

Addressing the Need for Accuracy and Traceability in Microbial Genomic Data: The ATCC Genome Portal

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Credible Leads to Incredible™





About ATCC

- Founded in 1925, ATCC is a not-for-profit organization with HQ in Manassas, VA, and an R&D and Services center in Gaithersburg, MD
- World's largest, most diverse biological materials and information resource for microbes – the "gold standard"
- Innovative R&D company featuring gene editing, microbiome, NGS, and advanced models
- cGMP biorepository

- Partner with government, industry, and academia
- Leading global supplier of authenticated cell lines, microorganisms, and molecular standards
- Sales and distribution in 150 countries,
 19 international distributors
- Talented team of 450+ employees, over onethird with advanced degrees



Overview

- The ATCC Genome Portal
- Traceability and authentication of microbial genomes
- Importance of authenticated reference genomes
- Development roadmap preview





The ATCC Genome Portal

The ATCC Genome Portal is a cloud-based platform that enables users to easily browse genomic data and metadata by simply logging into the portal



Download whole-genome sequences and annotations of ATCC materials



Search for nucleotide sequences or genes within genomes



View genome assembly metadata and quality metrics

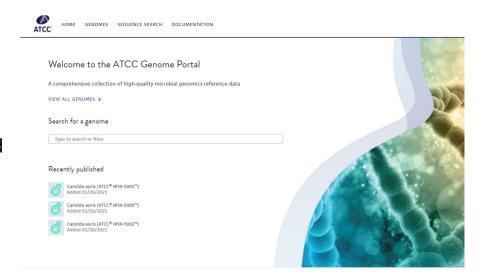
genomes.atcc.org



The ATCC Genome Portal

ATCC's authenticated reference genomes

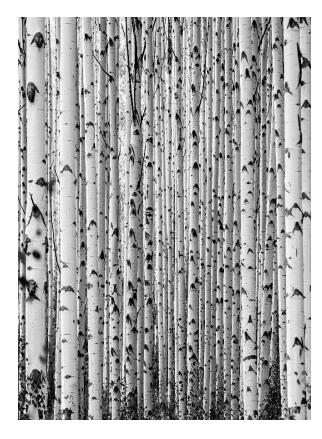
- 2017 2018 Planning and proof-of-concept experiments
- 2018 ATCC Commitment
 - Laboratory, staffing, resources, instrumentation, and bioinformatics development
- 2019 ATCC Enhanced Authentication Initiative
 - June 2019 beta launch at ASM Microbe 2019
 - Sept 2019 formal launch of the ATCC Genome Portal
- 2020+ Expansion
 - 1200+ authenticated, reference-grade whole-genome assemblies
 - Inclusion of viral, bacterial, and fungal genomes





Providing reference-quality genomes

Why - Challenge # 1



- Public databases routinely host genomic data that is cited as "ATCC," but...
 - Often no traceability back to genuine ATCC cultures
 - ATCC cannot authenticate 3rd party genomes
- So, how do researchers *know* which data set to use?
 - Which is the "correct" one?
 - Close enough?
- How do researchers have confidence in their selection?



Providing reference-quality genomes

Why - Challenge # 2



- How do we bring authentication into the genomics era while maintaining our commitment to our customers that we've fully and accurately authenticated our material?
- Typically, authentication* may refer to:
 - Morphology
 - Purity
 - Viability
 - Phenotypic testing
 - Genotypic testing
 - 16S ribosomal gene
 - ITS and D1D2



*not an inclusive list



Providing reference-quality genomes

Why - Challenge # 3



 Acknowledge there is a problem with reference genomes



 Work through a plan to address the problem



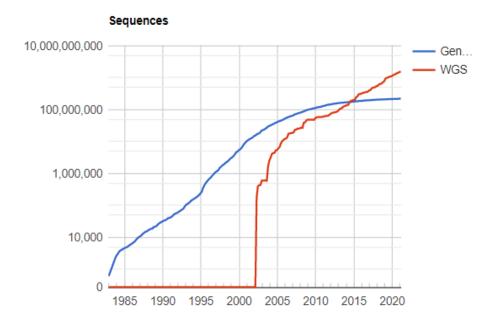
How do we effectively and easily provide customers with genomic data while not diluting it or burying it in a public database?



Reference genomes

Where can researchers turn to for "reference" genomes?

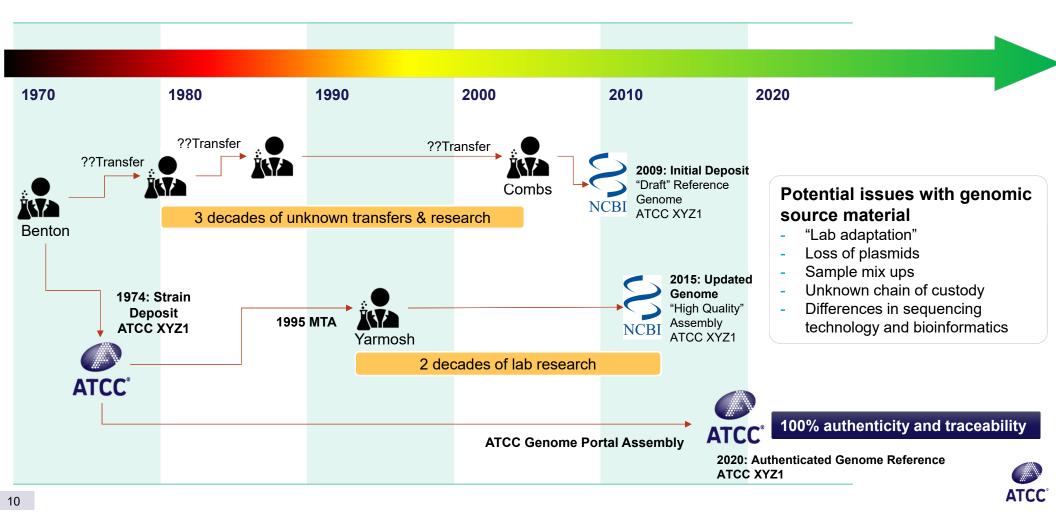
- De facto standard
 - The sequence database for the entire public scientific community
 - Contains numerous genomes
 - Genomes submitted by a variety of labs
- Relatively little curation
- Highly variable quality
- NEVER authenticated by ATCC



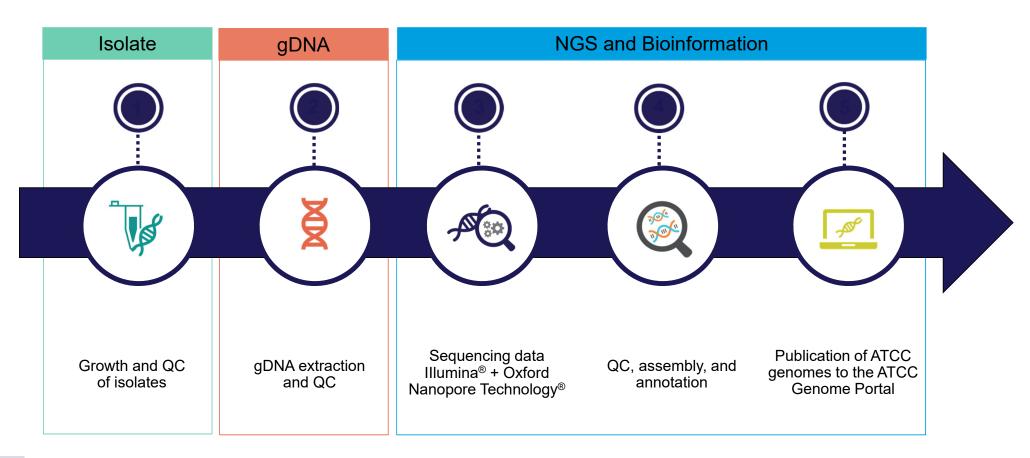
https://www.ncbi.nlm.nih.gov/genbank/statistics/



The elephant in the room: Authenticated reference genomes



Our process: Authenticated physical material coupled with reference-quality genome sequences



ATCC genome assembly process

ONT / Illumina reads uploaded to One Codex

Read-Based Contamination Quality Control with One Codex* Genome Assembly

Hybrid Assembly (Bacteria/Fungi)

Illumina-only Assembly (Viruses)

Quality Assessment

- Coverage
- Completeness
- Contamination

Genome Annotation Calculation of Assembly Level

Publish to the ATCC Genome Portal

Read Trimming
Illumina: fastp
ONT: FiltLong

Read-level kmer based taxonomic classification and estimation of strain abundance Bacteria: Unicycler

Fungi: MaSuRCA w/ FLYE

Viruses: Taxonomic bining

followed by SPADes

Bacteria: CheckM

Fungi: BUSCO

Viruses: curated

database*

Bacteria: Prokka

Fungi: BUSCO

Viruses: detect-viral-variants*

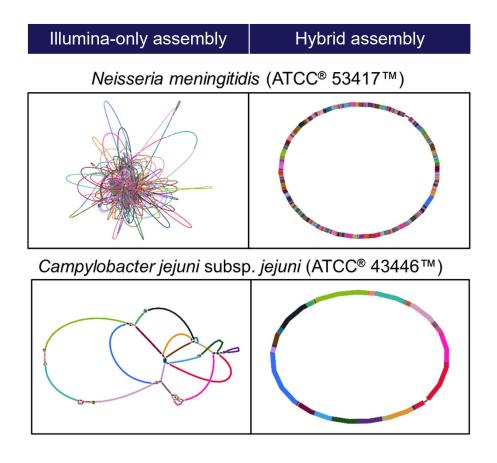
Based on NCBI's Assembly Level

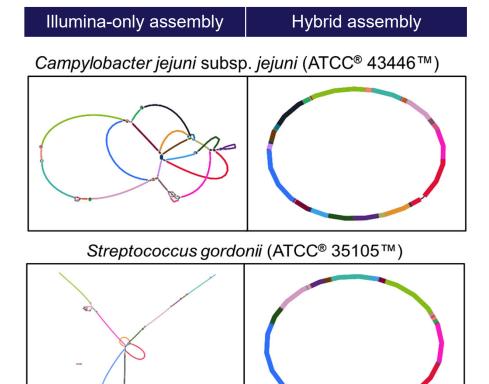
- Complete
- Scaffold



^{*} One Codex proprietary software

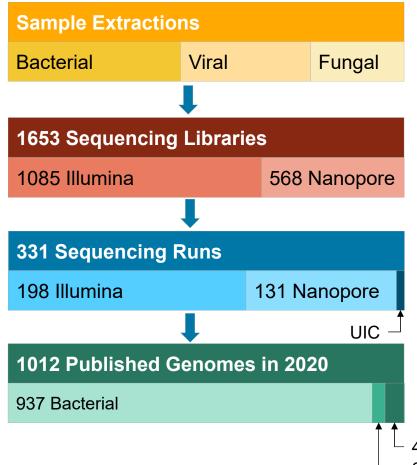
Advantage of hybrid assemblies







The ATCC Genome Portal





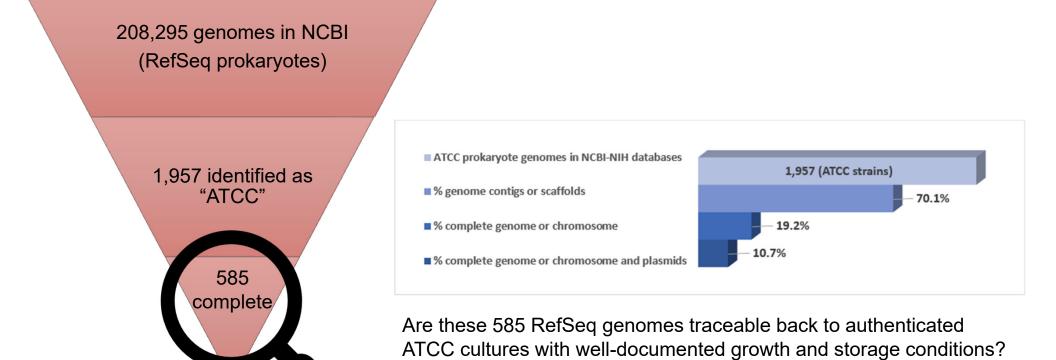
- 1,118 bacterial genomes (739 complete circularized, 391 type strains)
- 59 viral genomes
- 74 mycology genomes

- Monthly updates
- All genomes are traceable to ATCC's biomaterials
- Hybrid assemblies for all bacterial & fungal genomes
- All genomes annotated
- Additional improvements to fungal and viral genome annotations coming



42 Viral 33 Fungal

Reference genomes

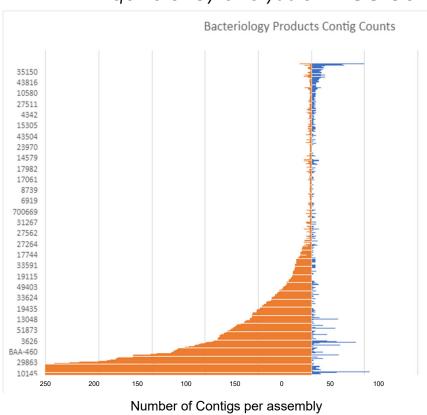




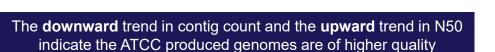
Genome assembly quality

Equivalency analysis of ATCC Genome Portal assemblies vs. those from public databases

Public Genome Assembly



ATCC GENOME PORTAL



ATCC Genome Portal Assembly

Bacteriology Assembly N50 Comparisons



#PUBLIC DATA

Evaluation of genome sequences from public databases

Product	NCBI existing reference genomes	NCBI assembly level (plasmids)	Sequencing technology and # of SNPs coverage		# of indels	Average coverage (variants)
	GCA_001593425.2	Complete Genome	Illumina (300.0x)	14	5	210.1
	GCA_000015425.1*	Complete Genome (2)	Not available	118	656	152.7
Acinetobacter baumannii (ATCC® 17978™)	GCA_014672775.1	Complete Genome (1)	PacBio (399.24x)	15	87	170.4
	GCA_013372085.1	Complete Genome (2)	Illumina, Nanopore (80x)	14	2	210.2
	GCA_004797155.2	Complete Genome (2)	PacBio (247.19x)	28	62	162.1
	GCA_001077675.1	Complete Genome (1)	Illumina, PacBio (153x)	15	6	135.9
	GCA_011067065.1	Complete Genome (2)	PacBio (231.08x)	60227	2486	165.6
Candida albicans	GCA_015227795.1	3, 081 Contigs	NovaSeq (16x)	10174	1573	265.6
(ATCC [®] 10231™)	GCA_002276455.1	2,219 Scaffolds	HiSeq (95x)	13408	2390	274.6
Meyerozyma guilliermondii (ATCC® 6260™)	GCF_000149425.1	9 RefSeq Scaffolds	Not available	505	1973	278.2
	GCA_006942155.1	9 Contigs	ONT+MiSeq (240x)	74	386	223.3
Clavispora lusitaniae (ATCC® 42720™)	GCF_000003835.1	9 RefSeq Scaffolds	Not available	587	2336	265.6
	GCA_003675505.1	109 Scaffolds	NextSeq (182x)	102	5142	236.9

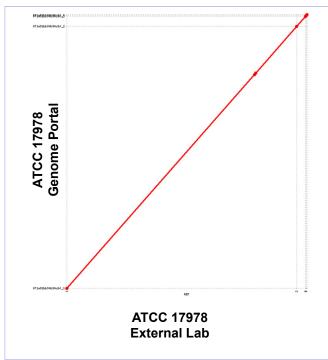
1 strain
7 assemblies
Unknown
origin of
materials

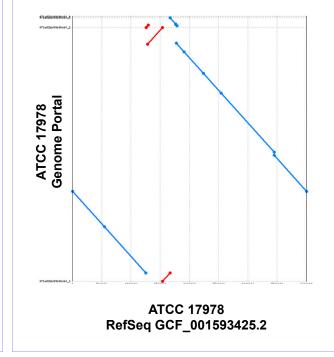


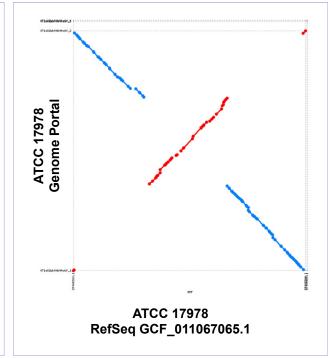
Evaluation of public sequences for ATCC 17978

MUMmer alignment with the de novo ATCC 17978 versus GenBank RefSeq genome assemblies GCF_001593425.2 and GCF_011067065.1

Acinetobacter baumannii strain 5377 (ATCC 17978)



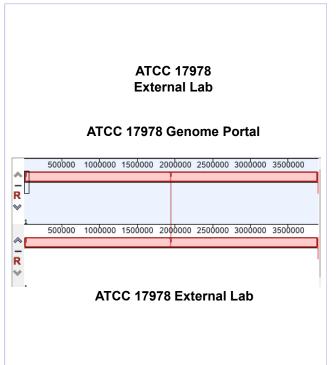


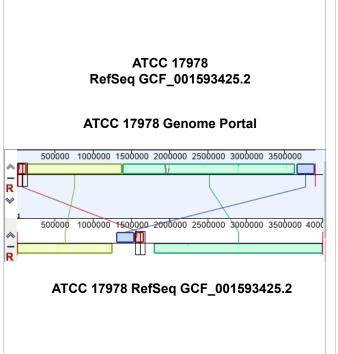


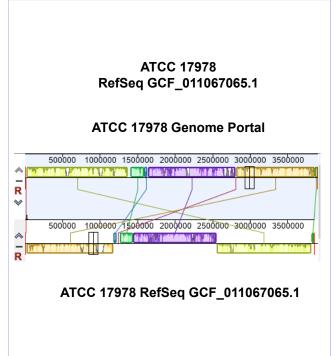


Evaluation of public sequences for ATCC 17978

Acinetobacter baumannii strain 5377 (ATCC 17978)









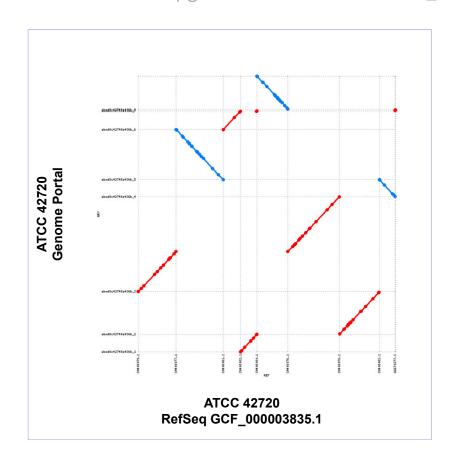
Evaluation of genome sequences from public databases

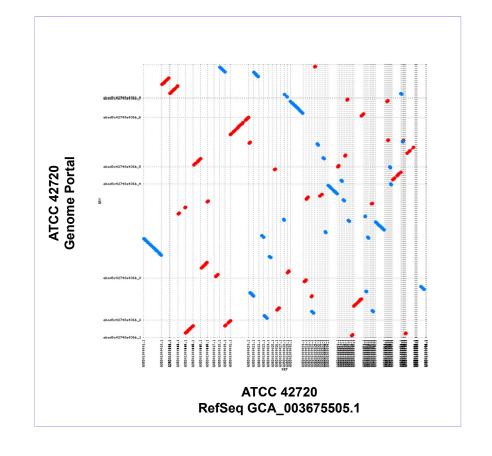
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Evaluation of public sequences for ATCC 42720

MUMmer whole genome alignments of ATCC de-novo genome assembly of ATCC 42720 versus GenBank RefSeq genome assemblies GCF_000003835.1 and GCA_003675505.1







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Selected timeline for (microbial) genomics standards

1970s	1980s 1982 – GenBank and ENA created	1990s	2000s 2005 – Genomic Standards Consortium established 2008 – Minimal Information on Genome Sequence (MIGS) specification 2009 – Genome Project Standards published by GSC	2010s 2012 – CDC NGS Standards for Clinical Testing (Nex-StoCT) 2014 – Viral Genome Reference Standards 2016 – FDA Draft Guidance on NGS for Pathogen Identification	2020s 2020 – ATCC Enhanced Authentication Initiative 2020 – ATCC Genome Portal Launch
1974 – Complete RNA genome bacteriophage MS2	1984 – Complete Epstein Barr virus genome	1995 – Complete genome of <i>Haemophilus</i> <i>influenzae</i>	2001 – Draft Human Genome 2007 - Genomic Encyclopedia of Bacteria and Archaea (GEBA) and Human Microbiome Project (HMP) launch.	2011 – GEBA II Launched	2020 – First "end to end" gapless genome for Human Chr. X



Recognition of the importance of traceability to biomaterials

PERSPECTIVE

nature biotechnology

The minimum information about a genome sequence (MIGS) specification

Dawn Field*¹, George Garrity², Tanya Gray¹, Norman Morrison^{3,4}, Jeremy Selengut⁵, Peter Sterk⁶, Tatiana Tatusova⁷, Nicholas Thomson⁸, Michael J Allen⁹, Samuel V Angiuoli^{5,10}, Michael Ashburner¹¹, Nelson Axelrod⁵, Sandra Baldauf¹², Stuart Ballard¹³, Jeffrey Boore¹⁴, Guy Cochrane⁶, James Cole², Peter Dawyndt¹⁵, Paul De Vos^{16,17}, Claude dePamphilis¹⁸, Robert Edwards^{19,20}, Nadeem Faruque⁶, Robert Feldman²¹, Jack Gilbert⁹, Paul Gilna²², Frank Oliver Glöckner²³, Philip Goldstein²⁴, Robert Guralnick²⁴, Dan Haft⁵, David Hancock^{3,4}, Henning Hermjakob⁶, Christiane Hertz-Fowler⁸, Phil Hugenholtz²⁵, Ian Joint⁹, Leonid Kagan⁵, Matthew Kane²⁶, Jessie Kennedy²⁷, George Kowalchuk²⁸, Renzo Kottmann²³, Eugene Kolker^{29–31}, Saul Kravitz⁵, Nikos Kyrpides³², Jim Leebens-Mack³³, Suzanna E Lewis³⁴, Kelvin Li⁵, Allyson L Lister^{35,36}, Phillip Lord³⁵, Natalia Maltsev²⁰, Victor Markowitz³⁷, Jennifer Martiny³⁸, Barbara Methe⁵, Ilene Mizrachi⁷, Richard Moxon³⁹, Karen Nelson^{5,40}, Julian Parkhill⁸, Lita Proctor²⁶, Owen White¹⁰, Susanna-Assunta Sansone⁶, Andrew Spiers⁴², Robert Stevens³, Paul Swift¹, Chris Taylor⁶, Yoshio Tateno⁴³, Adrian Tett¹, Sarah Turner¹, David Ussery⁴⁴, Bob Vaughan⁶, Naomi Ward⁴⁵, Trish Whetze¹⁴⁶, Ingio San Gil⁴¹, Gareth Wilson¹ & Anil Wipat^{35,36}

With the quantity of genomic data increasing at an exponential rate, it is imperative that these data be captured electronically, in a standard format. Standardization activities must proceed within the auspices of open-access and international working bodies. To tackle the issues surrounding the development of better descriptions of genomic investigations, we have formed the Genomic Standards Consortium (GSC). Here, we introduce the minimum information about a genome sequence (MIGS) specification with the intent of promoting participation in its development and discussing the resources that will be required to develop improved mechanisms of metadata capture and exchange. As part of its wider goals, the GSC also supports

can manipulate it to provide new solutions to critical problems. Such solutions include therapies and cures for disease, industrial products, approaches for biodegradation of xenobiotic compounds and renewable energy sources. With improvements in sequencing technologies, the growing interest in metagenomic approaches and the proven power of comparative analysis of groups of related genomes, we can envision the day when it will be commonplace to sequence tens to hundreds of genomes or more as part of a single study. At current rates of genome sequencing, it has been estimated that >4,000 bacterial genomes will be available soon after 2010 (ref. 1).

Given the importance of the growing genome collection, the capital investment in its creation and the benefits of leveraging its value "Source material identifier" is an exception; the GSC recommends this be a core descriptor, but as of yet, physical archives are not yet routinely created for all cases or types of biological material subjected to genome sequencing ...

This was in 2008.

We agree.

But, 12 years later "physical archives are [still] not yet routinely created" by groups doing whole genome sequencing.

Chain of custody of biomaterials is rarely or poorly documented.

Field, D. et al. (2008) 'The minimum information about a genome sequence (MIGS) specification', *Nature Biotechnology*, 26(5), pp. 541–547. doi: 10.1038/nbt1360.



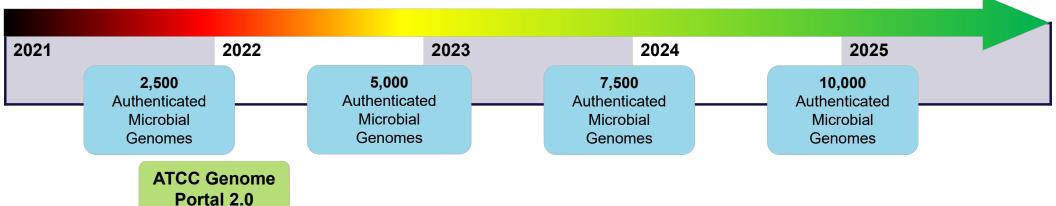
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ATCC Genome Portal development goals



- Fungal and viral genome annotation improvements
- Batch downloads
- API for bioinformatics
- Lab Protocols
 - Interactive taxonomy browser
 - Biosynthetic gene cluster analysis
 - Genome Forum
 - Microbial phenotypic data, including AMR

Authenticated Human & Animal Cell Line Data

RNAseq, WES, STR, cell imaging, flow cytometry, and more



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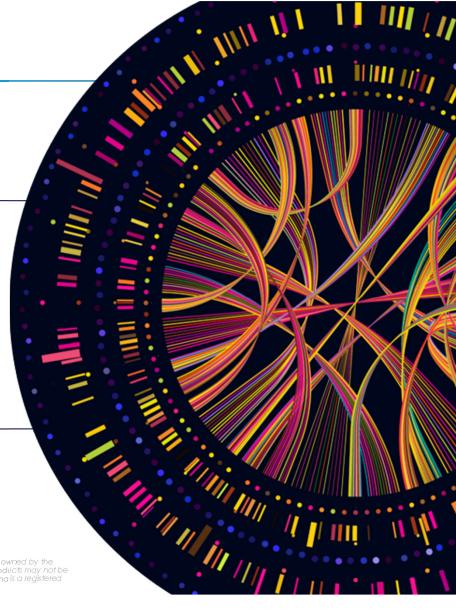
Partners

Juan Lopera, PhD Marco Riojas, PhD

... and One Codex!

JOIN OUR TEAM! We're hiring!

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Thank you

