The Biology of Anaerobic Bacteria and Predominant Propagation Practices

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- Founded in 1925, ATCC is a non-profit organization with headquarters in Manassas, VA
- World's premiere biological materials resource and standards development organization
- ATCC collaborates with and supports the scientific community with industry-standard biological products and innovative solutions
- Strong team of 400+ employees; over one third with advanced degrees



Established partner to global researchers and scientists





Outline



- 1. ATCC's anaerobe collection
- 2. Classification of anaerobes and specific examples
- 3. Nutritional and atmospheric considerations
- 4. Propagation methods



ATCC's anaerobe collection

942 active anaerobe holdings

465 are type strains

VPI Anaerobe Laboratory

- W.E.C. Moore, Lillian V.
 Holdeman-Moore, Tracy
 Wilkins, Elizabeth P. Cato
- Anaerobe Laboratory Manual





Top 5 anaerobes at ATCC

- Clostridium sporogenes (ATCC[®] 11437[™])
- Clostridium sporogenes (ATCC[®] 19404[™])
- Clostridium perfringens (ATCC[®] 13124[™])
- Bacteroides fragilis (ATCC[®] 25285[™])
- Porphyromonas gingivalis (ATCC[®] 33277[™])





Classification of anaerobes





Facultative anaerobes

- Can grow with or without the presence of oxygen
- Can metabolize energy aerobically (respiration) or anaerobically (fermentation)
- Ex: Propionibacterium acnes (ATCC[®] 6919™)
 - Isolated from facial acne
 - Has variable aerotolerance
 - Nutritional requirements:
 - All strains require Vitamin B5
 - Thiamine, biotin, and nicotinamide are stimulatory





Aerotolerant anaerobes

- Not inhibited by O₂, but does not use O₂ to generate ATP
- Uses fermentation to generate ATP
- Ex: Clostridium tertium (ATCC[®] 14573[™])
 - Originally isolated in 1917 by Captain Henry from war wounds
 - Type strain
 - Will grow slightly on an aerobic blood plate





Strict anaerobes

- Grows only in the absence of O₂ and may be inhibited or killed by O₂
- Generates ATP through anaerobic respiration or fermentation
- Ex: Fusobacterium nucleatum subsp. nucleatum (ATCC[®] 25586[™])
 - Isolated from a Cervico-facial lesion
 - Obligate anaerobic, Gram-negative rods with pointed ends
 - Most strains produce H₂S
 - All *Fusobacterium* species produce butyric acid
 - Require a rich medium for growth
- Ex: Clostridium sporogenes (ATCC[®] 19404[™])
 - Isolated from gas gangrene
 - Genetically similar to C. botulinum





Environmental anaerobes

- Includes extremophiles
- Selective media are often specific to genus or even species
- May require a gas mixture free of CO₂
- Ex: Thermoanaerobacterium thermosaccharolyticum (ATCC[®] 7956[™])
 - Requires a fermentable carbohydrate to grow
 - Type strain
 - Obligate anaerobe
 - Grows at 45°C 60°C





Methanogens

- Oregon Collection of Methanogens (OCM)
- Very strictly anaerobic with methane always being the product of catabolic metabolism
- Ex: Methanobrevibacter wolinii (ATCC[®] BAA-1170[™])
 - Isolated from sheep feces
 - Atmospheric requirements:
 - Media has a redox potential below -110 mV
 - Large amount of headspace and pressure
 - Nutritional requirements:
 - Nitrogen source (ammonia or N₂)
 - One or more B vitamins
 - Requires acetate and/or trypticase or yeast extract
 - Growth inhibited by bile salts





Media

- PRAS commercial media is superior
 - Boiled free of molecular oxygen
 - Autoclaved, dispensed, and packaged anaerobically
 - Light-proof packaging
- Indicator Rezazurin
 - Monitors redox potential of media
 - Non-toxic to bacteria and effective at low concentrations
 - Becomes colorless at a redox potential below -110 mV, remains pink above -51 mV





Common agar media

- Bases for blood agar media:
 - Brucella
 - Columbia-based blood agar
 - Tryptic Soy-based blood agar
 - Brain heart infusion w/ 0.5% yeast extract
- Supplements to enhance growth:
 - 5% sheep, horse, or rabbit blood
 - Vitamin K1 (1 μg/mL)
 - Hemin (5 µg/mL)
- Selective agars are commercially available
 - Bacteroides Bile Esculin agar (BBE)





Common broth media

- Chopped meat
- Reinforced clostridial
- Peptone yeast extract broth with glucose (PYG)
- Supplemented tryptic soy
 - ATCC Medium 2722
 - Additions of Yeast extract, hemin, and vitamin K1
- Broth media vessels
 - Hungate tubes
 - Balch tubes





Inhibitors

- Inhibitors that affect the quality of media
 - Oxygen
 - Light
 - Moisture/dehydration
- Inhibitors that affect cell growth
 - Bile
 - Reducing agents
 - Oxygen/incorrect gas mixture
 - Failure to add supplements





Reducing agents

- Reduces the redox potential of growth media
 - The reducing agent is oxidized by the oxygen in the media and therefore the media is reduced of the oxygen
- Ex: Coenzyme, cysteine, sodium sulfide
 - Select the reducing agent that is already used in the media formulation
 - Typically use 2 mL reducing agent per 100 mL media
 - Incubation at 37°C may speed up the process
 - Allow reducing agent to react for at least 1 hour, preferably overnight





Oxygen toxicity

Defenses

- Hemoglobin
- Superoxide dismutase
- Catalase
- Peroxidase
- Vitamin C
- Vitamin E
- Uric Acid

Damage

- Oxidation of membrane lipids
- Inactivation of enzymes
- Genetic damage



Why is oxygen toxic?







Why is oxygen toxic?





Gas mixtures

- 100% Nitrogen
- 97% N₂ 3% H₂
- 80% N₂ 10% H₂ 10% CO₂
- 80% N₂ 20% CO₂
- 80% H₂ 20% CO₂
- 100% Methane





Propagation methods: The roll tube







Image of anaerobic roll tubes courtesy of CEBTech Services. Please note that proper personal protective equipment is recommended.

Propagation methods: The roll tube





Propagation methods: The cannula system





Propagation methods: Syringe, needle, and oxygen-free gas





Propagation methods: Jars and catalysts





Propagation methods: Jars and catalysts







Propagation methods: Anoxomat[™]





Propagation methods: Anaerobe chambers









Propagation methods: Anaerobe chambers





References

- 1. Strother WH, Wallenstein P. From VPI to State University: President T. Marshall Hahn Jr. and the Transformation of Virginia Tech, 1962-1974. Mercer University Press, 2004.
- 2. Cox M. How to Isolate and Maintain Anaerobic Bacteria? ESCMID eLibrary, available online: <u>https://www.escmid.org/escmid_publications/escmid_elibrary/?q=+mike+cox&id=2173&L=0&x=0&y=0</u>
- 3. Jouseimies-Somer H, et al. Wadsworth-KTL Anaerobic Bacteriology Manual, 6th Edition, 2002.
- 4. Fukushima RS, Weimer PJ, and Kunz DA. Photocatalytic Interaction of Resazurin N -Oxide with Cysteine Optimizes Preparation of Anaerobic Culture Media. Anaerobe 8(1): 29-34, 2002.
- 5. Macy JM, Snellen JE, and Hungate RE. Use of syringe methods for anaerobiosis. The American Journal of Clinical Nutrition 25: 1318-1323, 1972.
- 6. Bacic MK, Smith CJ. Laboratory Maintenance and Cultivation of Bacteroides species. Current Protocols in Microbiology, 2008.
- 7. Willis AT. Anaerobic Bacteriology: Clinical and Laboratory Practice. 3rd Edition. Elsevier Ltd, 1977.
- 8. Collard PJ. The Development of Microbiology. CUP Archive, 1976.
- 9. Fildes P, McIntosh J. An improved form of the McIntosh and Fildes' anaerobic jar. British Journal of Experimental Pathology 2: 153-154, 1921.
- 10. DSMZ. Cultivation of Anaerobes. Available online: <u>https://www.dsmz.de/fileadmin/Bereiche/Microbiology/Dateien/Kultivierungshinweise/englAnaerob.pdf</u>
- 11. Remel. Technical Manual of Microbiological Media. Available online: Remel Technical Manual for Microbial Media <u>http://site3.auth.cscl.commonspotcloud.com/cms/intranet/msds/upload/MSDS-Remel-Tech-Manual-of-Microbiological-Media.pdf</u>
- 12. Bergey DH, Holt JG. Bergey's Manual of Determinative Bacteriology, 9th Edition. Lippincott Williams & Wilkins, Philadelphia, 2000.
- **13**. Goodfellow M, *et al.*, eds. Bergey's Manual of Systematic Bacteriology, 2nd Edition, Volume 5, Part A. Springer, New York, 2012.
- 14. Boone DR, Castenholz RW, eds. Bergey's Manual of Systematic Bacteriology, 2nd Edition, Volume 1. Springer, New York, 2001.
- 15. Krieg NR, *et al.*, eds. Bergey's Manual of Systematic Bacteriology, 2nd Edition, Volume 4. Springer, New York, 2010.
- 16. Vos P, *et al.*, eds. Bergey's Manual of Systematic Bacteriology, 2nd Edition, Volume 3. Springer, New York, 2009.
- 17. Miller TL, Lin C. Description of *Methanobrevibacter gottschalkii* sp. nov., *Methanobrevibacter thaueri* sp. nov., *Methanobrevibacter woesei* sp. nov. and *Methanobrevibacter wolinii* sp. nov. IJSEM 52: 819-822, 2002.
- 18. Douglas HC, Gunter SE. The Taxonomic Position of *Corynebacterium acnes*. Journal of Bacteriology 52: 25-32, 1946.
- 19. Holdeman LV, Cato EP, Moor WEC. Anaerobe Laboratory Manual, 4th Edition. Virginia Polytechnic Institute and State University, Anaerobe Laboratory, Blacksburg, VA, 1977.



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