

Modeling toxicity with Neural Progenitor Cellderived Neurospheres

Brian Shapiro, PhD Scientific Content Specialist, ATCC





About ATCC

- Founded in 1925, ATCC is a non-profit organization with HQ in Manassas, VA, and an R&D and Services center in Gaithersburg, MD
- World's largest, most diverse biological materials and information resource for cell culture the "gold standard"
- Innovative R&D company featuring gene editing, differentiated stem cells, advanced models
- cGMP biorepository

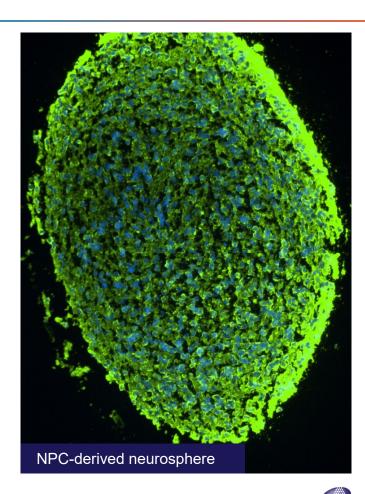
- Partner with government, industry, and academia
- Leading global supplier of authenticated cell lines, viral and microbial standards
- Sales and distribution in 150 countries, 19 international distributors
- Talented team of 450+ employees, over onethird with advanced degrees



Agenda

Neural Progenitor Cells (NPCs)

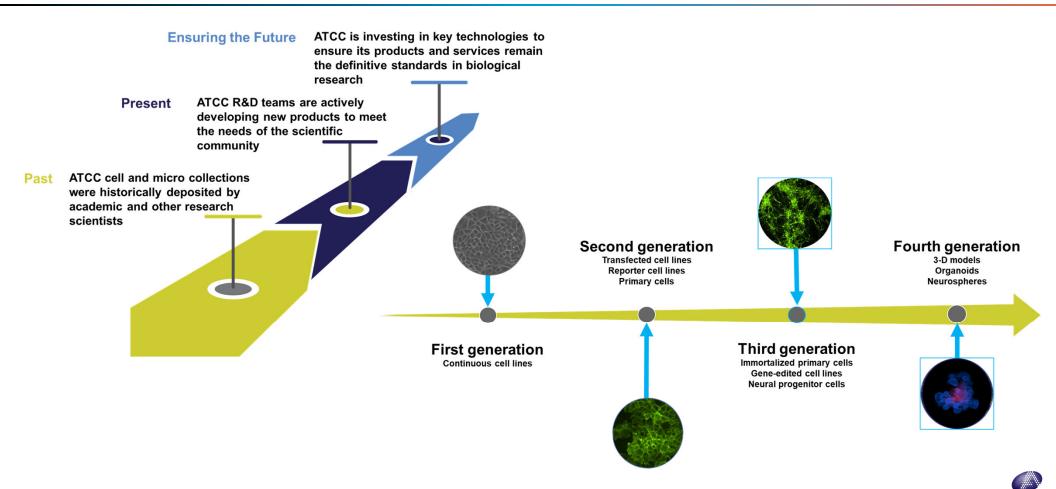
- ATCC roadmap and neurological resources
- Background information
- Differentiation potential of ATCC NPCs
- Neurosphere generation
- Toxicological studies using neurospheres
- Summary



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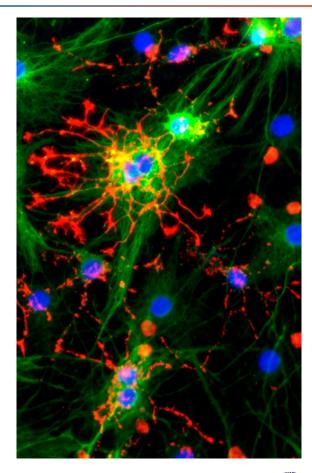
Modernization of the ATCC neurological portfolio: Evolution of in vitro cell models



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ATCC products for neurology

- ATCC is the complete solution supplier for neuroscientists
- From basic research through discovery and development to product testing
 - Continuous cell lines
 - hTERT-immortalized primary NTAP Schwann cells
 - Michael J. Fox Foundation cell lines
 - Neural progenitor cells
 - Human Cancer Model Initiative 2-D and 3-D products
- Portfolio features
 - Reliability
 - Fully characterized cells
 - Optimized growth protocols
 - Scalability into all aspects of the neuroscience workflow
 - Biological relevancy

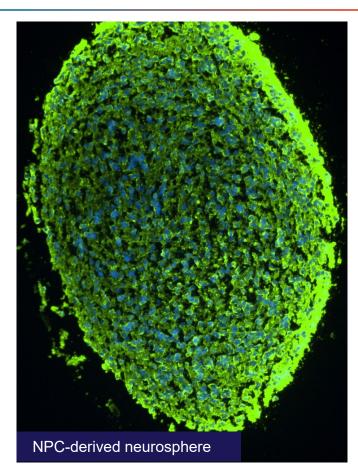




Agenda

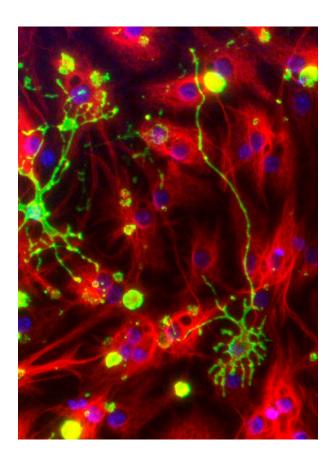
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Challenges associated with current models of the nervous system

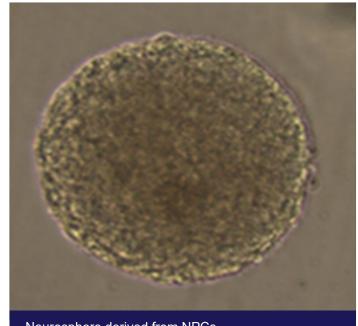


- Primary cells from animals (mouse and rat neurons)
 - -Not predictive
 - Donor variation
- Continuous cell lines (originally isolated from tumors)
 - -Not normal
 - -Not predictive
- Induced pluripotent stem cells (iPSCs; commercial or selfmade)
 - Time and labor intensive
 - -Often not validated for neural development
- 2D vs. 3D models



Neurosphere characteristics

- A neural model system
 - A 3-D model
 - Free-floating spherical clusters of neural stem cells
 - Can be used in undifferentiated state
 - Can be differentiated into multiple subtypes
 - At different timepoints the neurospheres can represent different neural developmental stages
 - Multiple methods of generation are available
- Neurosphere features
 - Expression levels of markers:
 - Nestin positive, neurofilament and GFAP negative
 - o Culture environment-dependent after differentiation
 - Size <50 to >600 µm
 - Amenable to growth in 96-well formats/high-throughput screening
 - Assays: apoptosis, neurite outgrowth, proliferation, ICC, etc...



Neurosphere derived from NPCs



Neurospheres in the literature

Published in final edited form as: *Cell Stem Cell*. 2017 April 06; 20(4): 435–449.e4. doi:10.1016/i.stem.2016.12.007.

Human iPSC-derived cerebral organoids model cellular features of lissencephaly and reveal prolonged mitosis of outer radial glia

Marina Bershteyn^{1,2,6,*}, Tomasz J. Nowakowski^{1,3}, Alex A. Pollen^{1,3}, Elizabeth Di Lullo^{1,3}, Aishwarya Nene⁴, Anthony Wynshaw-Boris^{2,5,*}, and Arnold R. Kriegstein^{1,3,7,*}

¹Eli and Edythe Broad Center of Regeneration Medicine and Stem Cell Research Put California, San Francisco, CA, USA

Published in final edited form as: Toxicol Appl Pharmacol. 2018 September 01; 354: 101–114. doi:10.1016/j.taap.2018.02.003.

Rotenone exerts developmental neurotoxicity in a human brain

spheroid model

David Pamies^a, Katharina Block^a, Pierre Lau^b, Laura Gribaldo^b, Carlos A. Pardo^c, Paula Barreras^c, Lena Smirnova^a, Daphne Wiersma^a, Liang Zhao^{a,d}, Georgina Harris^a, Thomas Hartung^{a,e}, and Helena T. Hogberg^{a,*}

Published in final edited form as: Cell Stem Cell. 2016 August 4; 19(2): 258–265. doi:10.1016/j.stem.2016.04.014. ^aCenter for Alternative to Animal Testing (CAAT), Johns Hopkins University, 615 North Wolfe Street, Baltimore, MD 21205, USA

Zika Virus Depletes Neural Progenitors in Human Cerebral Organoids through Activation of the Innate Immune Receptor TLR3

Jason Dang^{1,3}, Shashi Kant Tiwari^{1,3}, Gianluigi Lichinchi¹, Yue Qin¹, Veena S. Patil¹, Alexey M. Eroshkin², and Tariq M. Rana^{1,*}

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¹Department of Pediatrics and Institute for Genomic Medicine, University of California San Diego School of Medicine, 9500 Gilman Drive MC 0762, La Jolla, California, 92093, USA

Microglia Increase Inflammatory Responses in iPSC-Derived Human BrainSpheres

Celina Monteiro Abreu¹, Lucio Gama^{1,2}, Susanne Krasemann³, Megan Chesnut⁴, Shelly Odwin-Dacosta⁴, Helena T. Hogberg⁴, Thomas Hartung^{4,5} and David Pamies⁴⁺

¹ Department of Molecular and Comparative Pathobiology, Johns Hopkins School of Medicine, Baltimore, MD, United States, ³ Vaccine Research Center, National Institute of Allergy and Infectious Diseases, NIH, Bethesda, MD, United States, ³ Institute of Neuropathology, University Medical Center Hamburg-Eppendorf, Hamburg, Germany, ⁴ Center for Alternatives to Animal Testing, Johns Hopkins Bloomberg School of Public Health, Baltimore, MD, United States, ⁵ CAAT-Europa, University of Konstanz, Konstanz, Germany



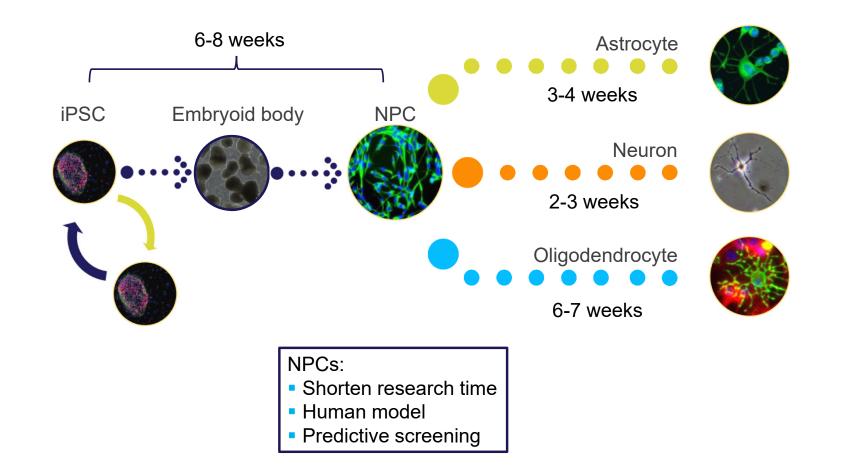
Published in final edited form as: *Nature*. 2014 November 13; 515(7526): 274–278. doi:10.1038/nature13800.

A three-dimensional human neural cell culture model of Alzheimer's disease

Se Hoon Choi^{1,†}, Young Hye Kim^{1,2,†}, Matthias Hebisch^{1,3}, Christopher Sliwinski¹, Seungkyu Lee⁴, Carla D'Avanzo¹, Jennifer Chen¹, Basavaraj Hooli¹, Caroline Asselin¹, Julien Muffat⁵, Justin B. Klee¹, Can Zhang¹, Brian J. Wainger⁴, Michael Peitz³, Dora M. Kovacs¹, Clifford J. Woolf⁴, Steven L. Wagner⁶, Rudolph E. Tanzi^{1,*}, and Doo Yeon Kim^{1,*} ¹Genetics and Aging Research Unit, MassGeneral Institute for Neurodegenerative Disease, Massachusetts General Hospital, Harvard Medical School, Charlestown, MA 02129, USA

> ORIGINAL RESEARCH published: 04 December 2018 doi: 10.3389/fmicb.2018.02766

Neural progenitor cells – Neuronal differentiation



ATCC[°]

ATCC NPC offerings

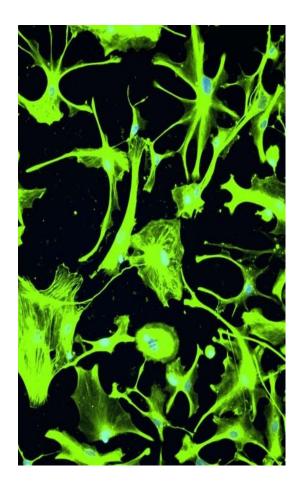
ATCC [®] No.	Designation
ACS-3003 [™]	NPC Growth Kit – add to DMEM/F12
ACS-3004 [™]	NPC Dopaminergic Differentiation Kit – add to DMEM/F12
ACS-5001™	NPCs derived from ATCC-DYS0530 Parkinson's Disease (ACS-1013) New!
ACS-5003™	NPCs derived from ATCC-BXS0117 (ACS-1031)
ACS-5004™	NPCs derived from ATCC-BYS0112 (ACS-1026)
ACS-5005 [™]	Neural Progenitor Cells derived from XCL-1 DCX-GFP (for late neuron differentiation)
ACS-5006 [™]	Neural Progenitor Cells derived from XCL-1 GFAP-Nanoluc®-Halotag® (for astrocyte differentiation)
ACS-5007 [™]	Neural Progenitor Cells derived from XCL-1 MAP2-Nanoluc®-Halotag® (for early neuron differentiation)
ACS-2103F [™]	Screening Fee – For Profit

ATCC[®] ACS-1026[™] – iPSC derived from bone marrow CD34+ cell from Caucasian male ATCC[®] ACS-1031[™] – iPSC derived from bone marrow CD34+ cell from Asian female

Reporter lines from iPSC derived from cord blood CD34+ from a Caucasian male (XL-1 iPSCs from NIH)



QC testing of NPCs



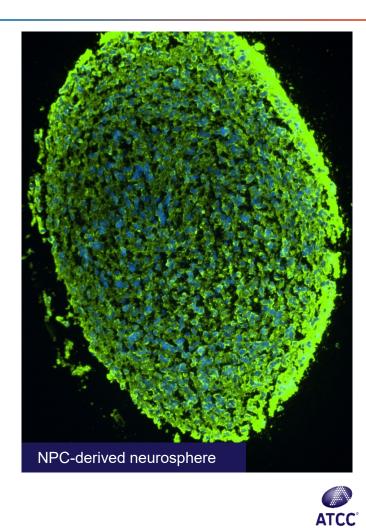
- Post-thaw cell viability: >80%
- Post-thaw viable cell number: >1x10⁶ cells/vial
- Longevity: >15 PDLs or 5 passages
- NPC marker expression: Nestin⁺, Pax-6⁺, and Tra-I-60⁻
- Differentiation potential:
 - -Tuj1⁺ early neurons
 - -TH⁺ dopaminergic neurons
- Identity: STR profile matching parental iPSC line
- Sterility, mycoplasma, and viral panel testing: None detected



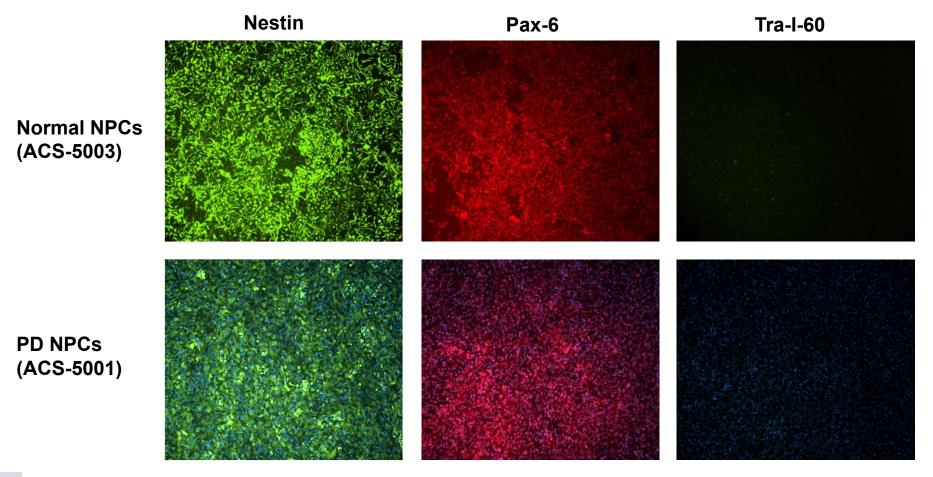
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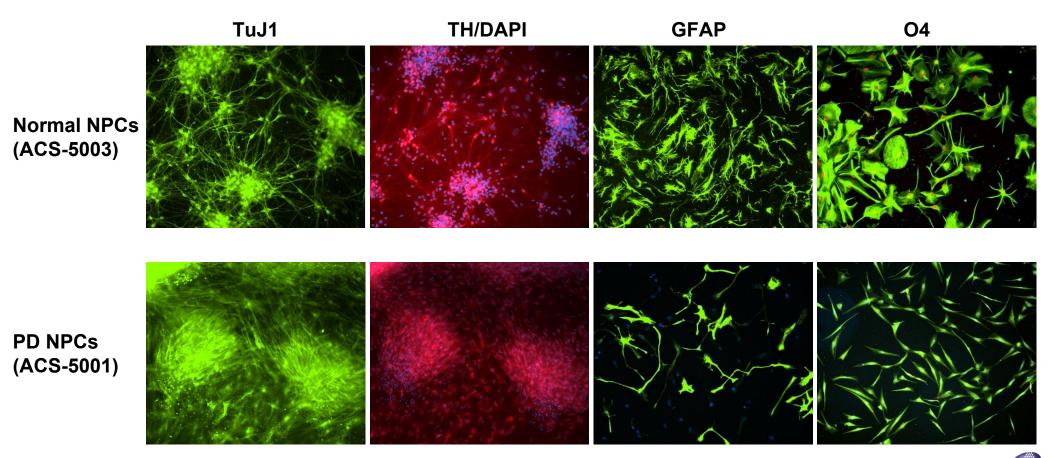


ATCC NPCs express NPC markers but <u>not</u> iPSC markers

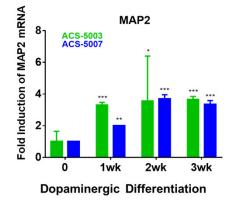


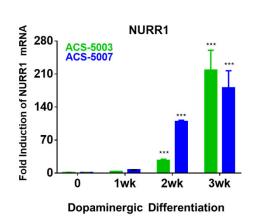


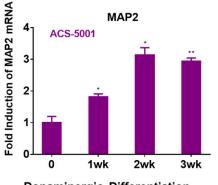
Early and dopaminergic neuron, astrocyte, and oligodendrocyte differentiation of normal and PD NPCs



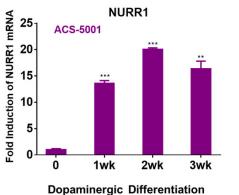
Expression of dopaminergic neuron genes

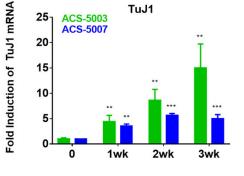




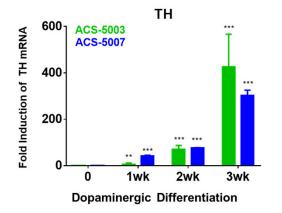


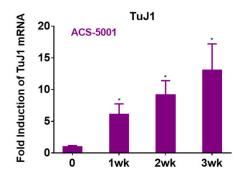
Dopaminergic Differentiation



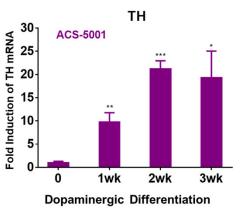


Dopaminergic Differentiation



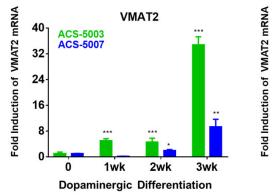


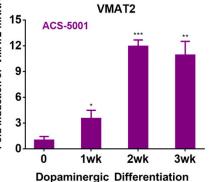
Dopaminergic Differentiation

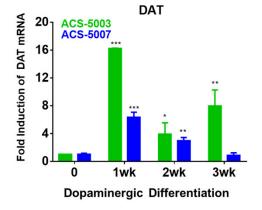


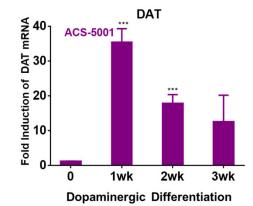
ATCC

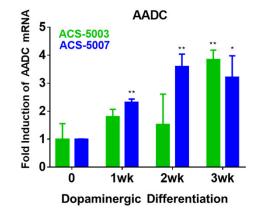
Expression of VMAT2, DAT, and AADC

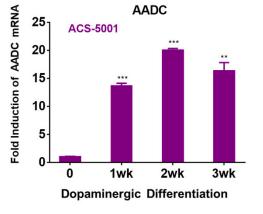










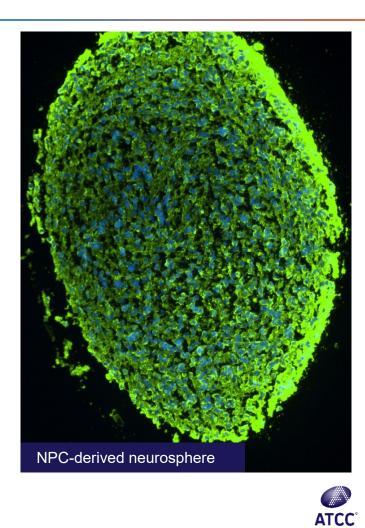




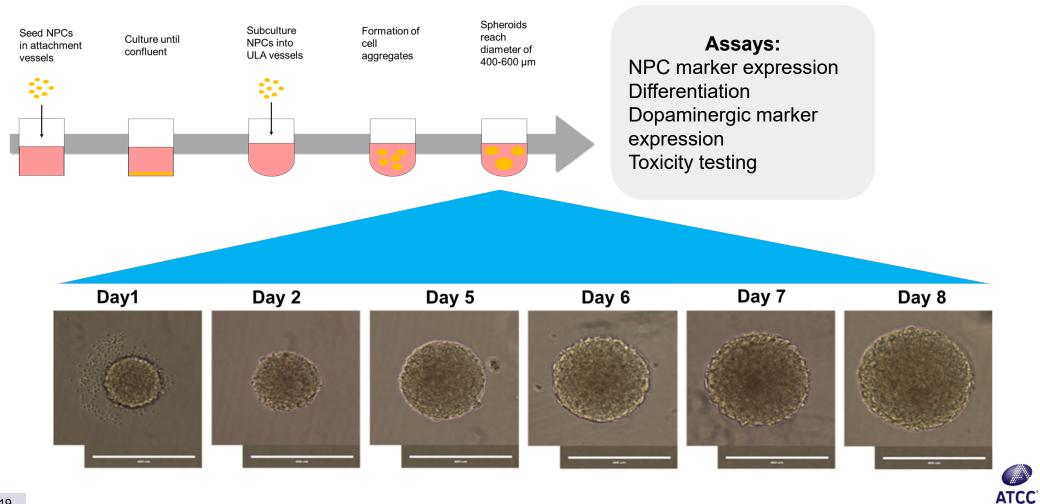
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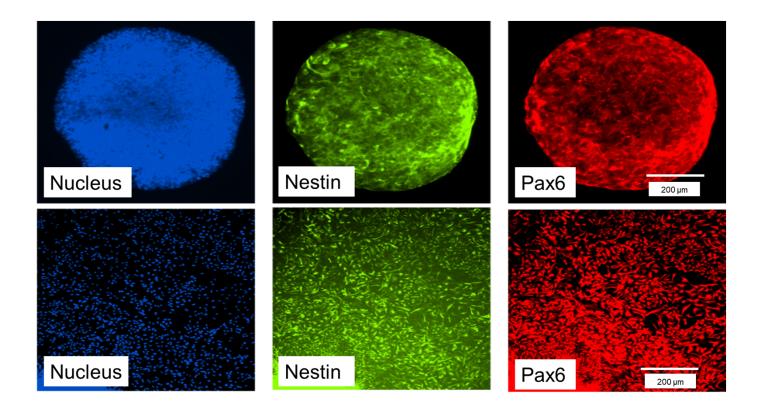
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NPC neurospheres grow in size over time

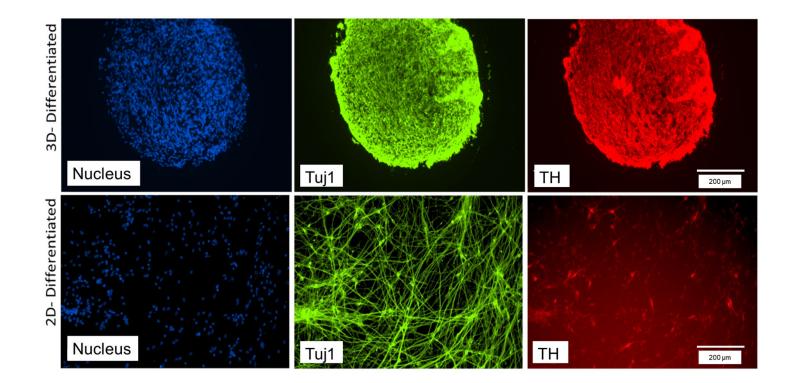


NPC derived-neurospheres and neurosphere-derived NPCs express NPC markers



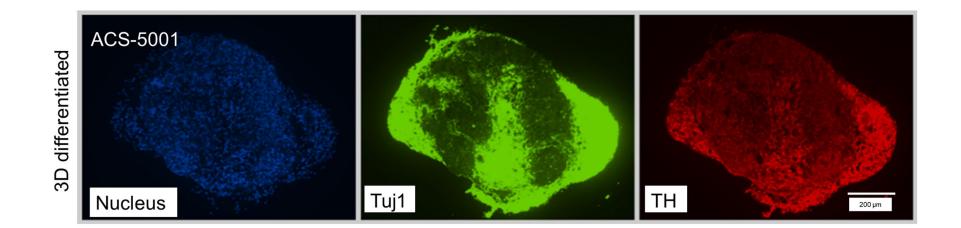


Normal neurospheres successfully differentiate and show higher TH positivity compared to 2D NPC cultures





Parkinson's disease donor derived neurospheres displayed different patterning after dopaminergic differentiation

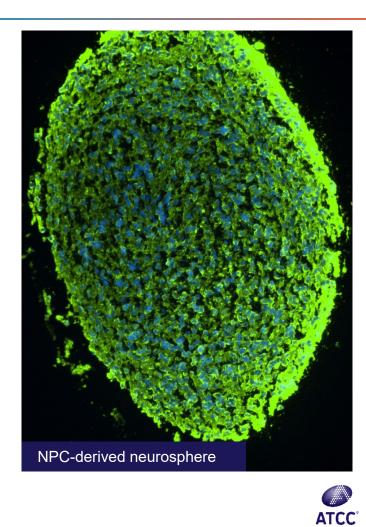




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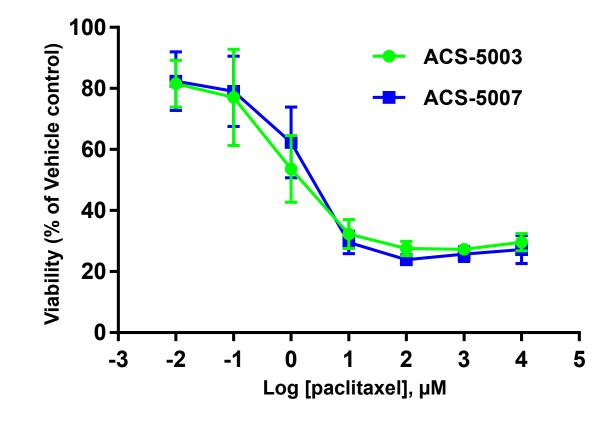


Compounds used in toxicity studies

Compound	Known effects of the Compound			
Paclitaxel	Microtubule stabilizer; known to cause neuropathies			
Cisplatin	Platinum-based apoptotic agent; known to have neuropathic affects but reputed to not be neurotoxic			
Piperine	Nociceptive agent; not known to cause neurotoxiciy			
Vincristine	Plant alkaloid; shown to cause peripheral neuropathy when used to treat pediatric A.L.L.			
Hydroxyurea	Antimetabolite; may cause severe peripheral neuropathy			
Amiodarone	Associated with peripheral neuropathies			
Chlorhexidine	Neurotoxic to neurons, SH-SY5Y cells, and Schwann cells			

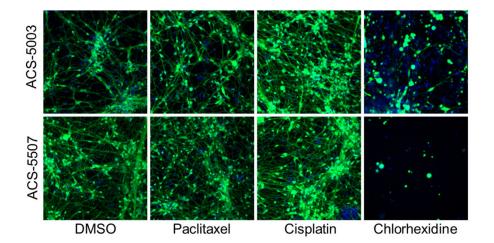


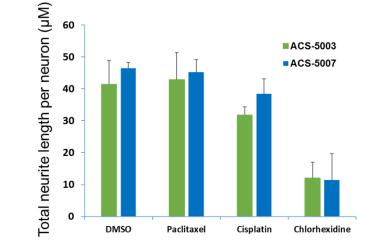
Dose-response curves for cell viability of normal, undifferentiated NPCs treated with paclitaxel for two days





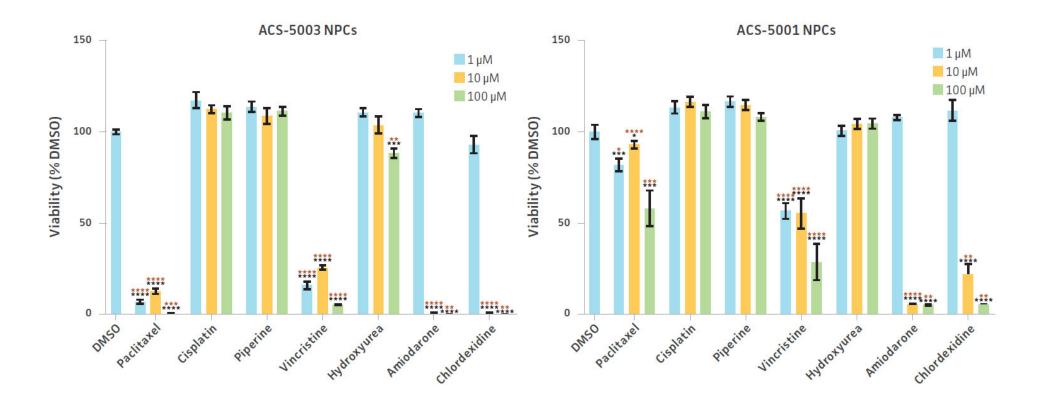
High content imaging of neurotoxicity in differentiated NPCs





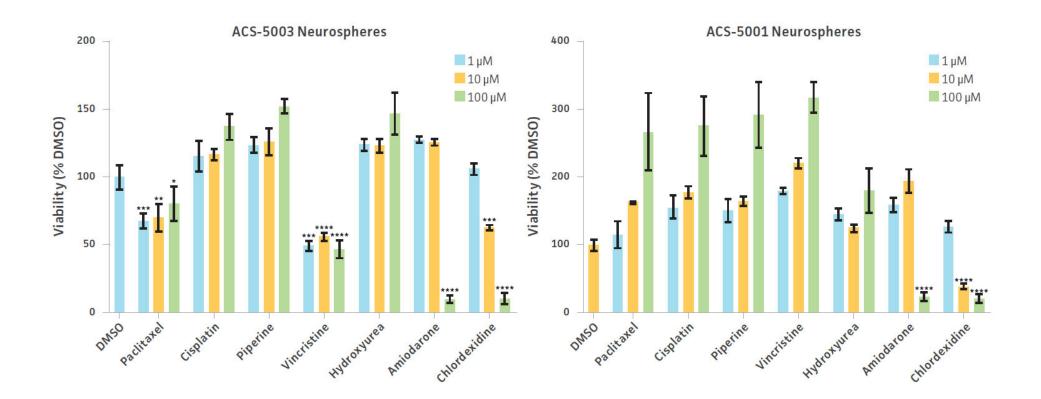


Normal and PD NPCs showed differential response to various drug treatments



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Normal and PD neurospheres showed differential response to various drug treatments; these responses varied with their 2D NPC counterparts





Overall neurotoxicity studies, NPCs vs. NPC-derived neurospheres

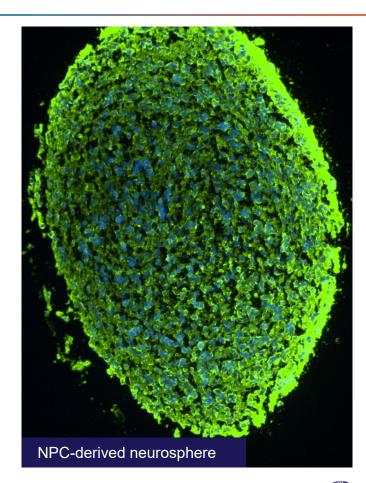
Toxin	ACS-5003 NPCs	ACS-5001 NPCs	ACS-5003 Neurospheres	ACS-5001 Neurospheres
Paclitaxel	Toxic	Weakly toxic	Weakly toxic	Resistant
Cisplatin	Resistant	Resistant	Resistant	Resistant
Piperine	Resistant	Resistant	Resistant	Resistant
Vincristine	Toxic	Toxic	Toxic	Resistant
Hydroxyurea	Weakly toxic	Resistant	Resistant	Resistant
Amiodarone	Toxic	Toxic	Weakly toxic	Weakly toxic
Chlorhexidine	Toxic	Toxic	Toxic	Toxic



Agenda

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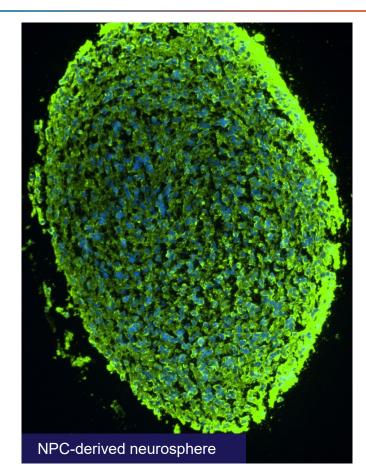
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Summary

- A complete solution:
 - Normal, PD, or reporter labeled NPCs
 - Expansion and Differentiation Medium
 - Easy-to-use protocols
- Human model with no donor variation
 - Ability to expand and bank
- Differentiation across a wide spectrum of neural and glial lineages
 - Various types of neurons
 - Astrocytes
 - Oligodendrocytes
- Live imaging of differentiation
 - GFP expression upon neural differentiation
- Advanced models of neurotoxicity that satisfy requirements for:
 - High cell yield
 - High physiological relevance

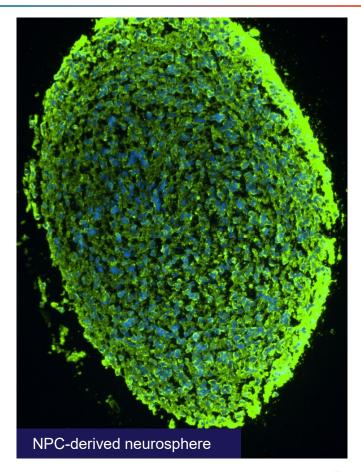




Summary – continued

- ATCC NPCs can convert to neurospheres with 100% efficiency
 - Ultra-low attachment culture vessels and NPC expansion medium
- NPC-derived neurospheres were similar to literature results
 - Formed solid circular spheres without formation of hollow cavities
 - Maintained their non-differentiated state for more than 2 weeks
 - Differentiated into dopaminergic neurons
 - More TH positivity than differentiated 2-D monolayer culture
- 3-D neurospheres are less sensitive to chemotherapeutics compared to 2-D NPC cultures
- Normal neurospheres were sensitive to vincristine but Parkinson's neurospheres were not
- NPCs cultured as 3-D neurospheres are amenable to drug toxicity studies using standard cell viability assays

For more information visit www.atcc.org/neuro





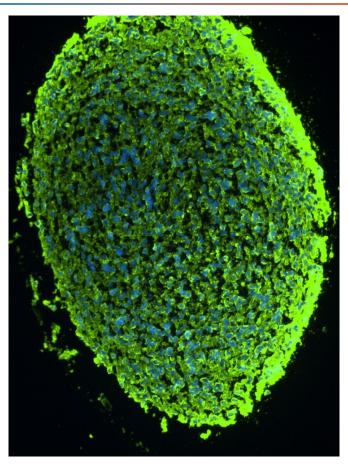
Thank you and questions?

Coming soon!

Reproducibility and Physiological Relevance: The ATCC Toxicology Portfolio Webinar Presented by Kevin Grady and Kevin Tyo March 3, 12:00 PM EST



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