

# Functionally characterized human PBMCs: An improved *in vitro* model of human immune response

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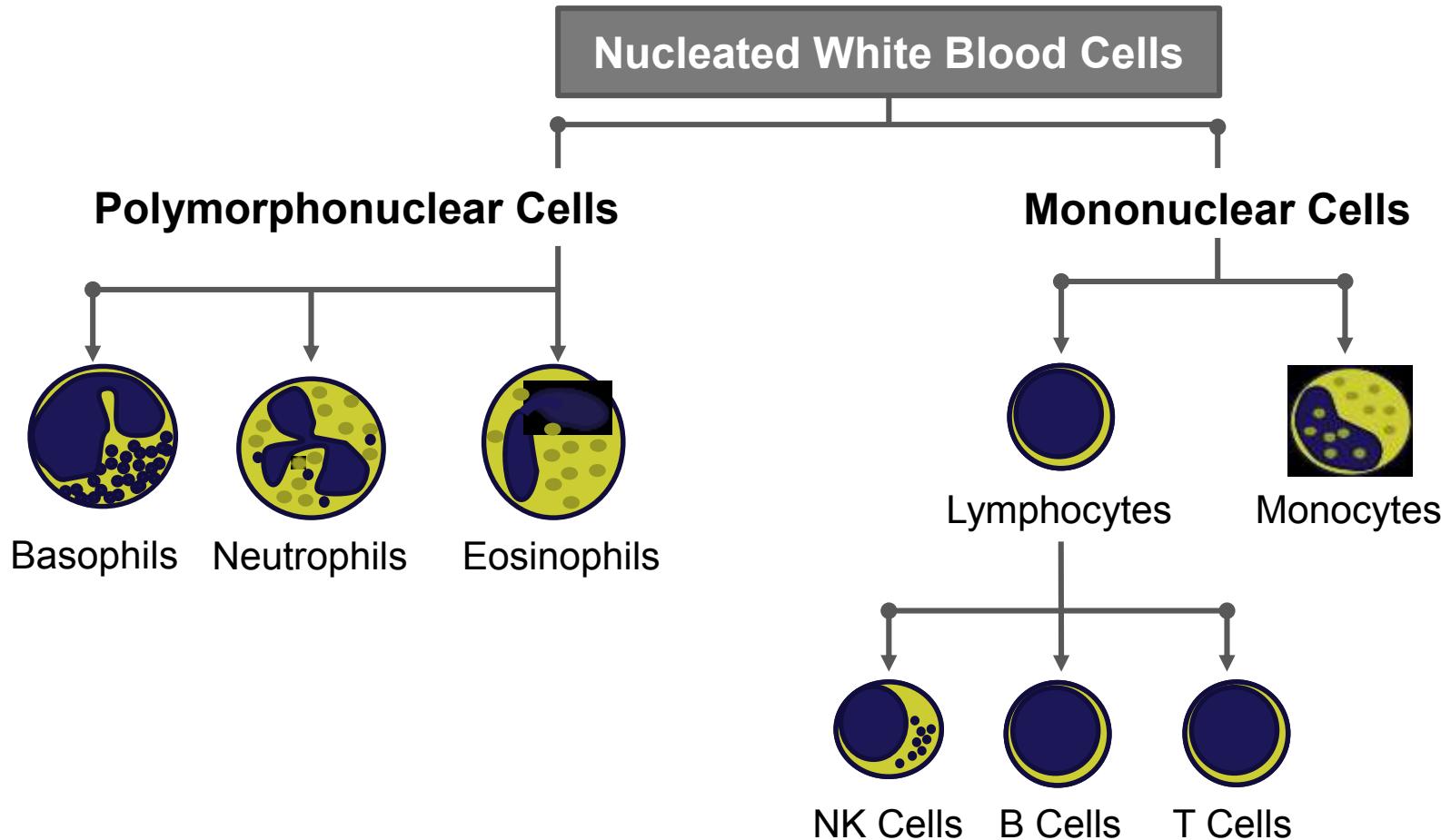
# About ATCC

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- World's premiere biological materials resource and standards development organization
- ATCC collaborates with and supports the scientific community with industry-standard biological products and innovative solutions
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Established partner to global researchers and scientists

# Peripheral blood mononuclear cells (PBMC): Cellular composition

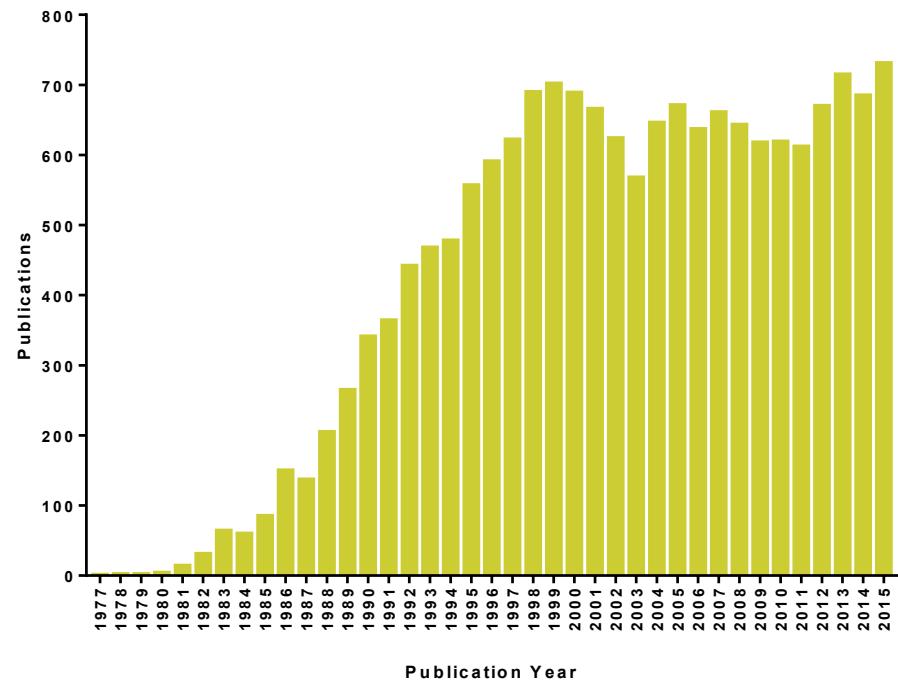


# Peripheral blood mononuclear cells

The main source of the primary human cells in immunology research

Currently used in:

- Basic research
- High-throughput drug screening
- Development of vaccines, biologics, and cellular therapeutics
- Assessment of immunogenicity and immuno-toxicity of new drug candidates



# Functional activity of human PBMC is extremely variable

In this webinar, we will discuss the variability of PBMC functional activity and the main approaches to minimizing the impact of PBMC variability on your research

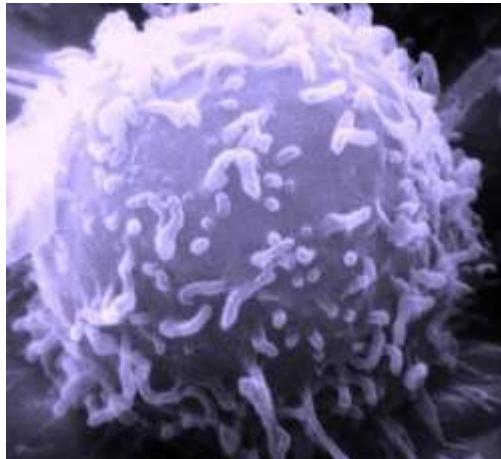


*Salmonella* invading an immune cell,  
photo credit: NIAID

# Factors affecting PBMC assay variability

## Donor Variability

- Genetic diversity
- Environmental factors
  - Immunizations
  - Nutrition
  - Latent infections

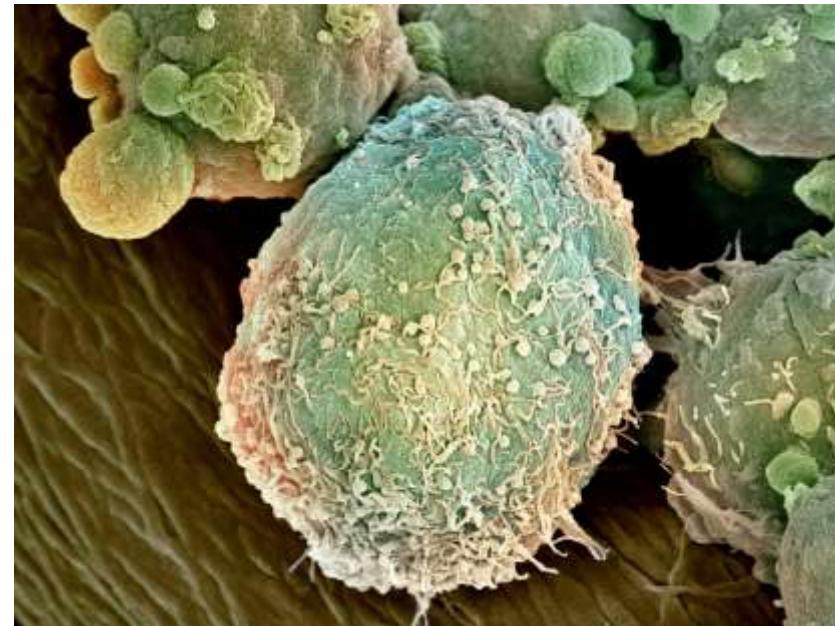


## Cells/Assay Variability

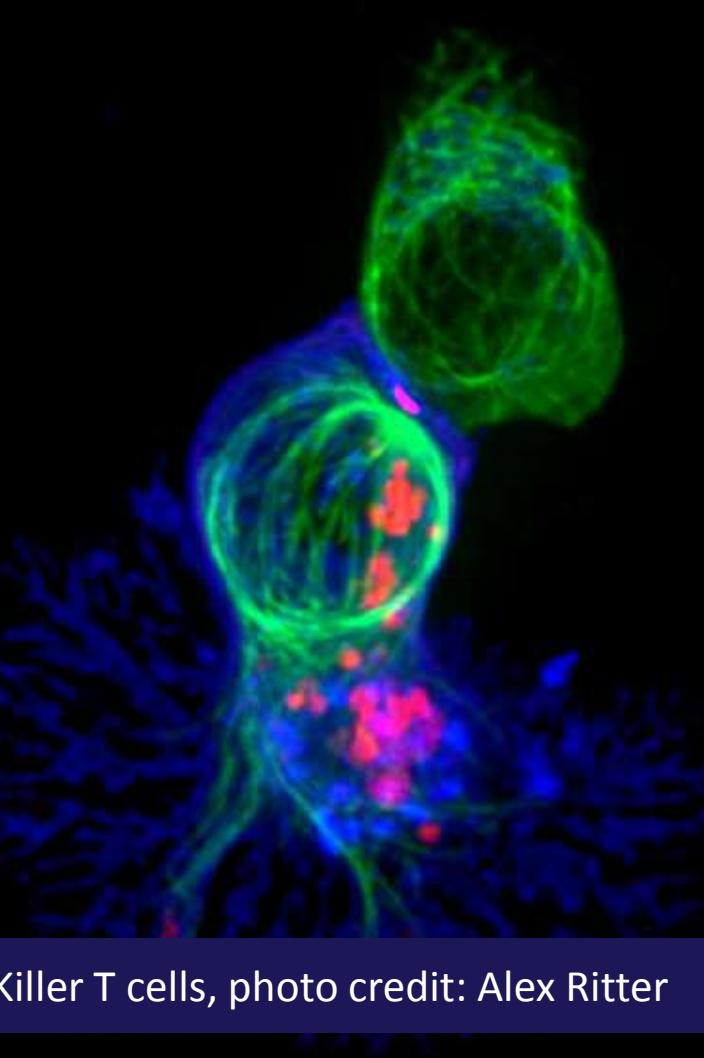
- Cryopreservation technique
- Cell viability
- Cellular composition (immunophenotype)
- Medium composition
- Quality of cell activating reagents

# Minimizing the impact of donor variability

- Stable, pre-qualified pool of adult healthy volunteer donors
- Complete donor information
  - Age, gender, ethnicity, blood type, and HLA class I and II
- At the time of collection, all donors were screened and certified to be free of:
  - Syphilis
  - Hepatitis B virus (HBV)
  - Human T-lymphotropic virus (HTLV)
  - Human immunodeficiency virus (HIV)
  - West Nile virus (WNV)
  - *Trypanosoma cruzi*



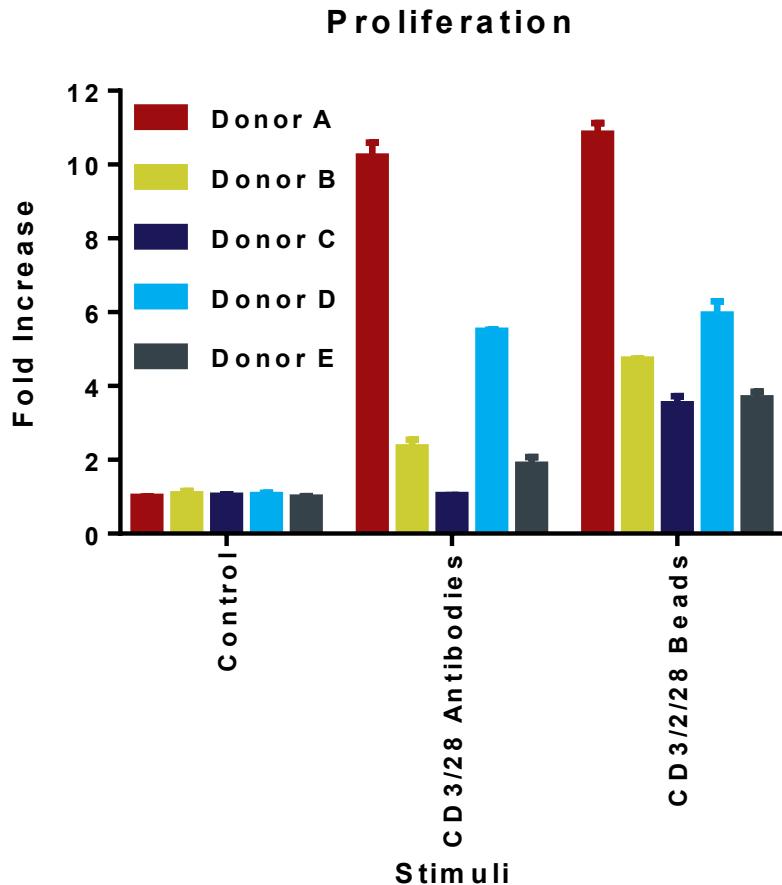
# Ensuring quality of the collected cells



Killer T cells, photo credit: Alex Ritter

- Cells purified by gradient centrifugation
- Greater than 95% viability at the time of collection
- Fully validated cryopreservation protocol
  - Optimized protein-free cryopreservation solution
  - Fully controlled freezing and storage conditions
- Comprehensive immunophenotype characterization of the collected cells
- Greater than 90% post-thaw viability

# Functional assay



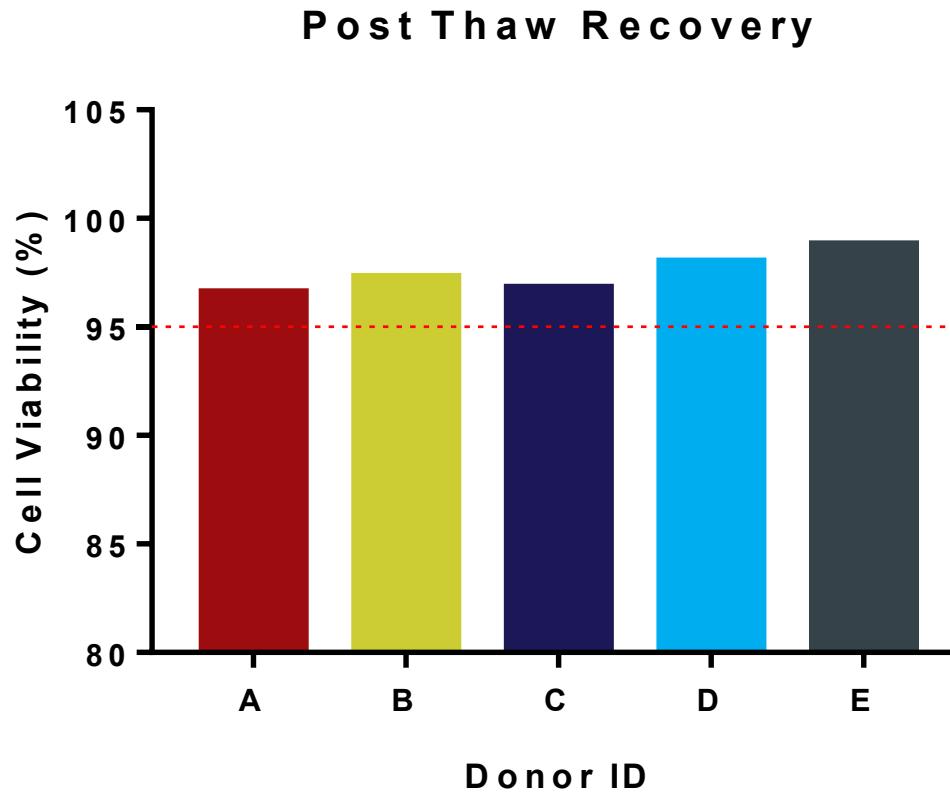
- Select lots of PBMCs can be also characterized for their functional activity in commonly used immunological assays
- We will discuss results of such characterization and discuss how the generated data can help you to select the PBMCs that are most appropriate for your experiment

# Functional assay: Donor profile

Group of genetically diverse donors

ID	Age	Gender	Ethnicity	Blood Type
Donor A	43	Male	Caucasian	A+
Donor B	51	Male	Caucasian	B-
Donor C	50	Female	Hispanic	O+
Donor D	30	Male	Hispanic	A-
Donor E	36	Male	Samoan	B+

# Functional assay: Cell viability

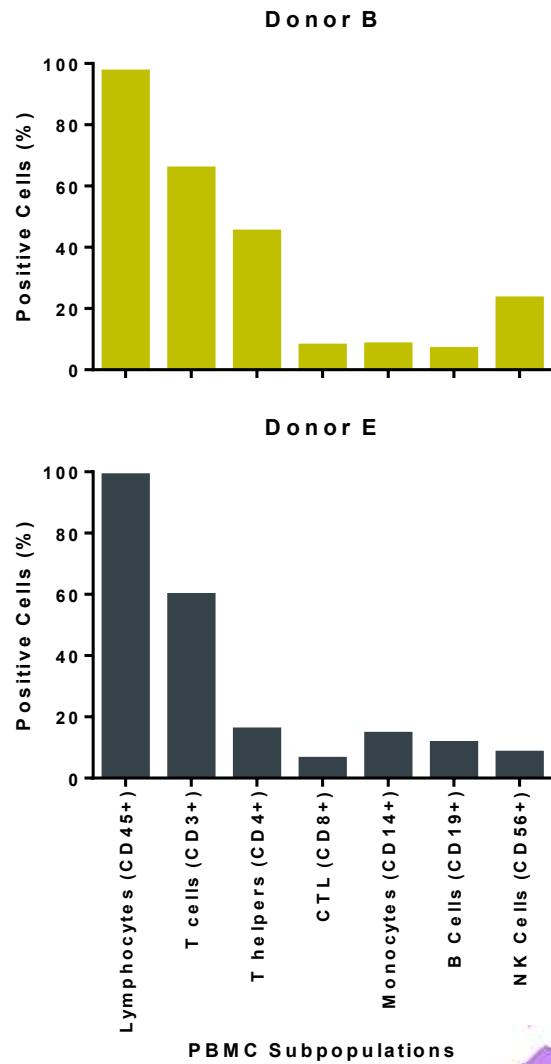
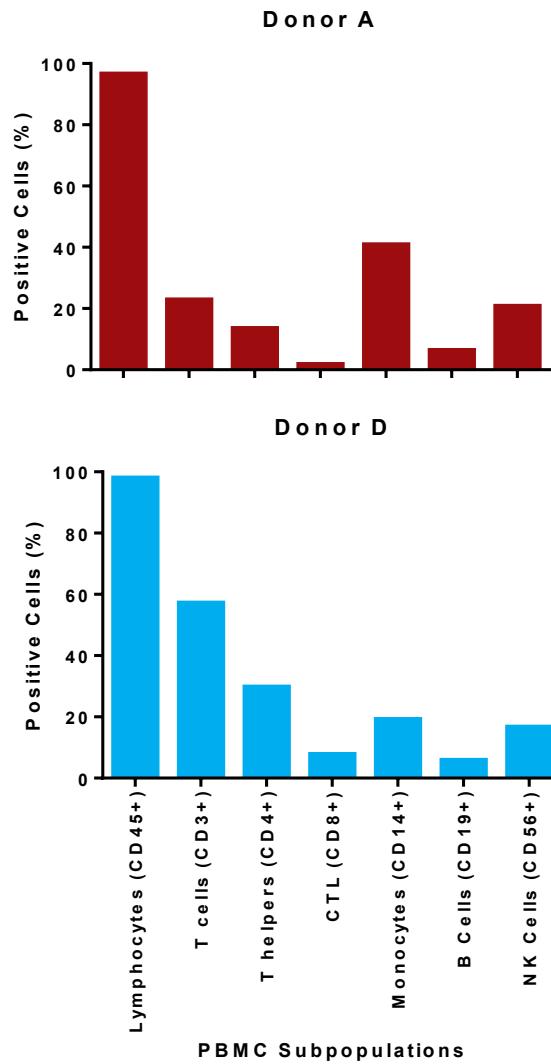
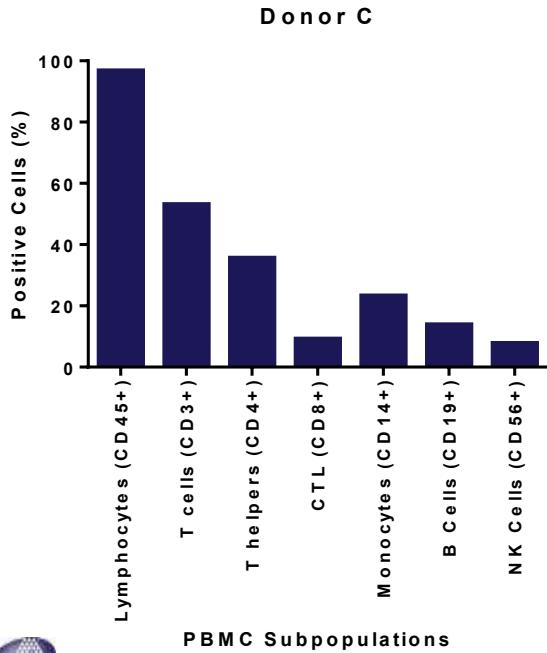


**Post-thaw viability of the tested PBMCs was greater than 95%**

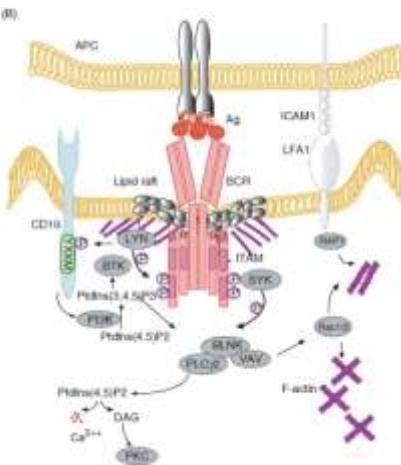
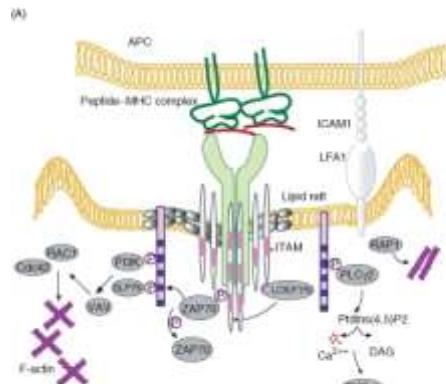
# Functional assay: Immunophenotype

All cells are tested for:

- CD45
- CD3
- CD4
- CD8
- CD14
- CD19
- CD56

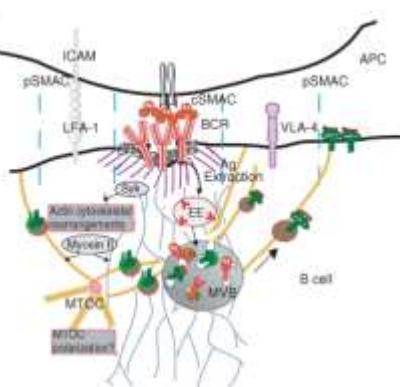
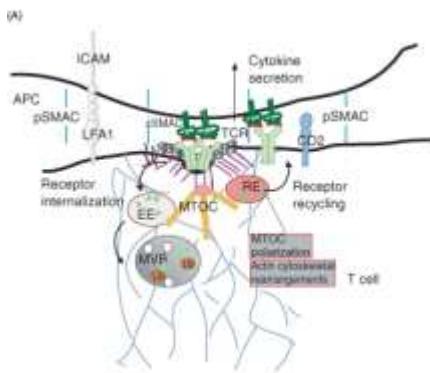


# Functional assay: Cell treatment



Cells are stimulated with commonly used and commercially available activators of immune response including:

- Soluble and bead-bound T cell specific mitogenic antibodies:
  - Anti-CD3/CD28 soluble
  - Anti-CD2/CD3/CD28 bead-bound
- Plant-derived mitogens:
  - Phytohemagglutinin (PHA)
  - Pokeweed mitogen (PWM)
- Agonists of Toll-like receptors:
  - polyIC (TLR3 agonist)
  - LPS (TLR4 agonist)
  - R-848 (agonist of TLRs 7 and 8)



Yuseff MI, et al. *Traffic* 10(6): 629-636, 2009.

# Functional assay: Assay readouts

We employed cell proliferation and cytokine expression as the primary assay readouts

T cell activation

- Proliferation
- IL-2
- IFN $\gamma$
- IL-10
- IL-13
- IL-17

Pro-inflammatory response

- TNF $\alpha$
- IL-1 $\beta$
- IL-6
- IL-12

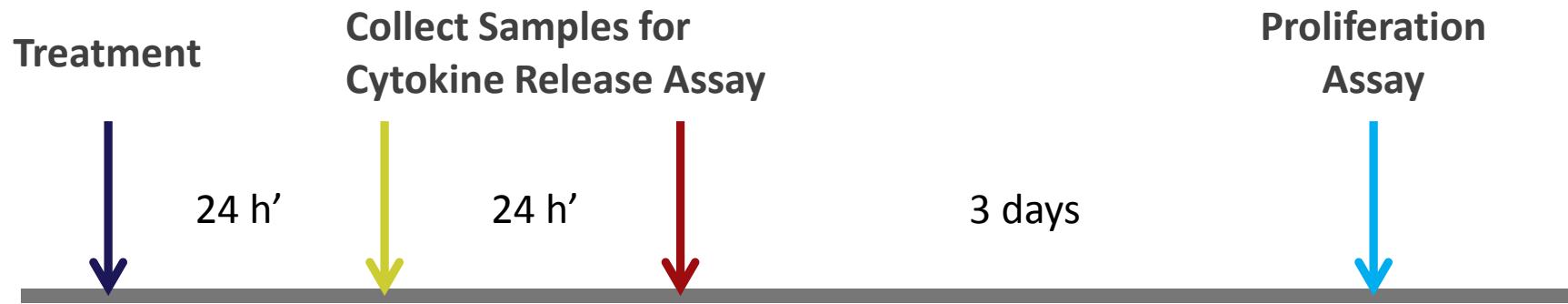
Anti-viral immunity

- IFN $\alpha$
- IP-10

The stimuli and the assay readouts were selected to demonstrate an activation of innate and adaptive immune cells within PBMCs

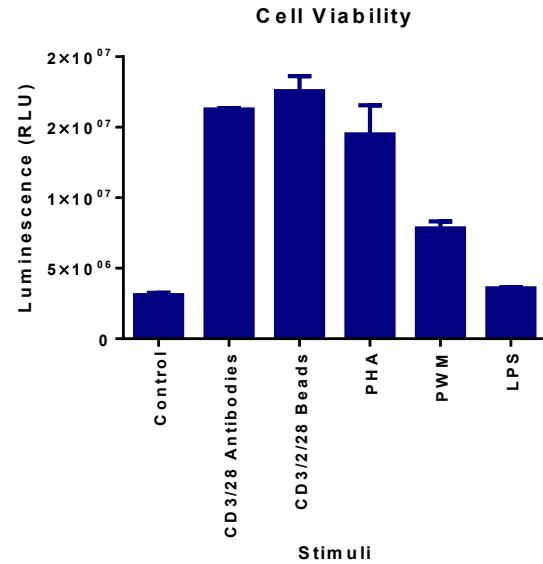
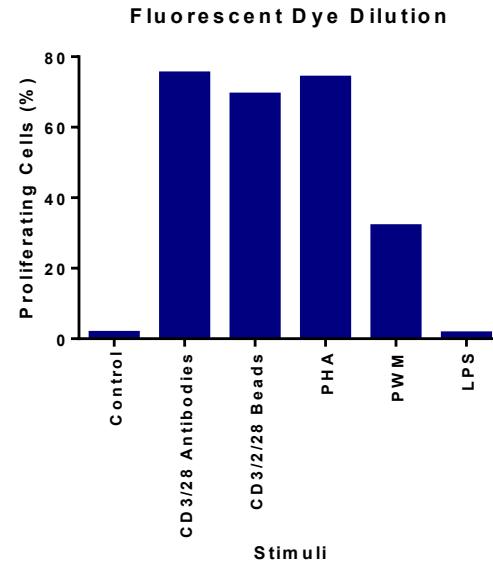
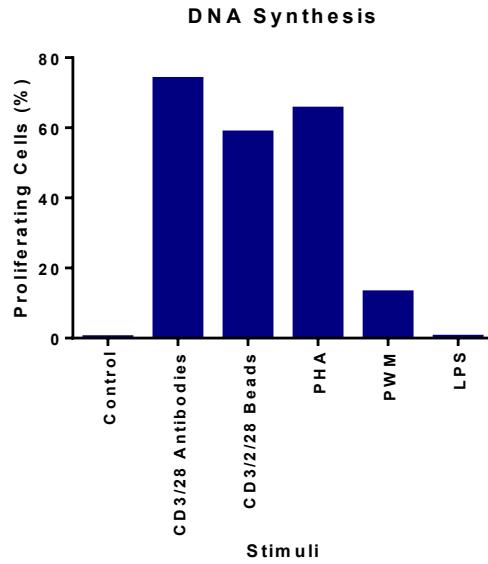
# Functional assay: Experimental design

Cells were treated and cultured in chemically defined, serum-free medium



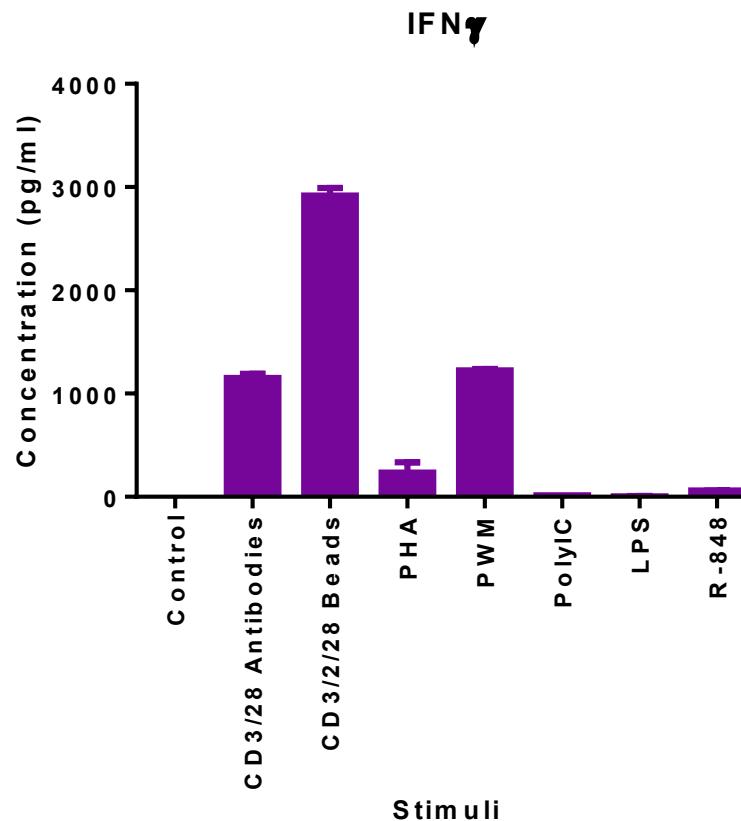
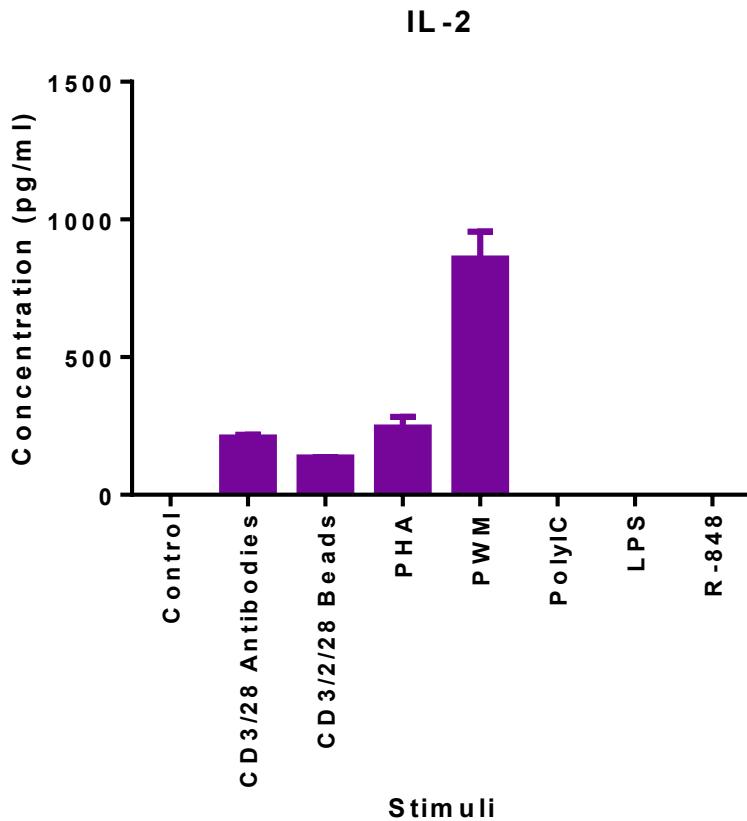
# Functional assay: Cell proliferation

Readout selection: DNA synthesis, fluorescent dye dilution, cell viability



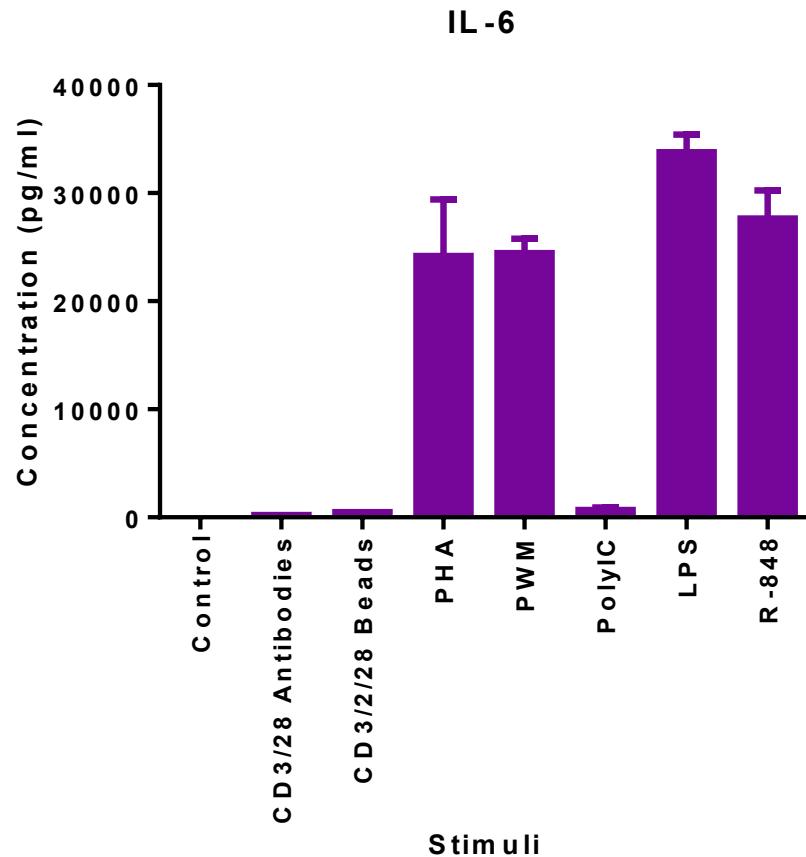
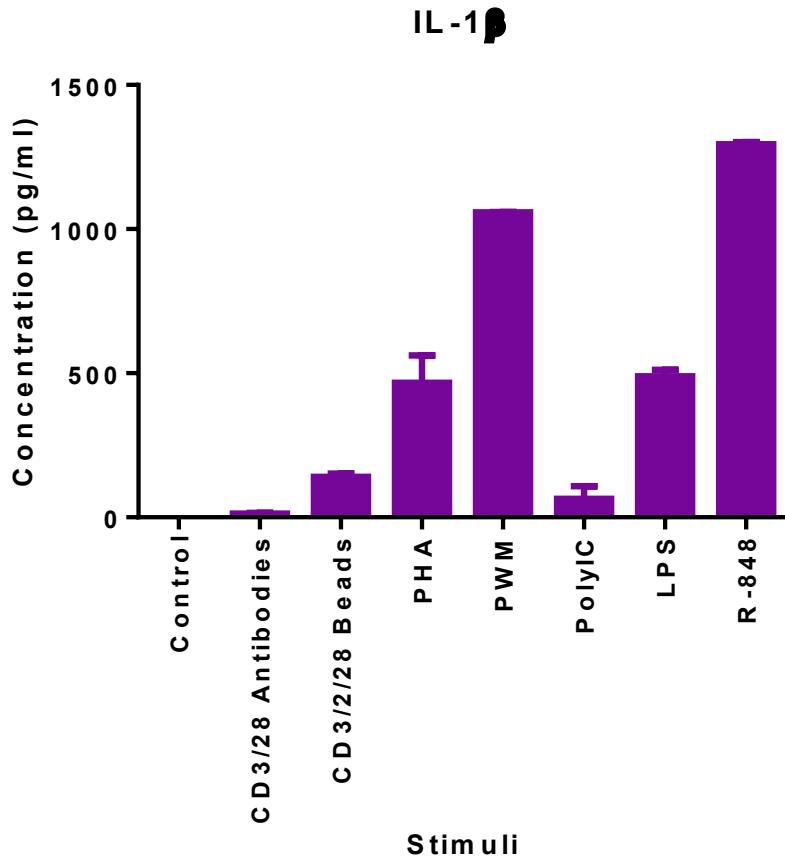
All methods provide very robust results; however, the majority of cell viability assays are faster and easier to perform

# Functional assay: Reagent validation

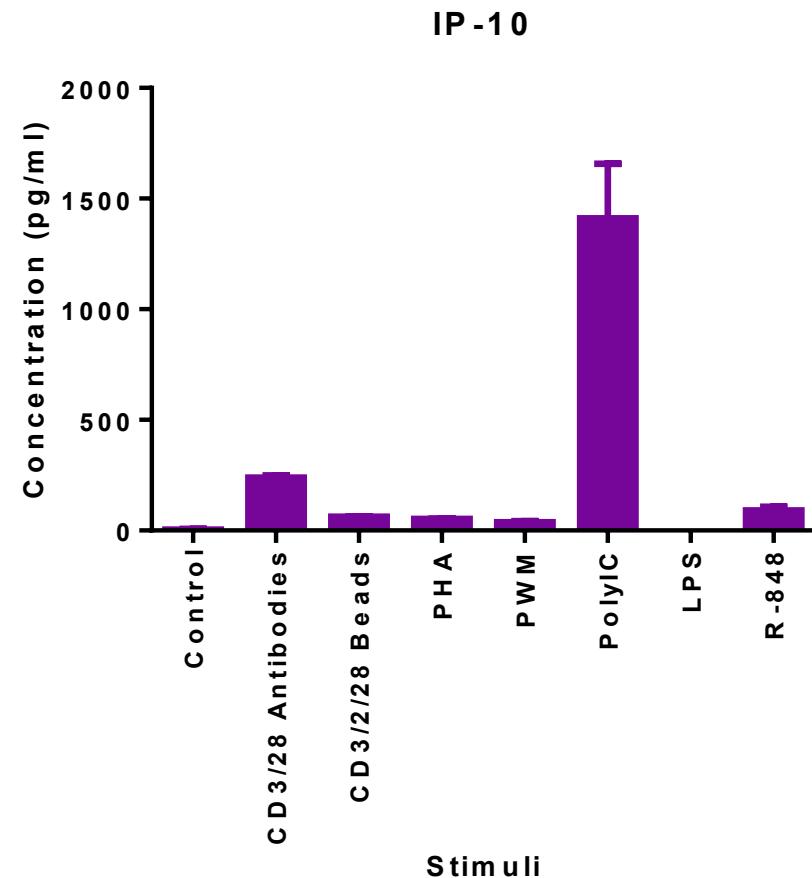
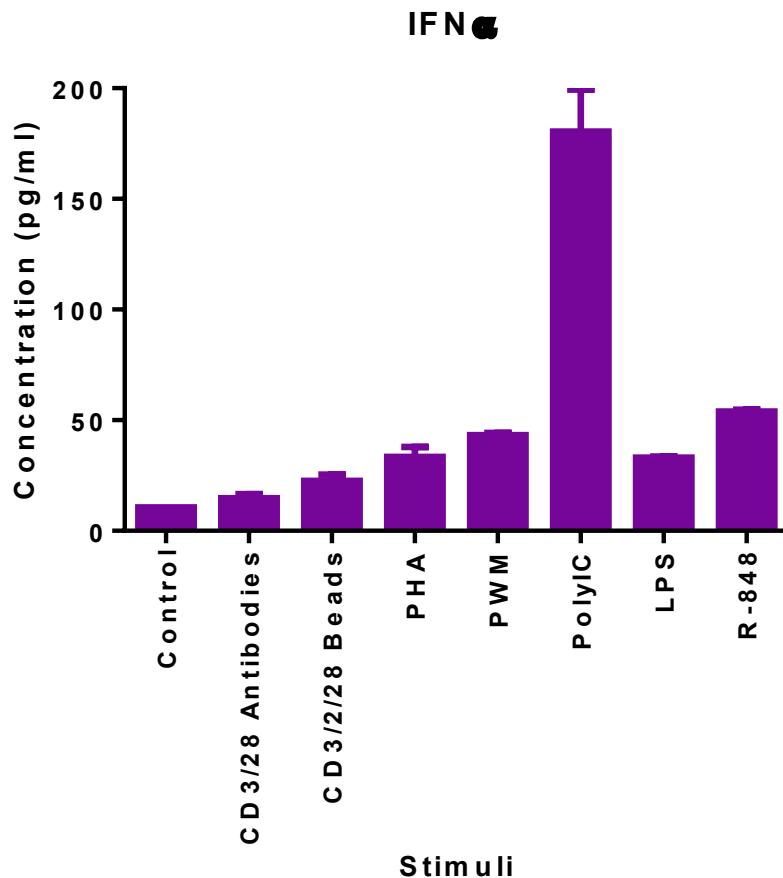


T cell dependent cytokines are induced exclusively by known T cell activators

# Functional assay: Reagent validation

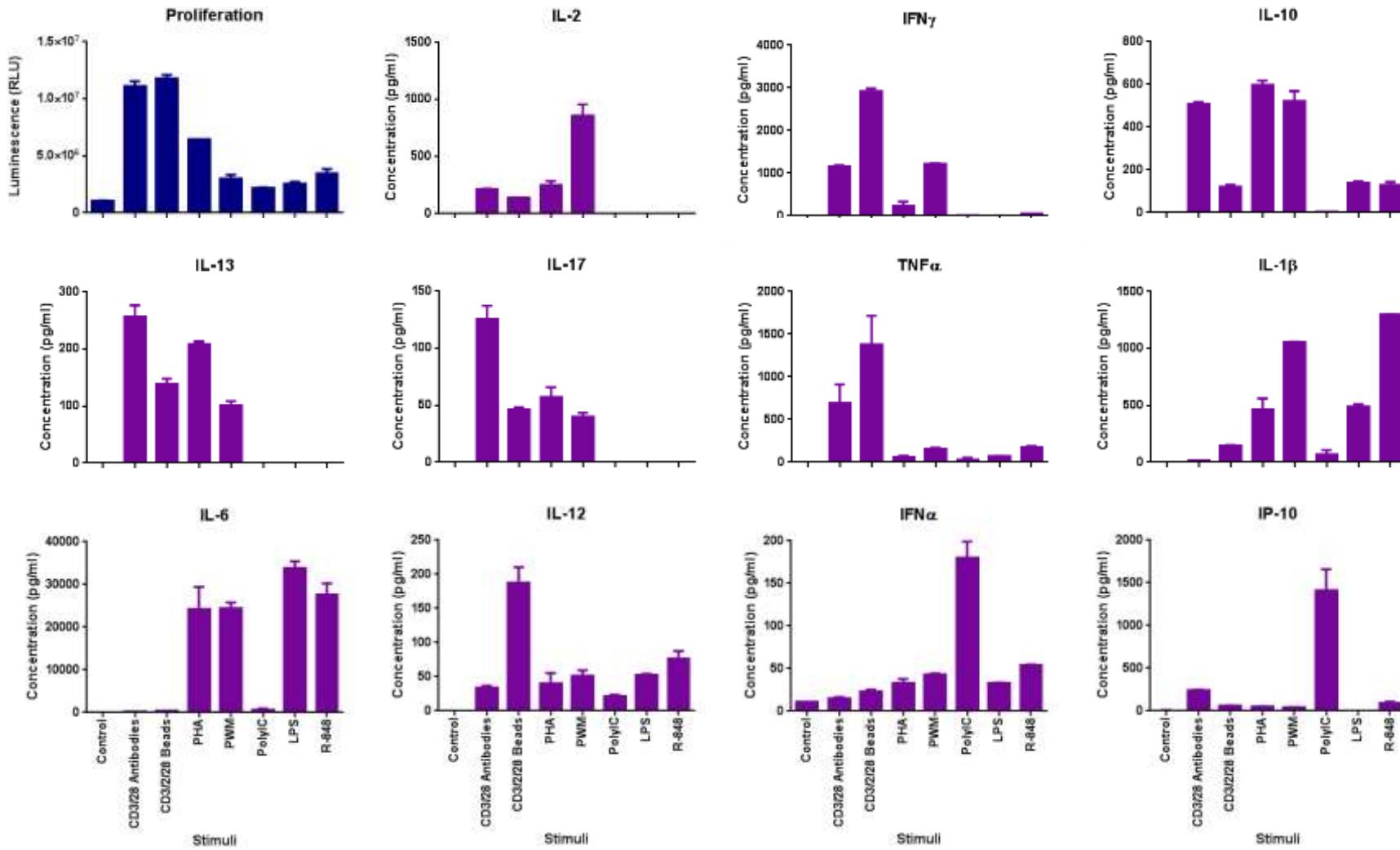


# Functional assay: Antiviral cytokines

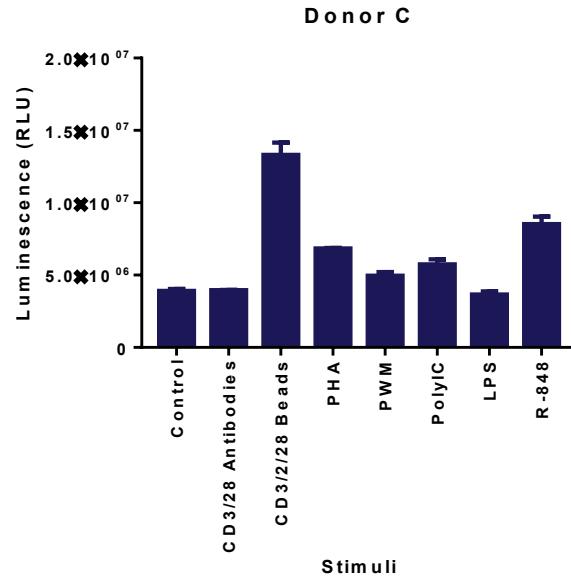
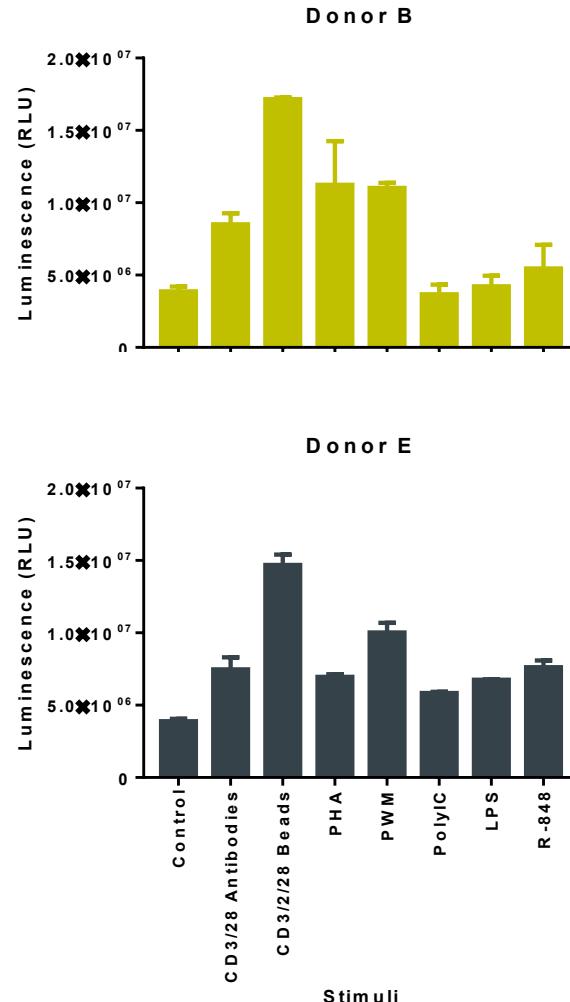
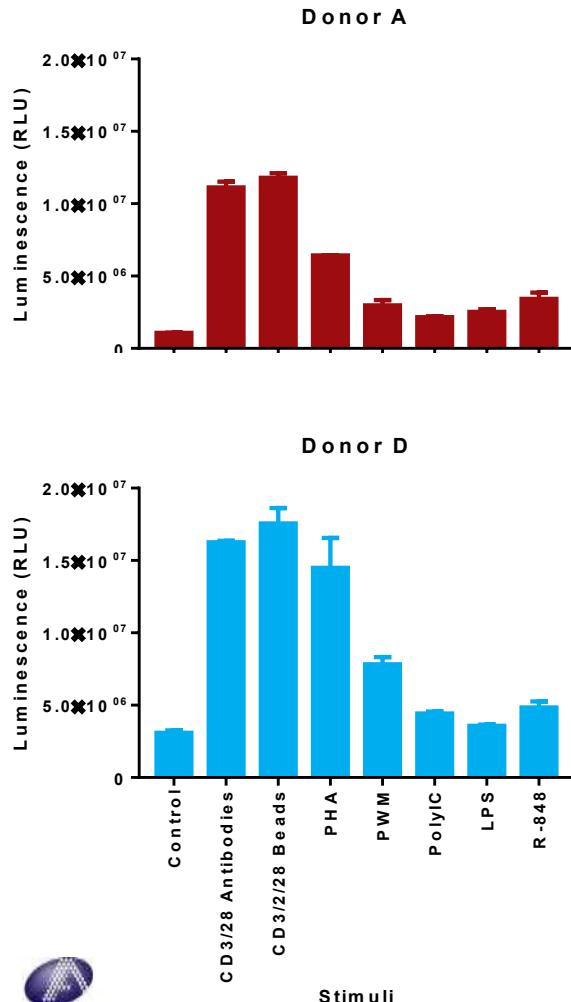


IFN $\alpha$  and IP-10 expression is induced primarily by TLR3 activation

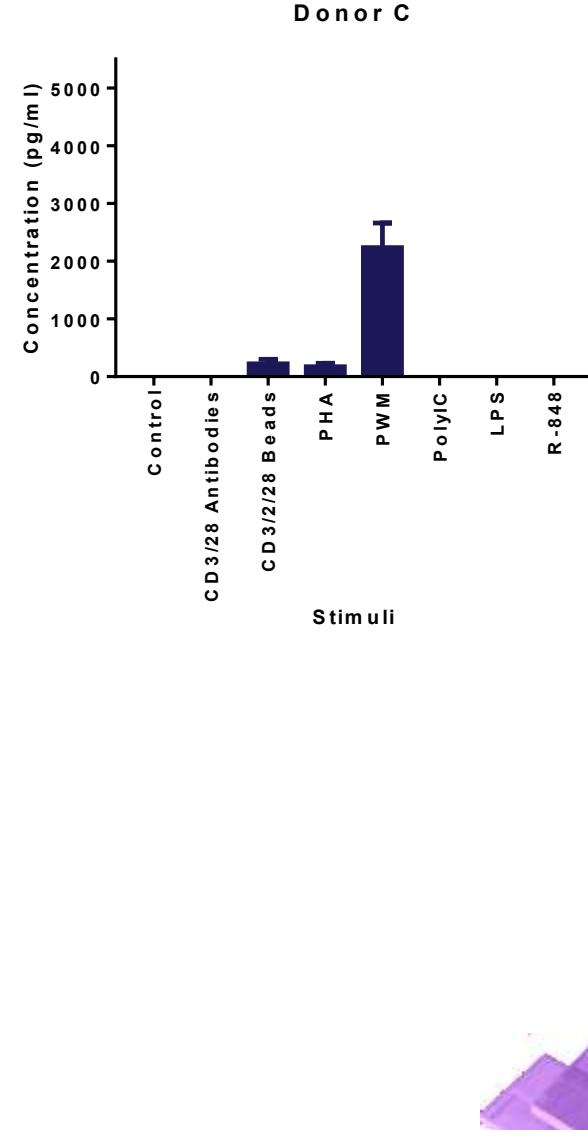
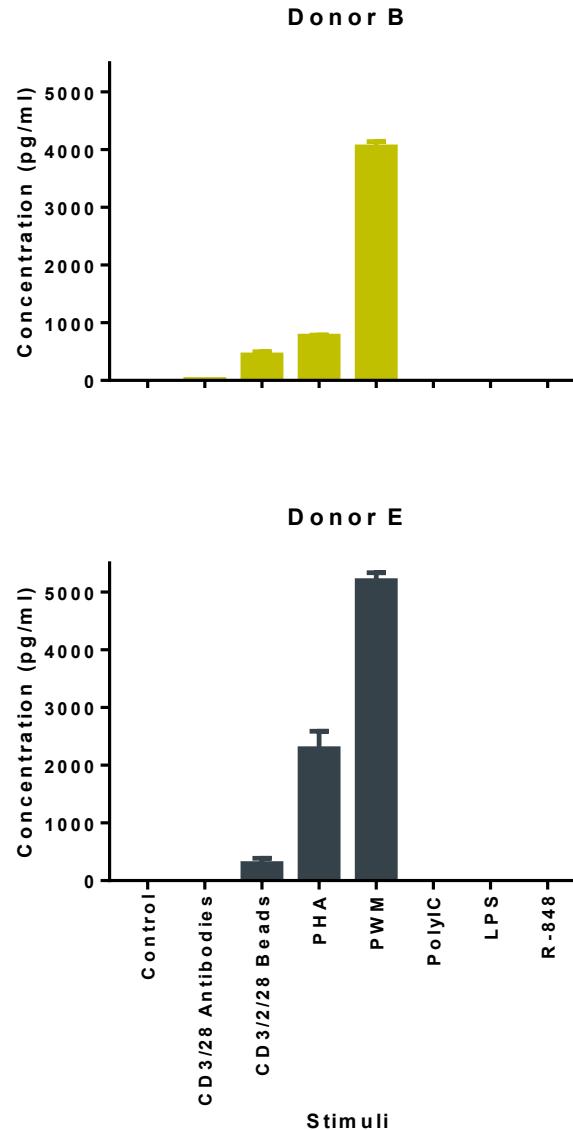
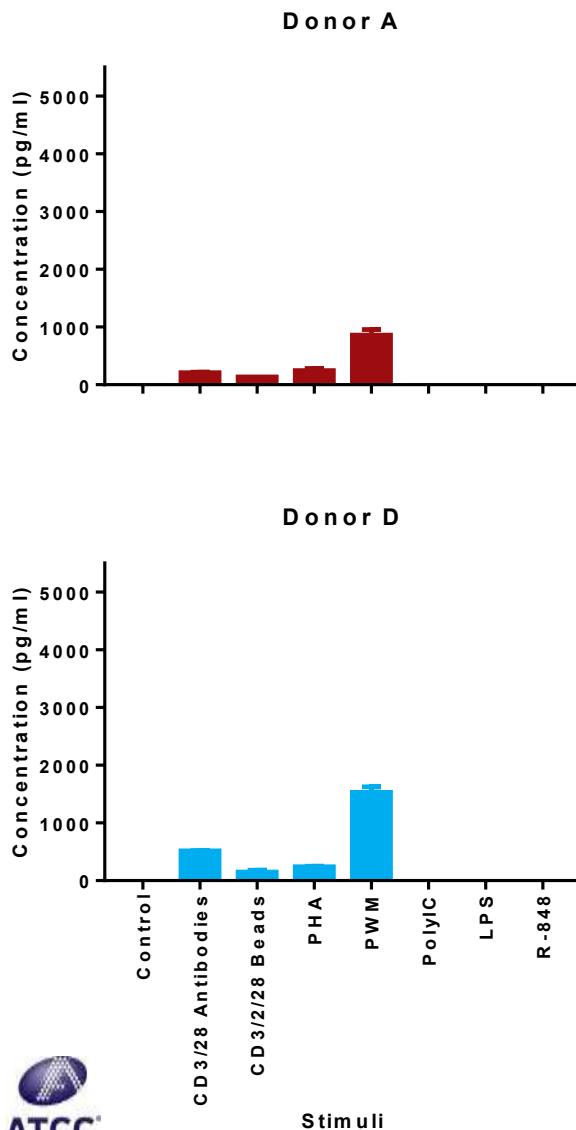
# Functional assay: Complete data set for a single donor



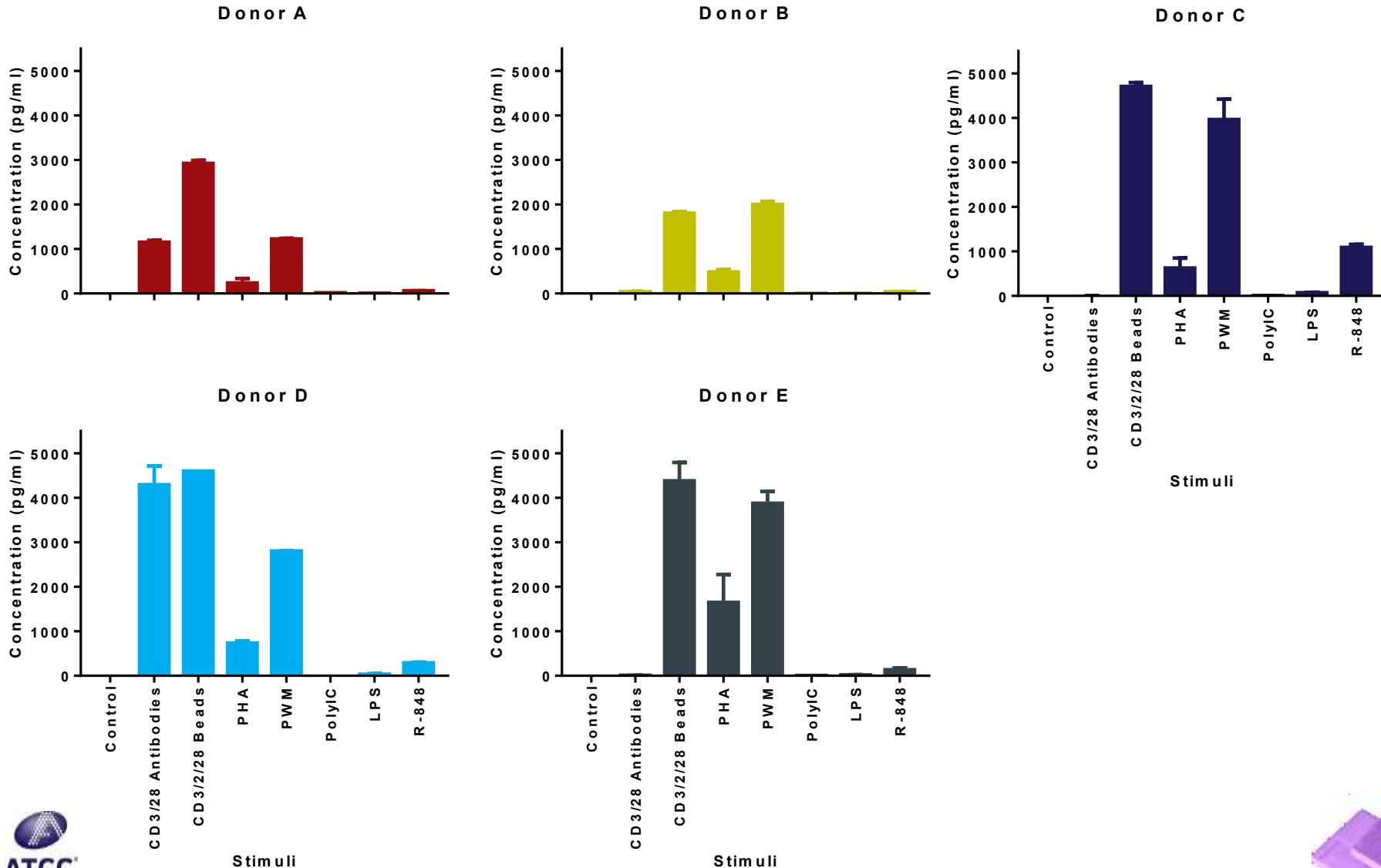
# Functional assay: Cell proliferation



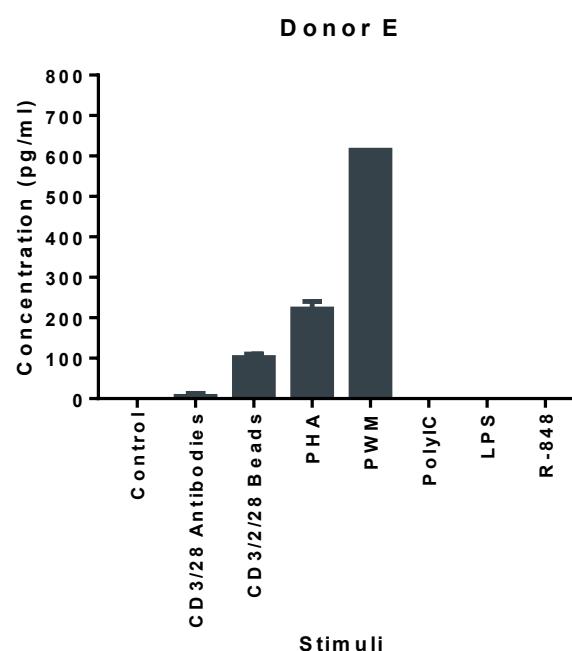
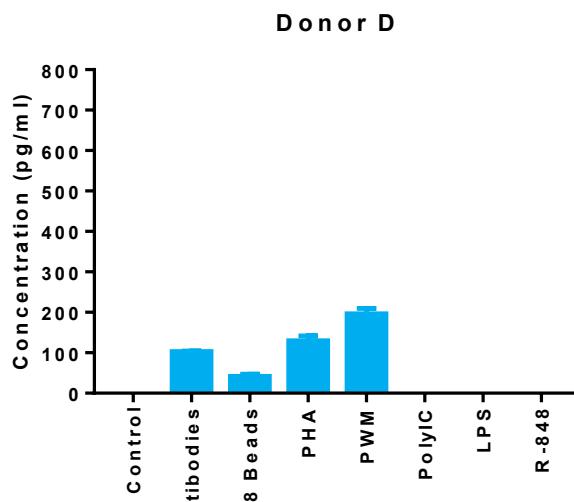
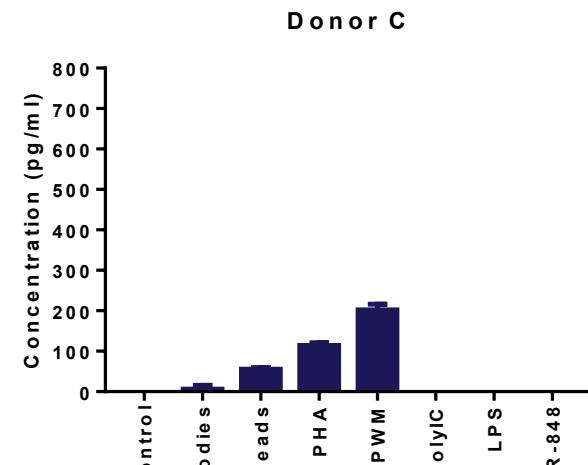
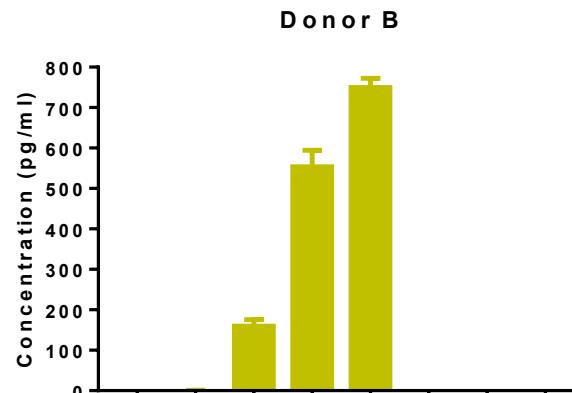
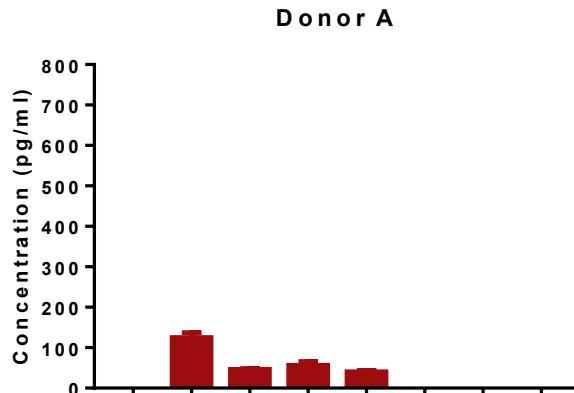
# Functional assay: IL-2 expression



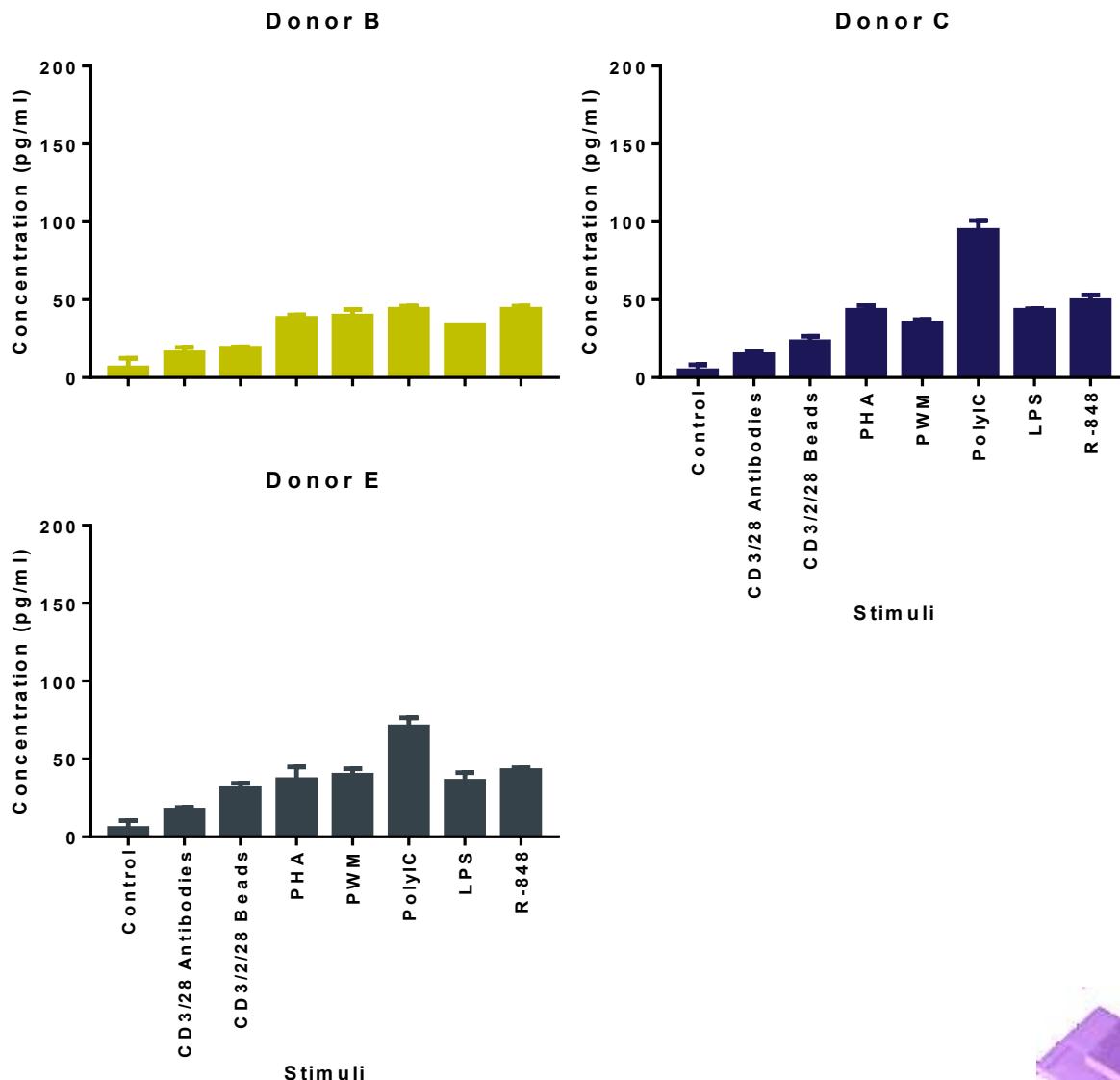
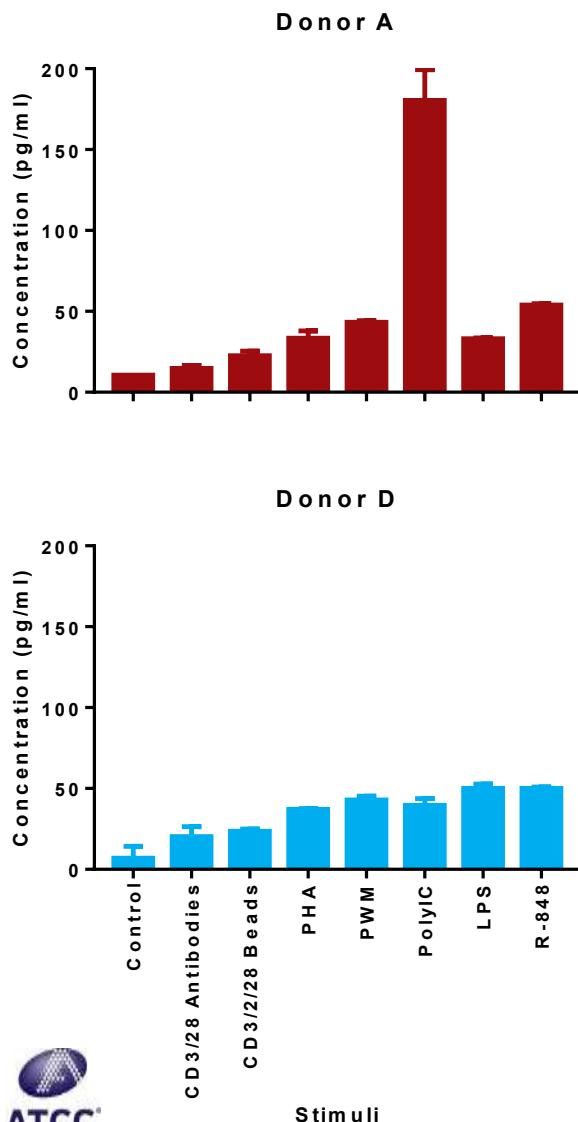
# Functional assay: IFN $\gamma$ expression



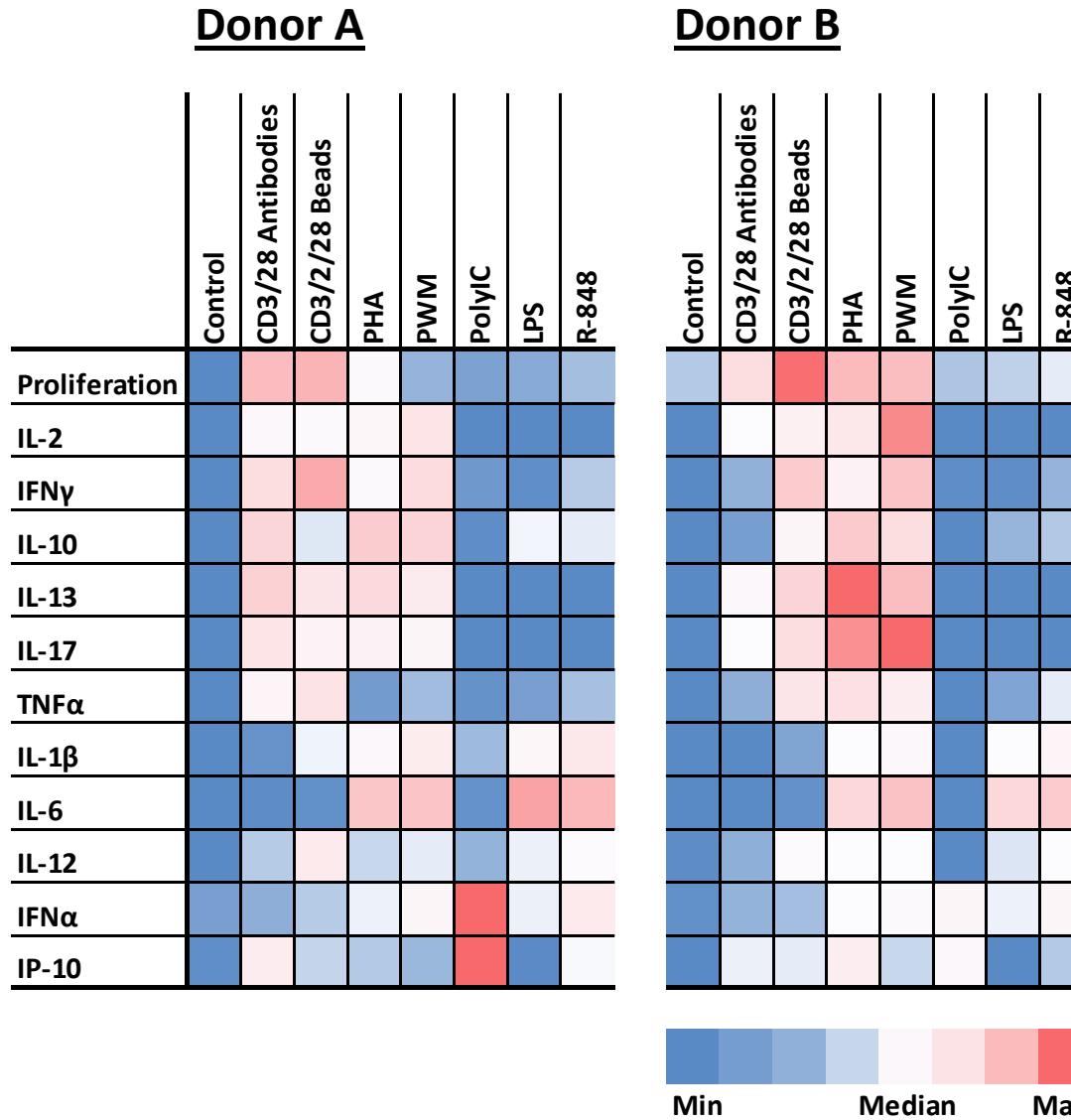
# Functional assay: IL-17 expression



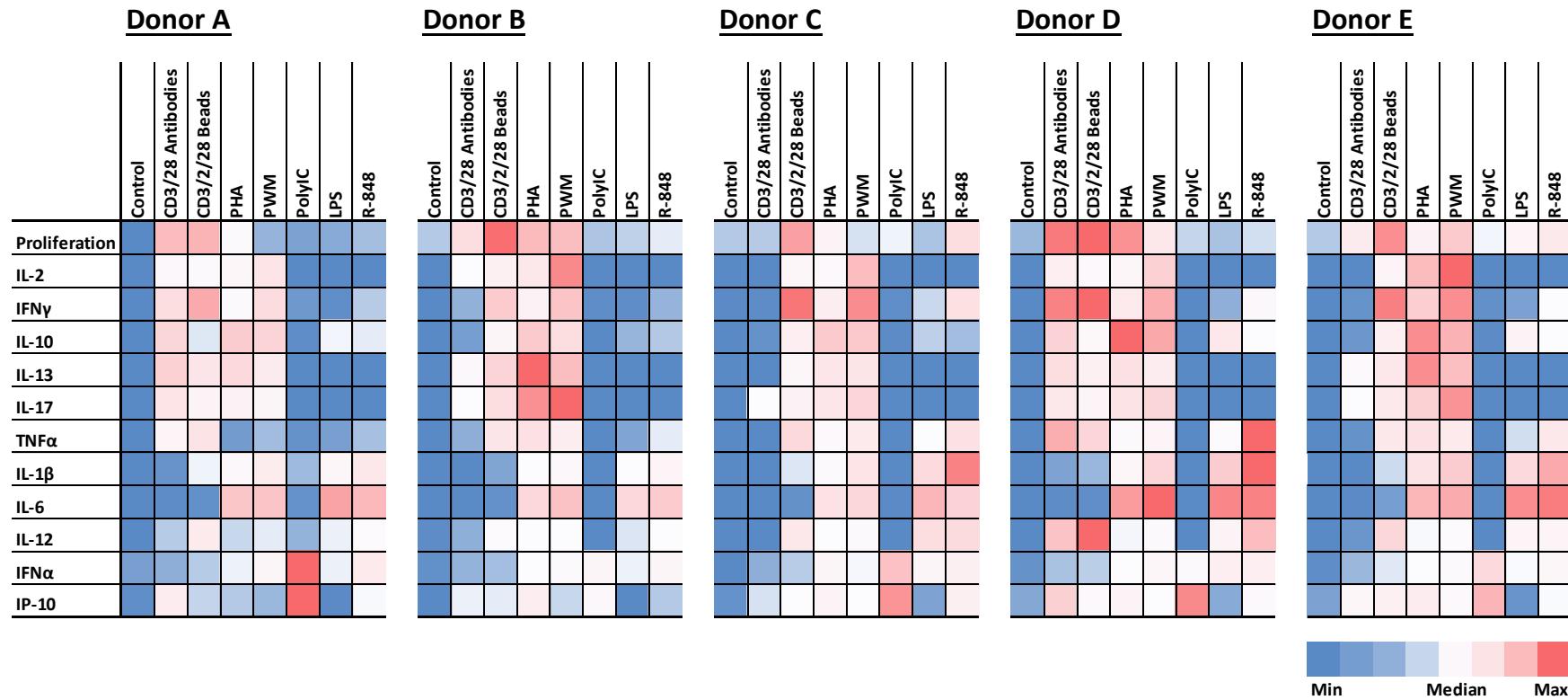
# Functional assay: IFN $\alpha$ expression



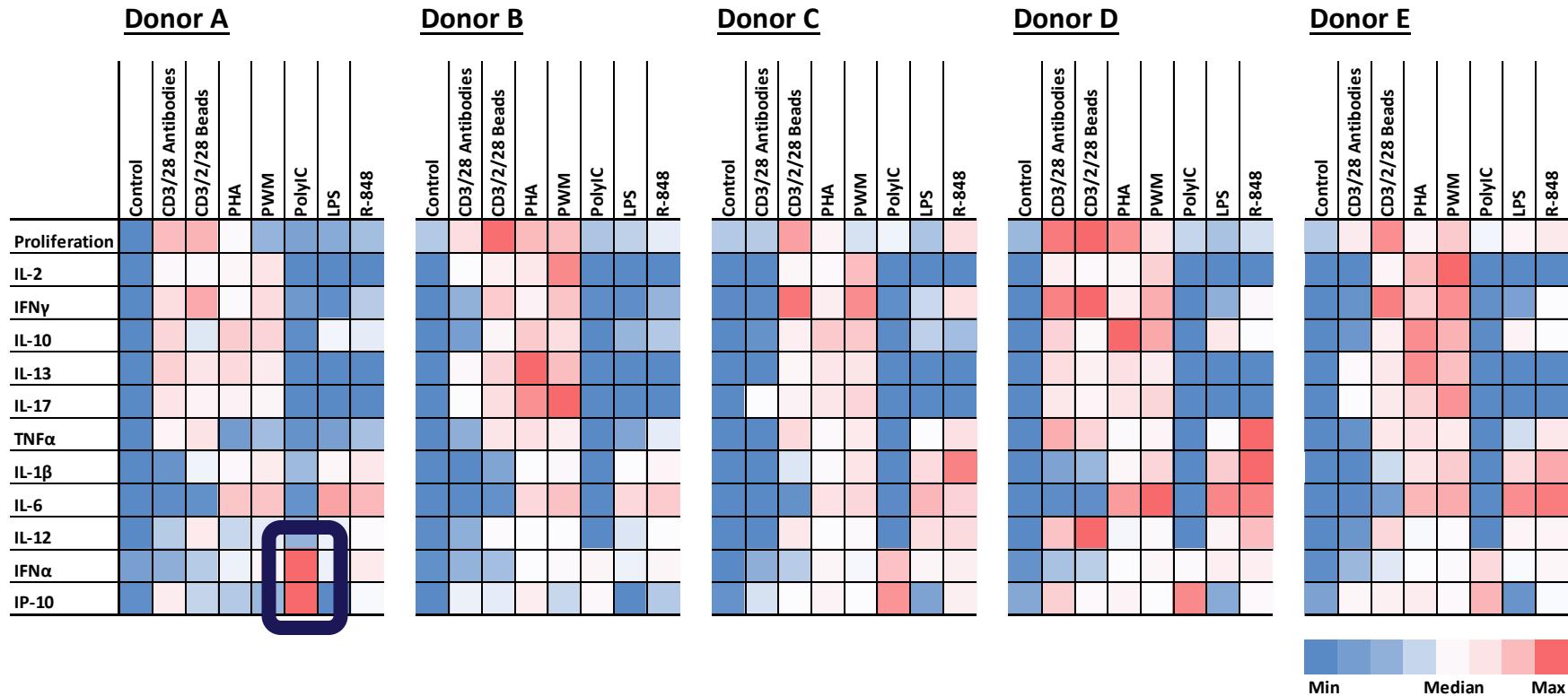
# Functional assay: Complete data set



# Functional assay: 5 donors complete data set

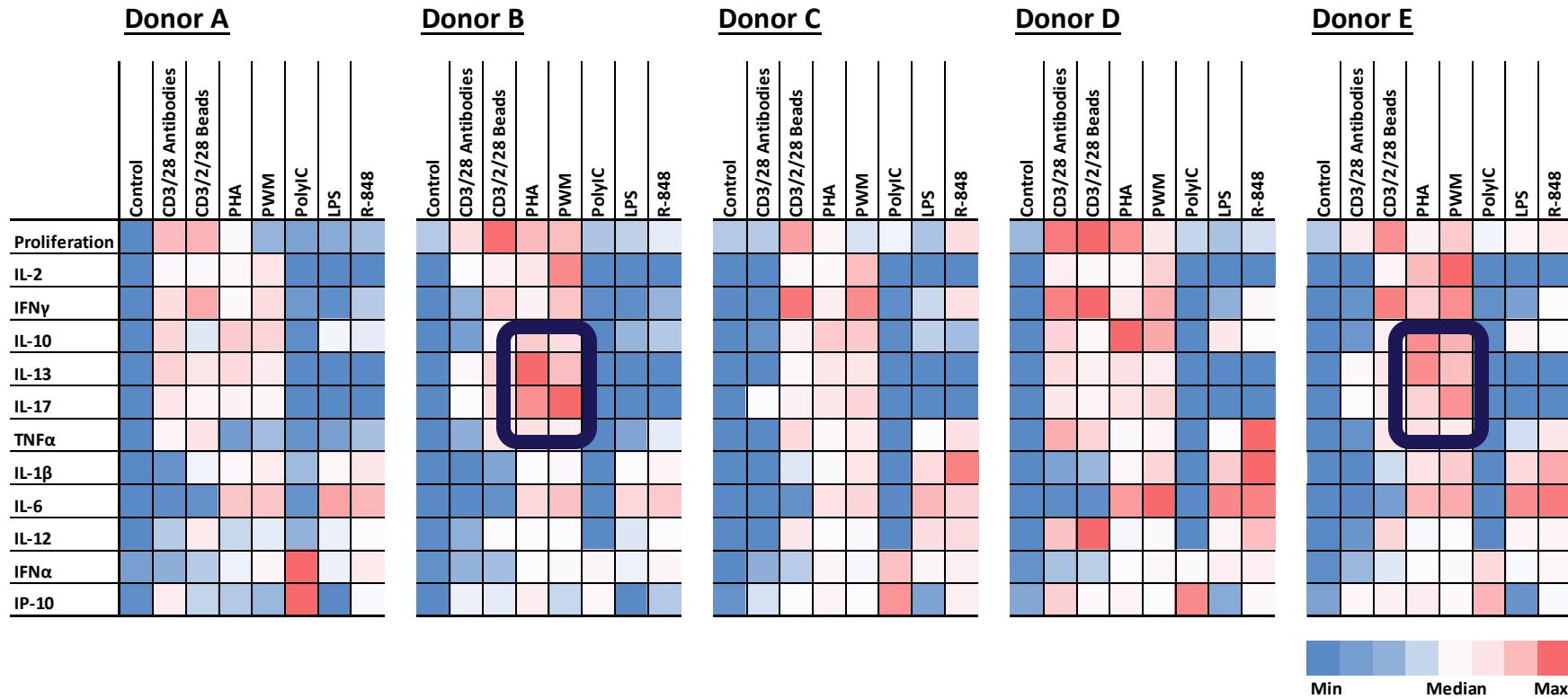


# Data driven donor selection



**Donor A is the most appropriate for investigation of IFN $\alpha$  expression**

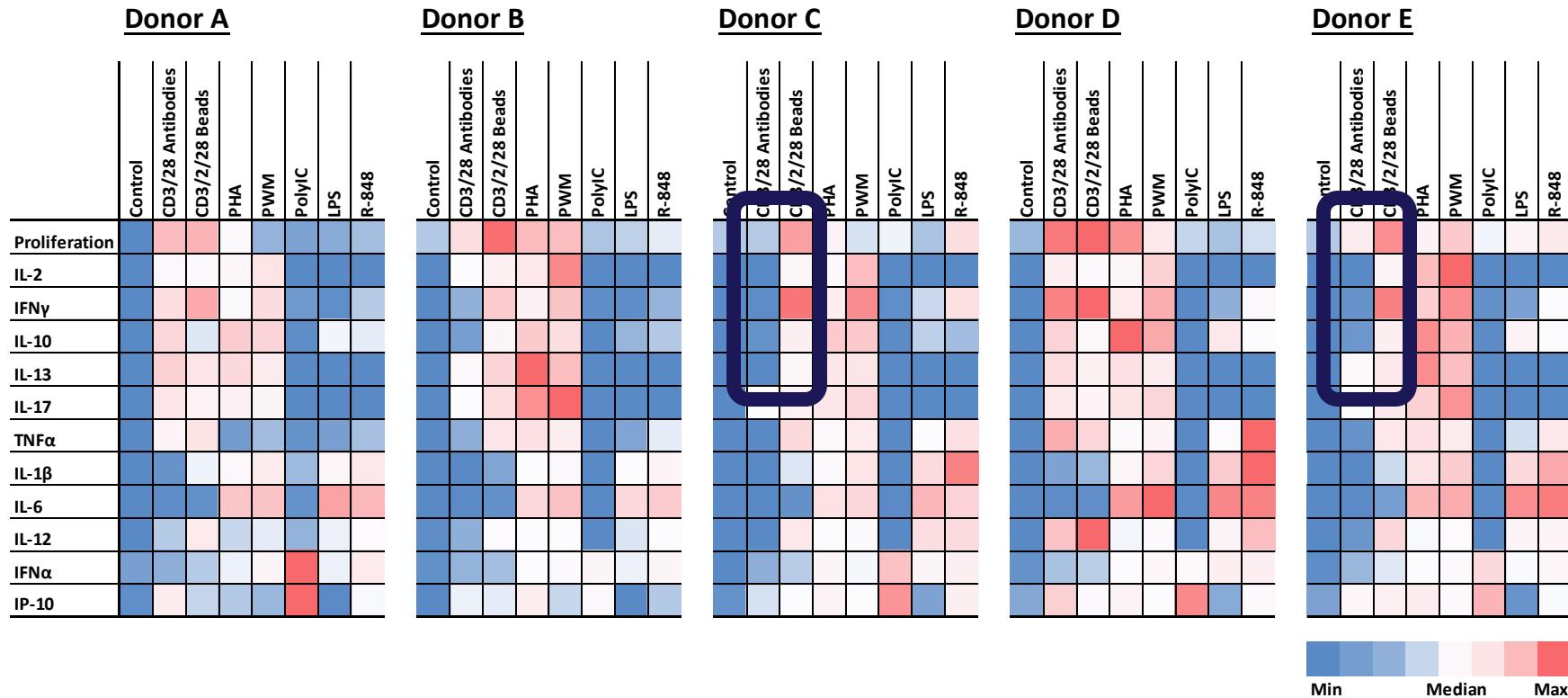
# Data driven donor selection



**Donors B and E are more suitable for investigation of IL-13 and IL-17 expression**



# Data driven donor selection



There is significant variability in cell responsiveness to commonly used stimuli

# Conclusion

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- 1. Functional activity of human PBMC *in vitro* is extremely variable**
- 2. Factors contributing to this variability can be divided into two groups:**

## I. Can be controlled

- Cryopreservation technique
- Cell viability
- Medium composition
- Quality of cell activating reagents

## II. Have to be accepted (Uncontrollable)

- Genetic diversity
- Environmental factors
  - Immunizations
  - Nutrition
  - Latent infections

- 3. To minimize assay variability we recommend to:**

- Use serum- or protein-free cryopreservation solution
- Ensure high viability of cells at the time of experiment
- Use prequalified FBS, autologous plasma/serum, or serum-free medium
- Validate cell activating (control) reagents

# Conclusion (cont'd)

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**4. Use of functionally characterized cells can significantly minimize assay variability and improve reproducibility of experimental results**

**5. When selecting a specific lot of characterized cells one should consider:**

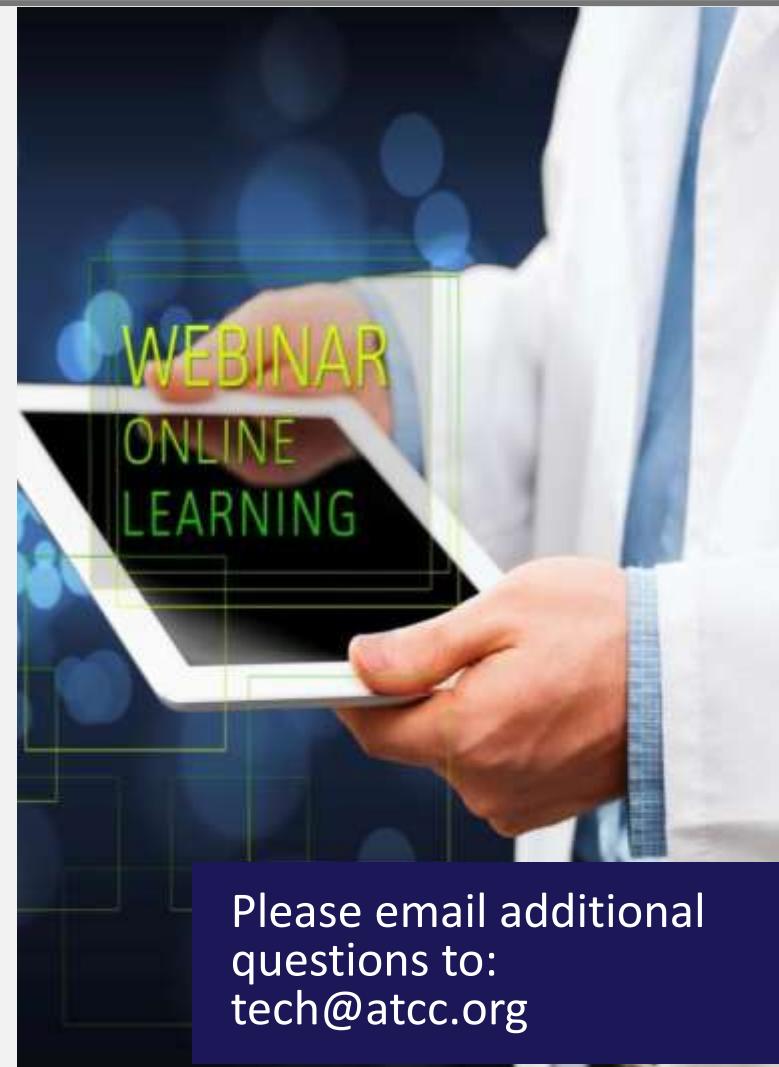
- Assay readout
- Cell reactivity (ability to respond to a specific stimuli)
- Magnitude of the response

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James Clinton, Ph.D., *Scientist, ATCC*  
Discovering ATCC Hematopoietic Progenitor Cells  
– Model Systems to Study the Immune and  
Cardiovascular Systems



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