

ATCC Microbiology – Best Practices for Stock Maintenance

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About ATCC

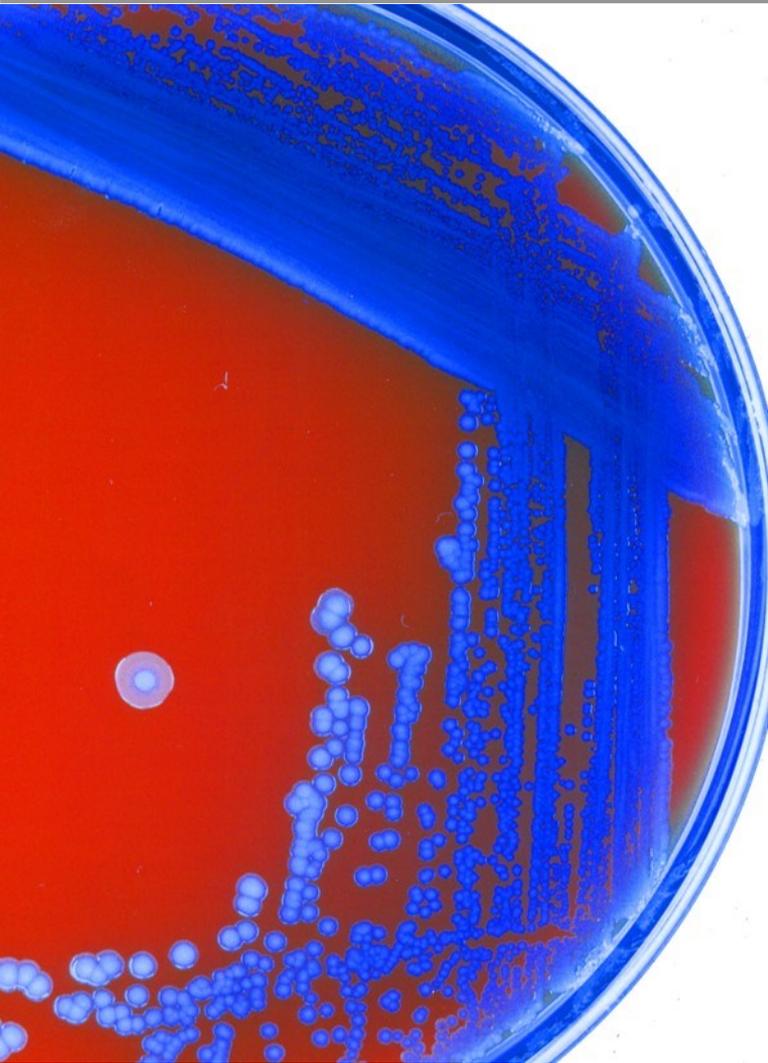
- Founded in 1925, ATCC is a non-profit organization with headquarters in Manassas, VA
- World's premiere biological materials resource and standards development organization
- ATCC collaborates with and supports the scientific community with industry-standard biological products and innovative solutions
- Strong team of 400+ employees; over one third with advanced degrees



Established partner to global researchers and scientists



Outline



- ATCC Microbiology workflow
- Passage matters
- Seed stock concept
- Principles of preservation and recovery
- Types of cultures and requirements
- Quality control
- Storage and safe-keeping

ATCC Microbiology workflow



- Deposits come from every continent
- Newly identified species, outbreak strains, type strains, etc.
- Grow/expand deposit to a large enough scale to distribute
- Preserve according to the best method for the species
- Some cultures have to be maintained in continuous culture
- Confirm identity noted by the depositor
- Minimum test is purity, viability, and sequence
- Goal is two platforms to confirm the item
- Worldwide distribution
- Ship frozen/live materials

Passage matters



Why is it important to minimize the number of passages?

- Reduce the chance of contamination
- Reduce/prevent genetic drift and mutations
- Minimize phenotypic variations

How many passages are acceptable?

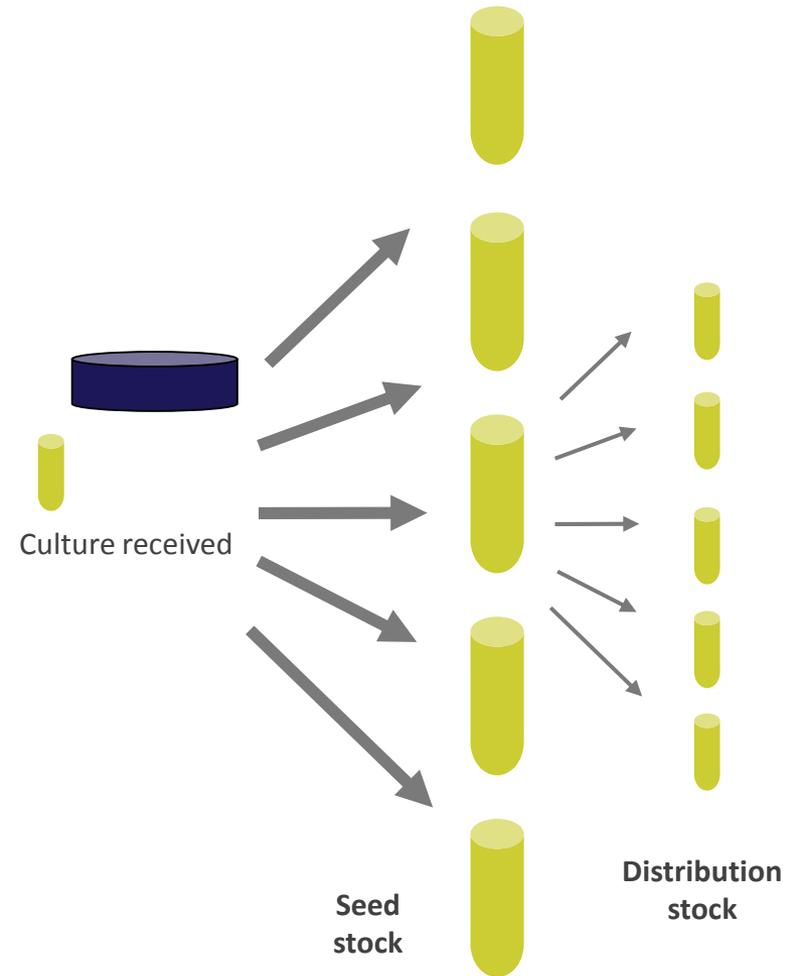
Passage matters



USP <51> Antimicrobial Effectiveness Testing

“The viable microorganisms used in the test must not be more than five passages removed from the original ATCC culture.”

Production



- Preserved cultures remain as close as possible to the original culture
- Seed stock is archived for future replenishment
- Distribution stock are used for distribution
- Authentication compares: Seed, Distribution, Initial culture

Preservation defined

- Importance of preserving cultures:
 - Minimize passage
 - Maintain valuable stocks
- Most common methods:
 - Cryopreservation (freezing)
 - Lyophilization (freeze-drying)



Principles of freezing

- Ice crystals develop outside of the cells first, increasing the concentration of solutes & causing water to migrate out of the cells
- The addition of cryoprotectants can have different effects on this process:
 - Most cryoprotectants bind either the electrolytes, which increase in concentration during freezing, or the water molecules to delay freezing (e.g., glycerol)
 - Some cryoprotectants alter cell permeability (e.g., DMSO) to facilitate movement of water out of the cells
- The amount of water remaining inside the cells is critical:

Too much water = too many ice crystals

Too little water = damage from high intracellular osmolality

- In addition to using a cryoprotectant, the rate of cellular dehydration during the freezing process can be managed by using a $-1^{\circ}\text{C}/\text{minute}$ cooling rate

Storage and recovery of frozen cultures

- Do not store frozen cultures in a freezer with a defrost cycle; this will expose the cultures to higher temperatures
- -130°C is the critical temperature for long-term storage of biological materials
- -80°C is sufficient for most bacteria and fungi for short-term storage (5 years or less)
- Thawing should be rapid at 37°C
- Initiate the culture in the medium recommended by ATCC or in the same growth medium used prior to freezing
- Incubate at optimum temperature & atmosphere

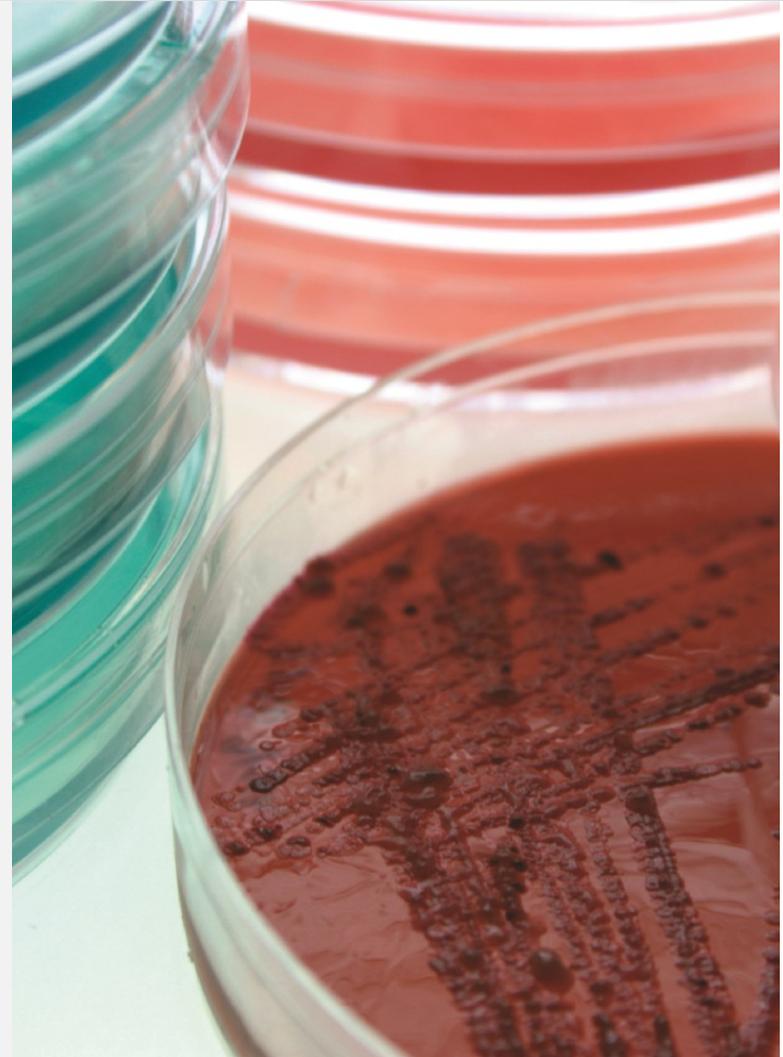


Principles of freeze-drying

- Most cultures in the ATCC collection are lyophilized
- This method of preservation is still focused on managing both water & solvents, which are removed by sublimation
- Three stage approach:
 1. Freeze the material
 2. Primary drying – Removes most of the water and is achieved by applying just enough heat & pressure to vaporize (but not melt) the frozen liquid
 3. Secondary drying – Removes residual moisture by applying heat and low pressure to desorb any remaining bound water
- Results in stable, dehydrated material with a relatively indefinite shelf-life when stored appropriately
- Cryoprotectants vary based on the organism and method used (e.g., commercial, component, or manifold systems) – the most common cryoprotectant used in lyophilization is 20% skim milk

Storage and recovery of freeze-dried cultures

- Store freeze-dried cultures at 2-8°C
- Lower temperatures (-20°C or lower) will yield a longer shelf life
- Keep sealed, away from moisture and oxygen (i.e., free radical that will react with the dried material) and moisture
- Open vial and revive organisms
 - Instructions online at www.atcc.org
 - Use media recommended by ATCC
 - Incubate under appropriate conditions



Bacteriology

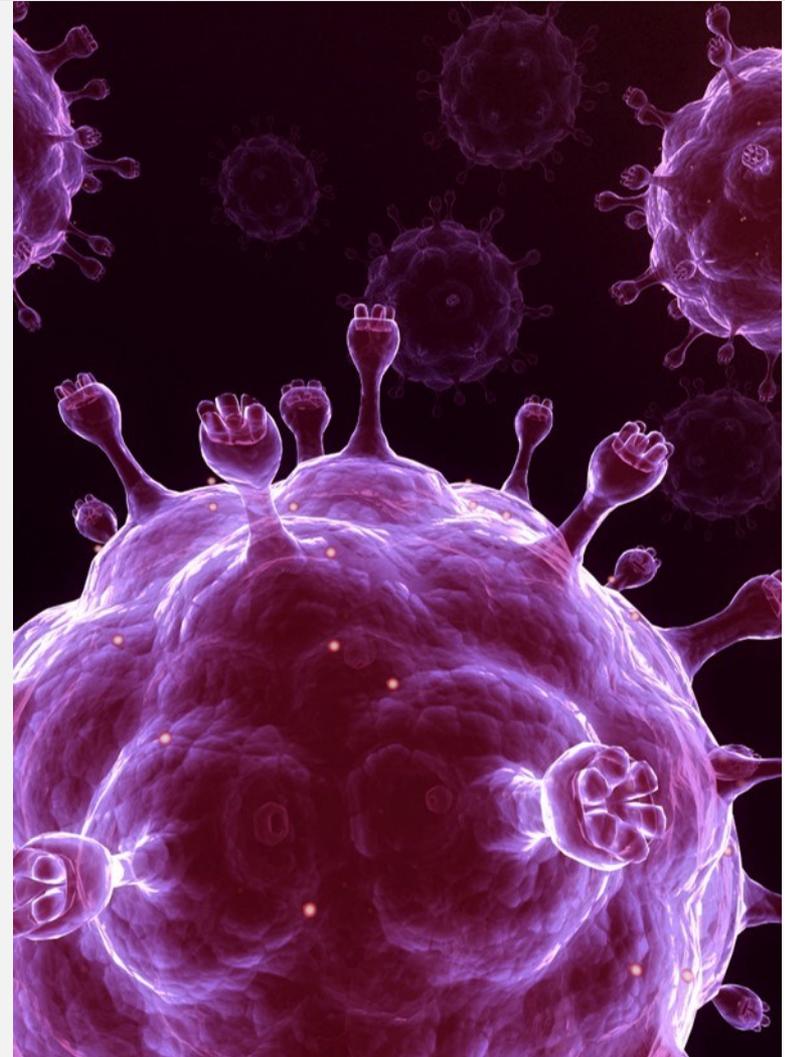
- Most can be freeze-dried
- Almost all can be frozen

- Special cases:
 - Acidophiles (e.g., *Thiobacillus acidophilus*)
 - Anaerobes (e.g. methanogens)
 - Phages
 - Chlamydia and Rickettsia



Animal Virology

- Most viruses can be preserved using an uncontrolled method of freezing
- Cultures that are dependent upon the viability of their host cell should be preserved with a controlled method of freezing
- Viruses grown in eggs are typically frozen in the allantoic fluid or yolk sac



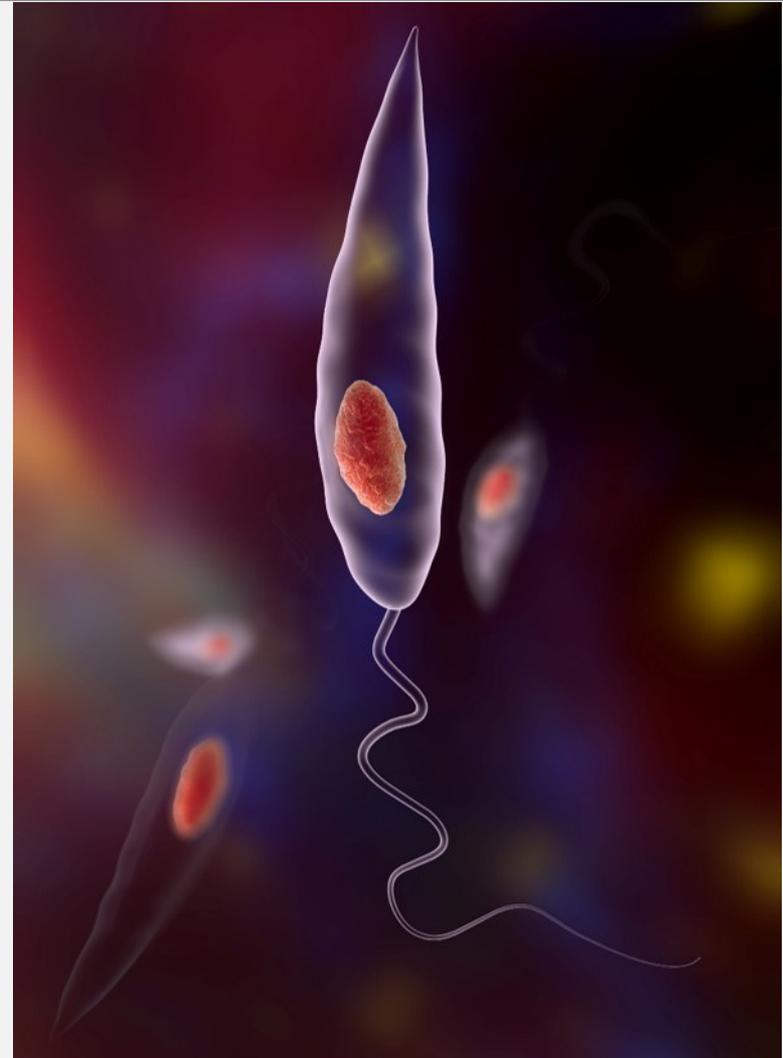
Mycology & Protistology

Mycology

- Yeasts can be preserved like bacteria
- The preservation method for filamentous fungi depends on whether the strain is sporulating or non-sporulating
- Slime molds can be frozen or freeze-dried

Protistology

- Over 100 protocols are used to preserve ATCC protozoa and algae
- Many strains are frozen, only a few can be freeze-dried or dried, and some are maintained as living cultures



Factors that affect post-preservation viability

- Storage temperature was not maintained
 - Avoid refreezing even without complete thawing (i.e., avoid frost-free freezers)
 - Avoid thawing near-by materials when retrieving one ampoule from the freezer
- Age of the culture was too young or too old
 - Best to harvest at late log or early stationary phase
- Culture or growth conditions
 - Aerated cultures are more permeable, allowing for more rapid water loss
 - Gram type (Gram-positive easier to freeze-dry) or fastidious culture



Microbial strain authentication

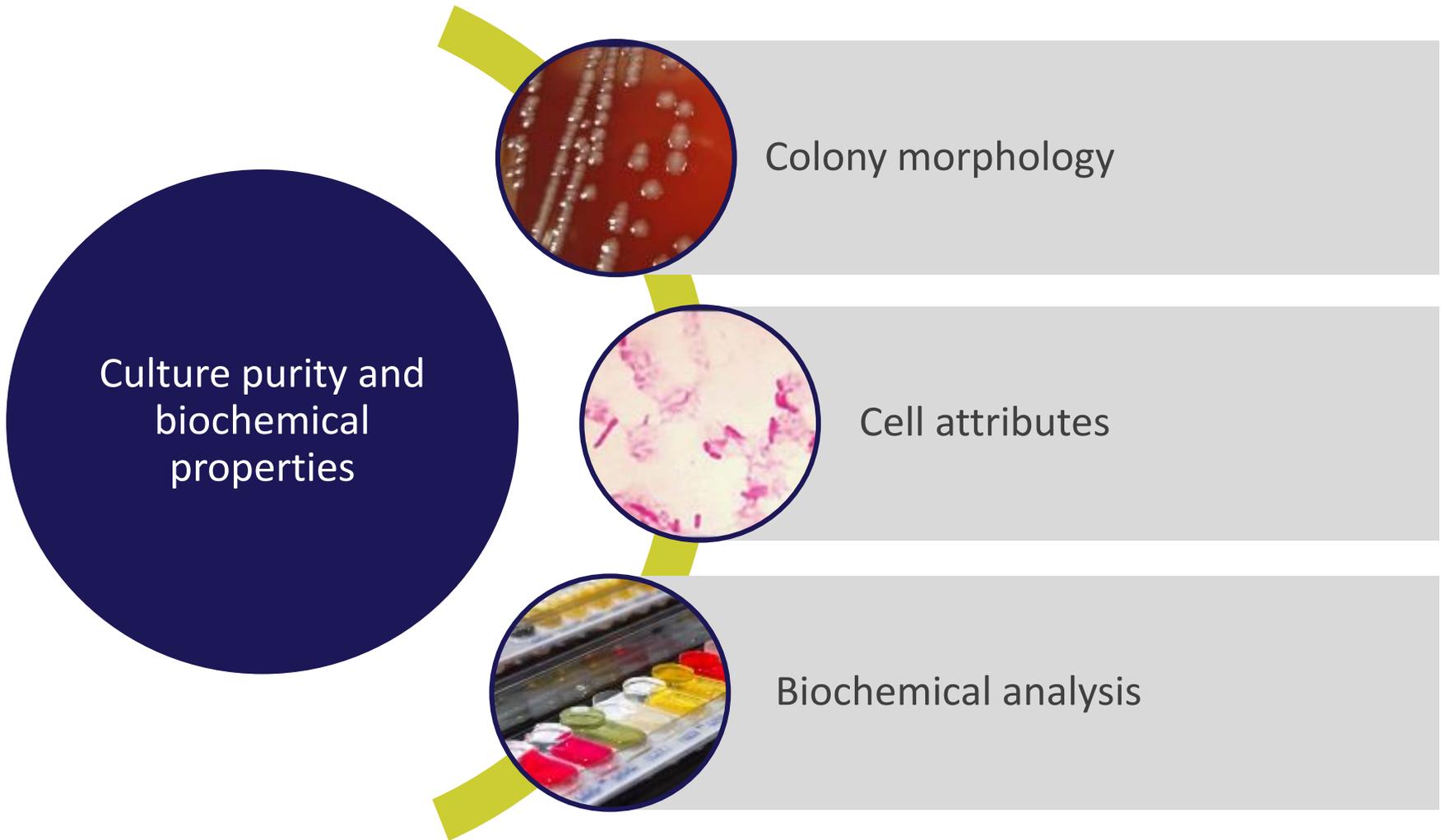


ATCC utilizes both classical and modern techniques

- Phenotypic analysis
- Genotypic analysis
- Proteotypic analysis
- Functional analysis

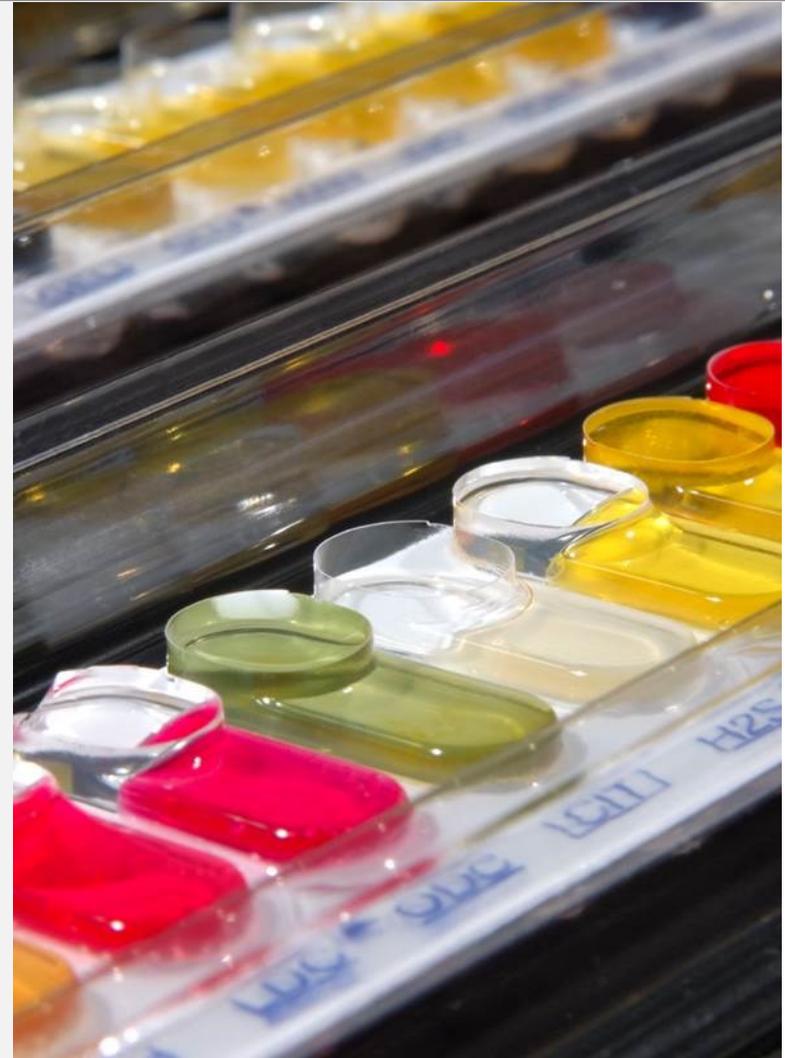
No single method of identification is sufficient

Phenotypic testing



Phenotypic testing platforms at ATCC

- API® ID
- Remel RapID™
- Biolog Microbial ID
- VITEK® 2
- VITEK® 2/MS
- Serotyping
- Hundreds of biochemical tests



Genotypic testing

Sequencing



Toxinotyping



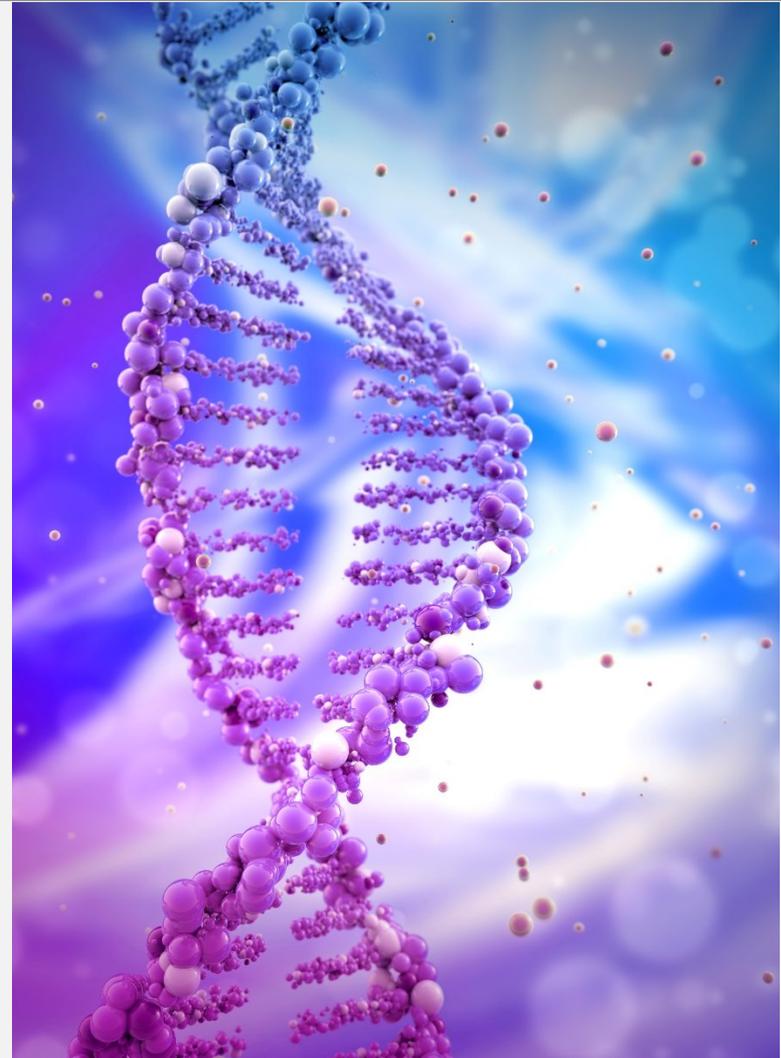
Ribotyping



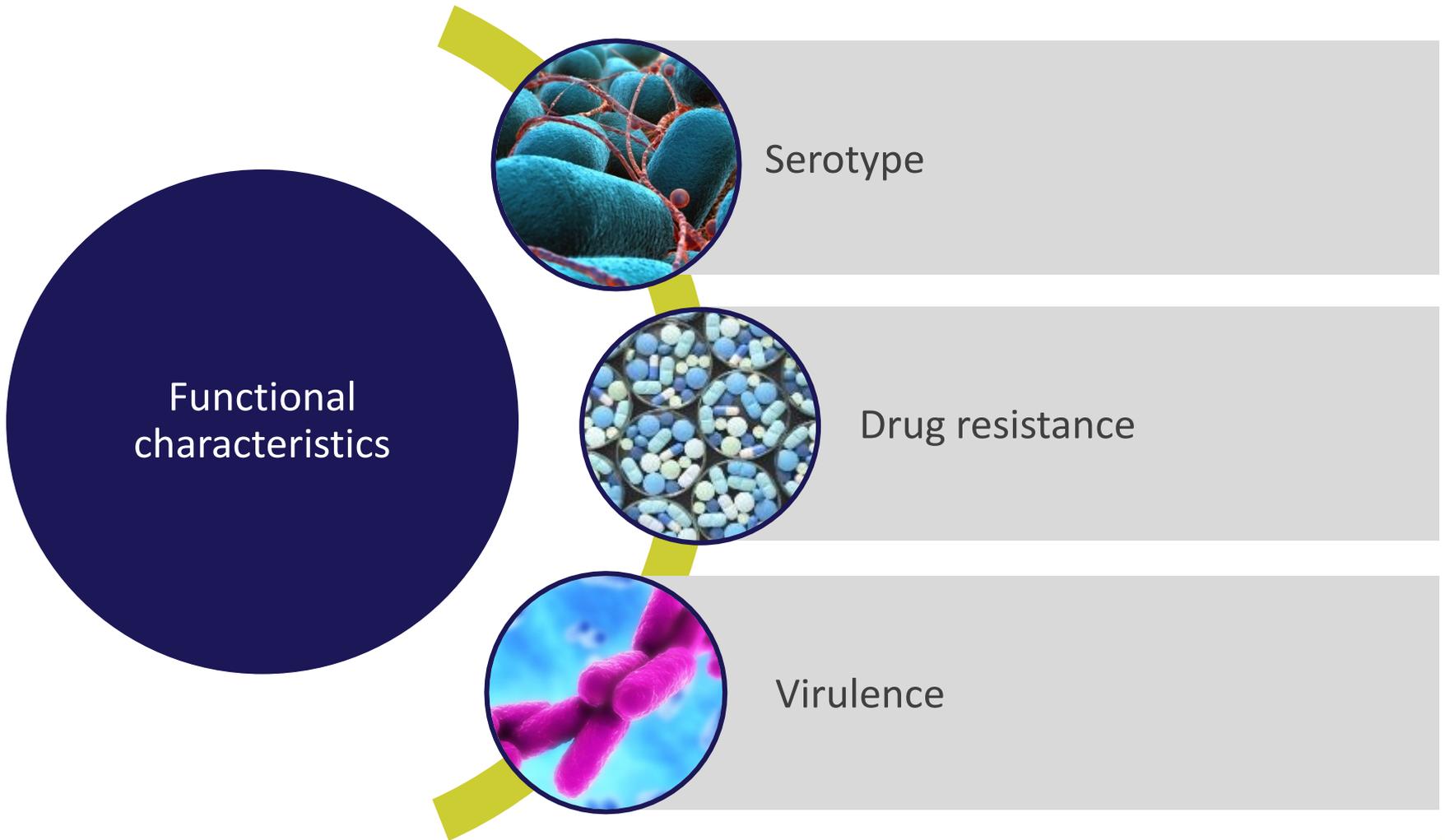
Targeted gene sequencing

Genotypic testing platforms at ATCC

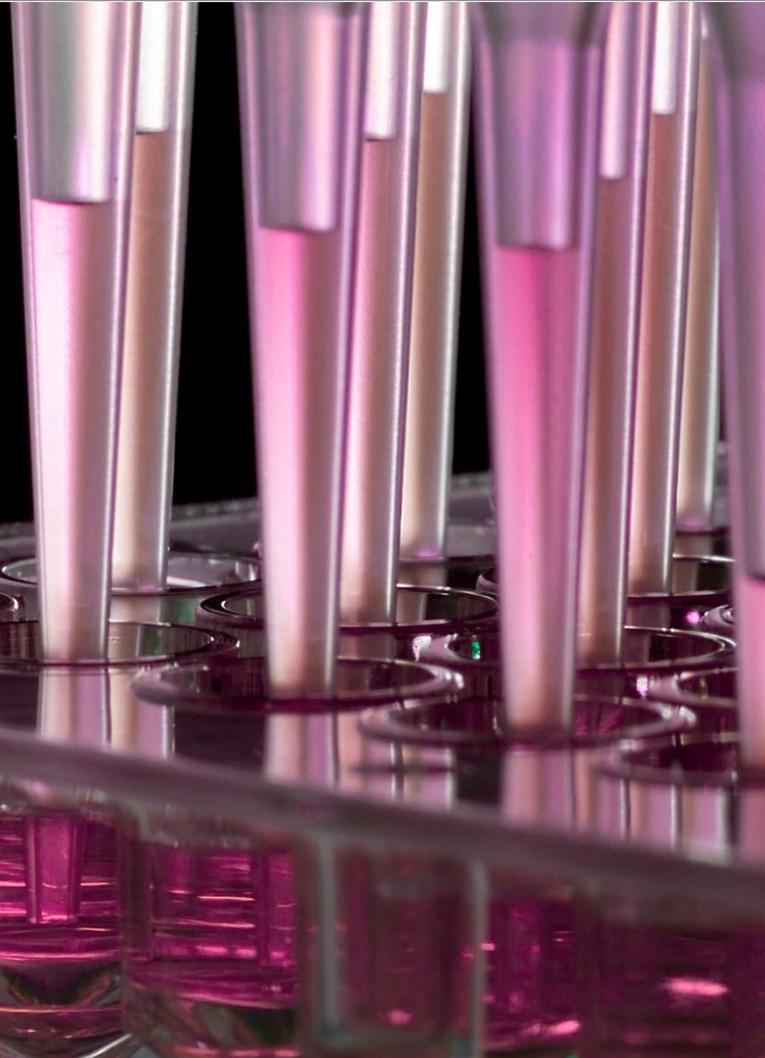
- ABI 3500 XL analyzers
 - MicroSeq® database
 - Public databases
 - Sequence provided by the depositor
- PCR
- DuPont™ RiboPrinter®
- Illumina® MiSeq



Functional testing



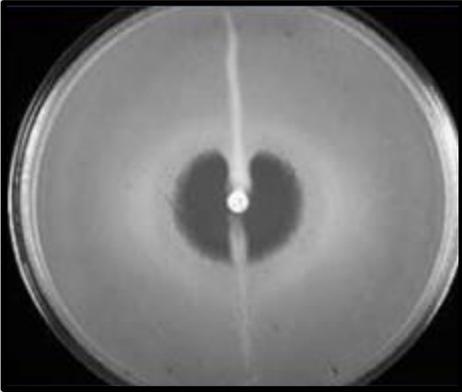
Functional testing platforms at ATCC



- AST
- Toxin assay
- IFA
- ELISA
- Cytotoxicity assay

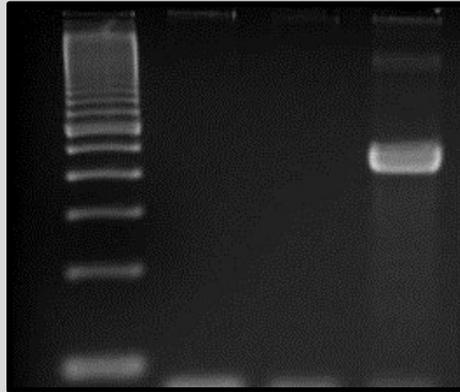
Verification of drug resistance

Modified Hodge Test



- Recommended by CLSI and the CDC for the detection of carbapenemase production

Endpoint PCR



- Endpoint PCR used to detect the presence or absence of genes required for antibiotic production

Antibiotic Profiling



- VITEK used to analyze resistance to various antibiotic classes

Safekeeping and security

- Keep good records
- Freezer alarms
- Monitor freezer temperature
- Call list for alarm response
- Spare freezers
- Backup storage (separate freezer or off-site)
- Dry ice cooling if necessary for -80°C freezers



Storage temperature

- Storage temperature must be maintained at all times
- Warming and re-cooling, even in the absence of thawing, can be detrimental
- Avoid removing the entire inventory when retrieving one item
- -80°C is sufficient for most bacteria and fungi for short-term storage (5 years or less)
- -130°C is the critical temperature for biological materials
- -20°C is only adequate for freezing organisms when the stability has been established at this temperature



Summary points

Know your cultures and minimize passage

- Understand *in vitro* requirements of the organism
- Use recommended media and atmospheric conditions
- Track and minimize passage

Preserve valuable stocks using a method that best suits the organism

- Determine the optimal preservation method for the culture based on the organism's attributes and length of time in storage
- Use a cryoprotectant that best suits the method of preservation

Verify stock identity when preparing MCB/WCB

- No one method is satisfactory for determining identity of an organism
- Select phenotypic and genotypic methods of testing to validate authenticity – polyphasic approach

Maintain safekeeping and optimal storage

- Store materials under appropriate conditions
- Monitor equipment and temperatures
- Keep good records

Disclaimers



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