#### SEEING IS BELIEVING - REPORTER LABELED MICROBIAL CONTROL STRAINS

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THE ESSENTIALS OF LIFE SCIENCE RESEARCH GLOBALLY DELIVERED\*

#### **Today's discussion**







Reporter labels and their use



**GFP-labeled** strains



NanoLuc<sup>®</sup>-labeled Shiga toxin-producing *Escherichia coli* strains



# About ATCC

- Founded in 1925, ATCC is a non-profit organization with headquarters in Manassas, VA
- World's premiere biological materials resource and standards development organization
- ATCC collaborates with and supports the scientific community with industry-standard products and innovative solutions
- Strong team of 400+ employees; over one third with advanced degrees
- Broad range of biomaterials
  - Cell lines, iPSCs, primary cells, and hTERT immortalized cells
  - Bacteria, yeasts, protists, and viruses
  - Tumor cell panels
  - Media, sera, and reagents





# Why label bacteria?

- Visualize bacteria under experimental conditions
- Differentiate between sample contamination and false positives from control cross-contamination
  - Saves money, time, and worry





## Why label bacteria?



Microbial quantification and detection



Host-pathogen interactions



Drug discovery



#### Food testing

Gene expression

**Biochemical interactions** 







Quality control assays

### **Reporter characteristics**

	Green Fluorescent Protein (GFP)	NanoLuc <sup>®</sup> Luciferase
Live-cell assays	Yes	No
Well-characterized	Yes	No
Highly stable	Yes	Yes
Substrate required	No	Yes
Glow response	Yes	Yes
Use in vivo	Yes	No



# **GFP-labeled strains**

- Labeled Gram-negative organisms with GFPmut3a
  - Escherichia coli
  - Salmonella enterica subsp. enterica serovar Typhimurium
  - Shigella flexneri
  - Pseudomonas aeruginosa
- *gfp* expressed behind the P<sub>lac</sub> constitutive promoter
- Bright, consistent label can be used for:
  - Microbial quantification and detection
  - Host-pathogen interactions
  - Drug discovery
  - Food testing





#### **Visual detection of GFP-labeled organisms**





The expression of a bright GFP variant on a high-copy number plasmid facilitates visual identification when exposed to UV light (A) or imaged using a detection system (B &C)

### **Bacterial fitness and quantification**



• GFPmut3a is not detrimental to bacterial fitness

• Fluorescence can be used for bacterial quantification



# **Plasmid stability**



Plasmid is stable for at least 20 generations at 37°C in the absence of antibiotic pressure



## **Photo bleaching**











- P. aeruginosa-GFP (ATCC<sup>®</sup> 10145GFP<sup>™</sup>) was continuously exposed to UV light to determine resistance to photo bleaching
- Fluorescence remained stable during the first 5 minutes
- Significant loss of signal after 7 minutes of exposure





# Fluorescent microscopy detection of pathogen-host interactions





*P. aeruginosa* GFP (ATCC<sup>®</sup>10145GFP<sup>™</sup>) interaction with A549 (ATCC<sup>®</sup> CCL-185<sup>™</sup>) airway epithelial cells (100X magnification)

# High-throughput detection of pathogen-host interactions

#### Invasion study

- P. aeruginosa GFP (ATCC<sup>®</sup>10145GFP<sup>™</sup>) was incubated in the presence of A549 (ATCC<sup>®</sup> CCL-185<sup>™</sup>) airway epithelial cell monolayers
- Cells were washed and medium supplemented with 100 µL gentamicin was added to kill extracellular bacteria (Adhesion)
- Cells were washed again (Invasion) and measured on a microplate reader





# High-throughput detection of pathogen-host interactions



#### **Flow Cytometry**

P. aeruginosa (ATCC<sup>®</sup> 10145<sup>™</sup>) (purple) and P. aeruginosa GFP (ATCC<sup>®</sup> 10145GFP <sup>™</sup>) (green) suspensions were analyzed by flow cytometry



# High-throughput detection of pathogen-host interactions



#### Macrophage uptake of Pseudomonas aeruginosa

P. aeruginosa (ATCC<sup>®</sup> 10145<sup>™</sup>) and P. aeruginosa GFP (ATCC<sup>®</sup> 10145GFP<sup>™</sup>) were incubated in the presence or absence of Cytochalasin D and analyzed by flow cytometry

#### In vivo detection of GFP-labeled P. aeruginosa



10<sup>8</sup> CFU 10<sup>6</sup> CFU 10<sup>4</sup> CFU Blank

- Various doses of *P. aeruginosa* GFP (ATCC<sup>®</sup> 10145GFP<sup>™</sup>) were injected into the mid-rib of *Lactuca sativa* L. var. *longifolia*
- The bacteria was easily detected at higher concentrations in the plant host, indicating that this vector can be successfully employed to monitor bacterial growth within a plant host



#### **ATCC GFP-labeled strains**

ATCC <sup>®</sup> No.	Species	Reporter	Parental Strain (ATCC <sup>®</sup> No.)
25922GFP™	Escherichia coli	GFP	25922™
14028GFP™	Salmonella enterica subsp. enterica serovar Typhimurium	GFP	14028™
12022GFP™	Shigella flexneri	GFP	12022™
10145GFP™	Pseudomonas aeruginosa	GFP	10145™
15692GFP™	Pseudomonas aeruginosa	GFP	15692™

www.atcc.org/reporters



### **Promega NanoLuc<sup>®</sup> reporter**



**Cotton Swab** 

**Filter Paper** 

**Cell Pellet** 

- Intensely bright reporter
- Glow response
- Portable, does not require instrumentation



# Shiga toxin-producing *E. coli*

- >265,000 cases of STEC infection in the United States each year
  - E. coli O157:H7 accounts for about 36% of STEC infections
  - ~5-10% of diagnosed infections develop into hemolytic uremic syndrome, a life threatening complication which can cause permanent health damage
- Food Safety and Modernization Act calls for expanded testing to include Non-O157 strains
  - O26 O111
  - O45 O121
  - O103 O145





## **Microbial strain authentication**



ATCC utilizes both classical and modern techniques

- Phenotypic analysis
- Genotypic & proteotypic analyses
- Functional analysis

No single method of identification is sufficient



### **Phenotypic testing**





## **Genotypic & proteotypic testing**





### **Functional testing**





# **STEC Testing**

- Bacterial identification
  - VITEK<sup>®</sup> 2, VITEK<sup>®</sup> MS, API<sup>®</sup> Strips, 16S rRNA sequencing
- Molecular characterization to assess the presence of the stx1, stx2, and eaeA genes
  - PCR
- Serogroup identification
  - Immunoprecipitation assay





#### Minimal reporter effects on bacterial fitness





#### Plasmid stable for ≥2 days



Observe reporter in >65% of c.f.u. after 2 days at 42°C



# Shiga toxin-producing *E. coli* (STEC) control strains

Labeled controls rule out cross-contamination of samples with the control strain





#### **NanoLuc®-labeled control detection**

#### **Cotton Swab**





Broth



External testing confirmed a clear signal after FSIS protocol for ground beef



### NanoLuc<sup>®</sup>-labeled *Escherichia coli*

ATCC <sup>®</sup> No.	Species	Serotype	Genotype
BAA-2580-PACK™	Escherichia coli	O26:H11	<i>stx</i> 1+, <i>stx</i> 2+, <i>eae</i> A+
BAA-2581-PACK™	Escherichia coli	O45:H2	<i>stx</i> 1+, <i>stx</i> 2-, <i>eae</i> A+
BAA-2582-PACK™	Escherichia coli	O103:H11	<i>stx</i> 1+, <i>stx</i> 2-, <i>eae</i> A+
BAA-2583-PACK™	Escherichia coli	O111	<i>stx</i> 1+, <i>stx</i> 2-, <i>eae</i> A+
BAA-2584-PACK™	Escherichia coli	O121:H19	<i>stx</i> 1-, <i>stx</i> 2+, <i>eae</i> A+
BAA-2585-PACK™	Escherichia coli	O145	<i>stx</i> 1-, <i>stx</i> 2+, <i>eae</i> A+
BAA-2586-PACK™	Escherichia coli	O157:H7	<i>stx</i> 1+, <i>stx</i> 2+, <i>eae</i> A+
BAA-2587-PACK™	Escherichia coli	O157:H7	<i>stx</i> 1-, <i>stx</i> 2-, <i>eae</i> A-

www.atcc.org/reporters

*Coming soon* – Big-Six *Escherichia coli* NanoLuc<sup>®</sup> Strains Panel



#### **Additional testing resources**





# Conclusions

- Reporter labels are a flexible research tool
- Using labels for microbial detection enhances safety and reliability in food testing
- ATCC strains provide
  - Authenticated reference standards
  - Low Passage
  - Significant savings in time and effort

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Thank you for joining today! Please send additional questions to <u>tech@atcc.org</u>

