to adding lysate.	Filter the culture to rid of bacteria cells and perform a spot titer to determine the plaque forming units (pfu).	After 24-hour incubation, lysis should be visible. At the higher dilutions, individual plaques should be countable.		

- Method suggestions to consider
  - Make sure host is at an optimal Optical Density (0.1-0.3) prior to introducing the lysate.
  - Consider the appropriate ratio of virus-to-cells called multiplicity of infection (MOI) to enhance lysate growth.

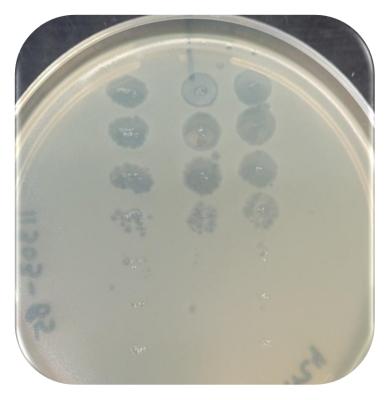
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 Try adding MgCl + CaCl (2<sup>+</sup> salt) to media which increases the potential for phage aggregation through cation binding.

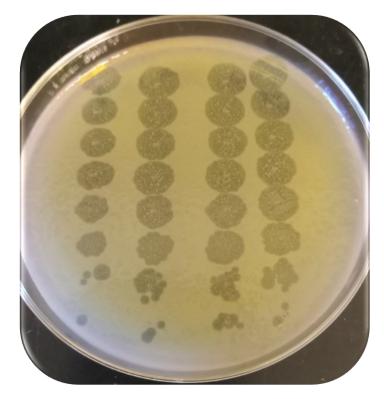
# Plaques

Zones of clearing are observed within the lawn of bacterial growth on a plate.

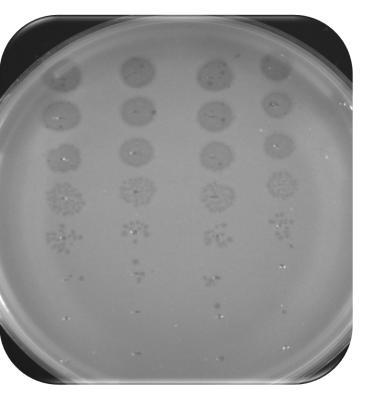
Escherichia coli bacteriophage T5 (ATCC<sup>®</sup> 11303-B5™)



Pseudoalteromonas espejiana bacteriophage PM2 (ATCC<sup>®</sup> 27025-B1<sup>™</sup>)



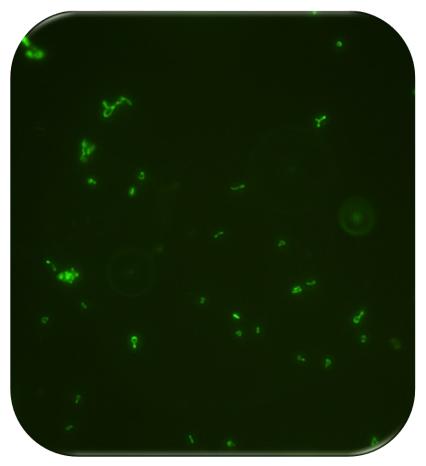
Escherichia coli bacteriophage Mu-1 (ATCC<sup>®</sup> 23724-B9™)





- Very small, pleomorphic cells without a cell wall
- Genera include:
  - Mycoplasma, Ureaplasma, Spiroplasma, and Acholeplasma
- Most are aerobic or facultative anaerobes
- Shape depends on osmotic pressure, nutritional quality of the culture medium, and growth phase
- Most media includes pH indicator to track growth cycle
  Unfavorable pH will inhibit growth
- If the organism is struggling to grow, try adding 10% fetal bovine serum as a supplement
- It's recommended to use at least a 10% inoculum when scaling up growth with mollicutes

### Mycoplasma ovipneumoniae (ATCC<sup>®</sup> 29419™)







# Troubleshooting



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# Bacteriology troubleshooting

### **Growth optimization**

- Growth optimization assays are performed to test different media with kinetic experiments.
- Comparing preservation methodologies to determine best vial type for numerous organisms.

### Specialty troubleshooting

- Antibiotic resistance testing
- Construct PCR protocols to test for the presence/absence of various genes of interest
- Whole-genome sequencing is conducted if suspicious of hidden contaminants

### **New accessions**

Our team determines the ideal growth conditions for new deposits based on depositor notes and research.







# Quality Assurance

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# Advancing authentication through credible standards

Quality control

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MALDI-TOF MS

- Phenotypic analysis Cell morphology, colony description, viability, and purity.
- Genotypic analysis Sequencing conserved regions of the genome and whole-genome sequencing (ATCC<sup>®</sup> Genome Portal).
- Proteomic analysis MALDI-TOF MS
- Functional testing Antimicrobial susceptibility testing, serotyping, virulence detection

ATCC uses a variety of advanced techniques to characterize and authenticate biomaterials—no single method of identification is sufficient. Researchers look to ATCC for a wide range of authentication resources to safeguard reproducibility and meet requirements for funding, publication, and quality control.



# ATCC<sup>®</sup> Genome Portal

A cloud-based platform that enables users to easily browse authenticated and traceable reference genomes and metadata.



Download whole-genome sequences and annotations from your browser or via our secure API.



Search for nucleotide sequences or genes within genomes.



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View genome assembly metadata and quality metrics.

### genomes.atcc.org

### 5,000 Authenticated Microbial Reference Genomes

4,205 bacteria and archaea 439 viruses 352 fungi 4 protists New

New genomes released every quarter!

**REST-API** for bioinformatics applications available



# Depositing at ATCC<sup>®</sup>



### How to deposit at ATCC<sup>®</sup>

- 1. Visit <u>www.atcc.org</u>
- 2. Select "Services"

- 3. Select "Depositing with ATCC"
- 4. Choose the type of deposit
- 5. Fill out the electronic deposit request form provided
- 6. Submission is sent to our Content & Accessioning team for review.
- 7. A project manager will reach out with further details once a deposit is approved.



# Deposit service options

### General Collection Deposit



Preserve your research biomaterials and make them available to researchers worldwide by depositing them into the ATCC General Collection.

### Patent Deposit



ATCC is an International Depositary Authority under the Budapest Treaty and also accepts deposits under the rules of the US Patent and Trademark Office.

### National Park Service Deposit



Deposit biological specimens collected from a US national park into the National Park Service Special Collection.

### Type strain deposit



ATCC conducts viability and identity testing, and then, if applicable, provides depositors with a Certificate of Deposit required for publication.

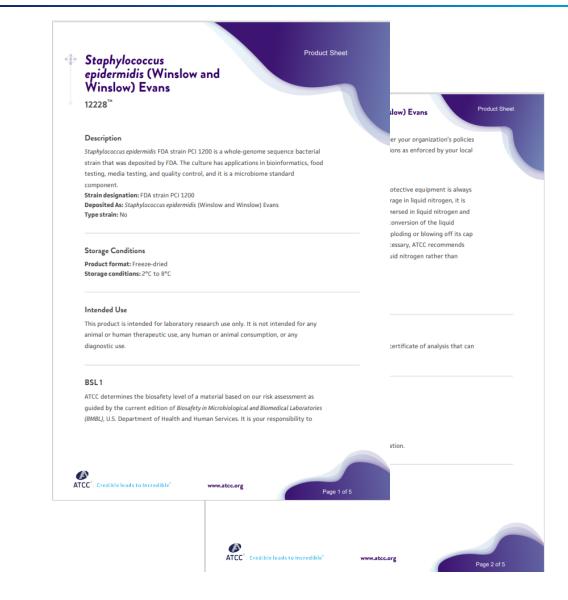


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### Resources



### Access detailed product sheets



- Propagation instructions
- Maintenance media
- Atmospheric needs
- Growth temperature
- Incubation time
- Helpful handling notes
- Growth optimization assay results

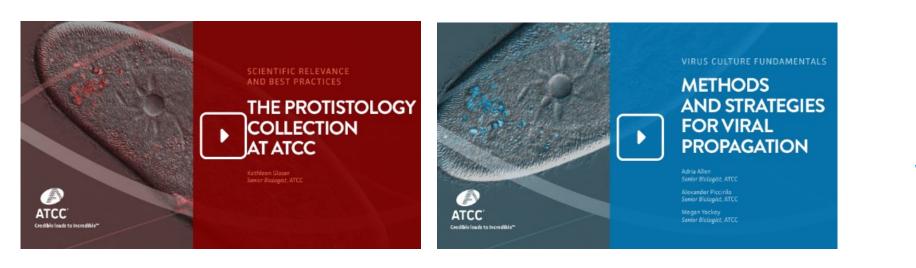


### Get expert tips in our culture guides





### View our webinars



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### Read our blog posts



The science and innovation blog from inside ATCC, the world's premier biomaterial organization

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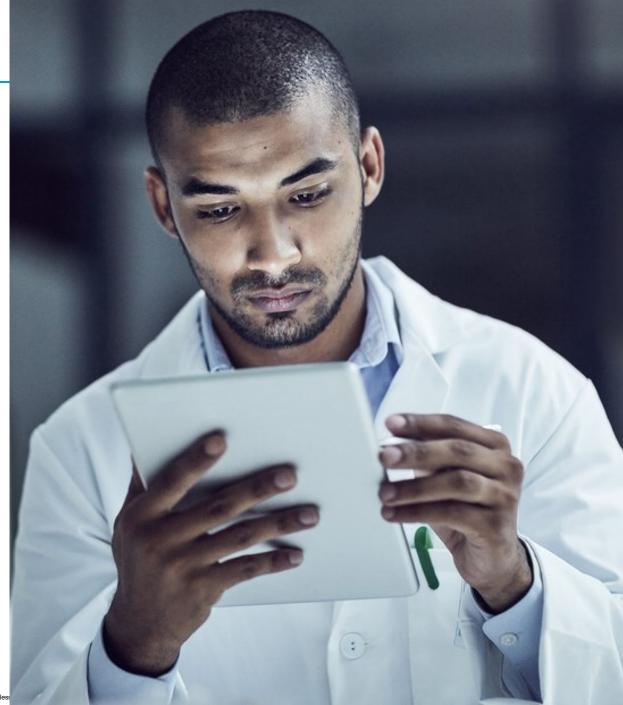




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# Conclusions

### Things to consider...



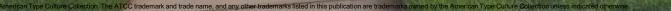
- Follow a standard propagation method. ATCC<sup>®</sup> product sheets provide helpful handling information.
- Use the appropriate medium, growth conditions, and length of incubation.
- Reminder that certain organisms prefer shaking incubation over traditional static incubation.
- Make sure your media is room temperature before inoculation.
- Commercial vs. self-made media
  - -Commercially made media will often help with poor growth.
  - -Don't forget to <u>reduce</u> anaerobic media prior to inoculation.
- Consider specific techniques necessary for specialized strains.
- Bacteriophages If you don't see plaques, confirm that the host is viable and check the NaCl concentration of the media.

# Acknowledgments

Thank You!



- Graham Hogg
- Nancy Krueger
- Joe Thiriot
- ATCC<sup>®</sup> Bacteriology Team
- Microbiology Bioproduction Teams: Bacteriology | Mycology | Protistology | Molecular Genomics | MSAT-BacT



# Learn more about Bacteriology from ATCC

# Submit your questions for the ATCC team

- Jeanette Rimbey, Lead Biologist Bacteriology (presenter)
- Graham Hogg, Biologist
- Joe Thiriot, Lab Operations
- Genome Portal Team

# View all culture guides at www.atcc.org/guides



