



ENHANCING VECTOR-BORNE RESEARCH WITH BIOLOGICAL AND MOLECULAR STANDARDS

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Lead Biologist, ATCC
March 5, 2015



THE ESSENTIALS OF LIFE SCIENCE RESEARCH
GLOBALLY DELIVERED™

About ATCC

- Founded in 1925, ATCC is a non-profit organization with headquarters in Manassas, VA
- World's premiere biological materials resource and standards development organization
- ATCC collaborates with and supports the scientific community with industry-standard products and innovative solutions
- Broad range of biomaterials
 - Continuous cell lines, iPSCs, primary cells, and hTERT immortalized cells
 - Bacteria, fungi, yeasts, protists, and viruses
 - Microbial and tumor cell panels
 - Genomic and synthetic nucleic acids
 - Media, sera, and reagents



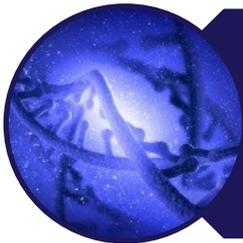
Outline



Introduction and clinical significance



Prevention and treatment



Solutions for vector-borne research

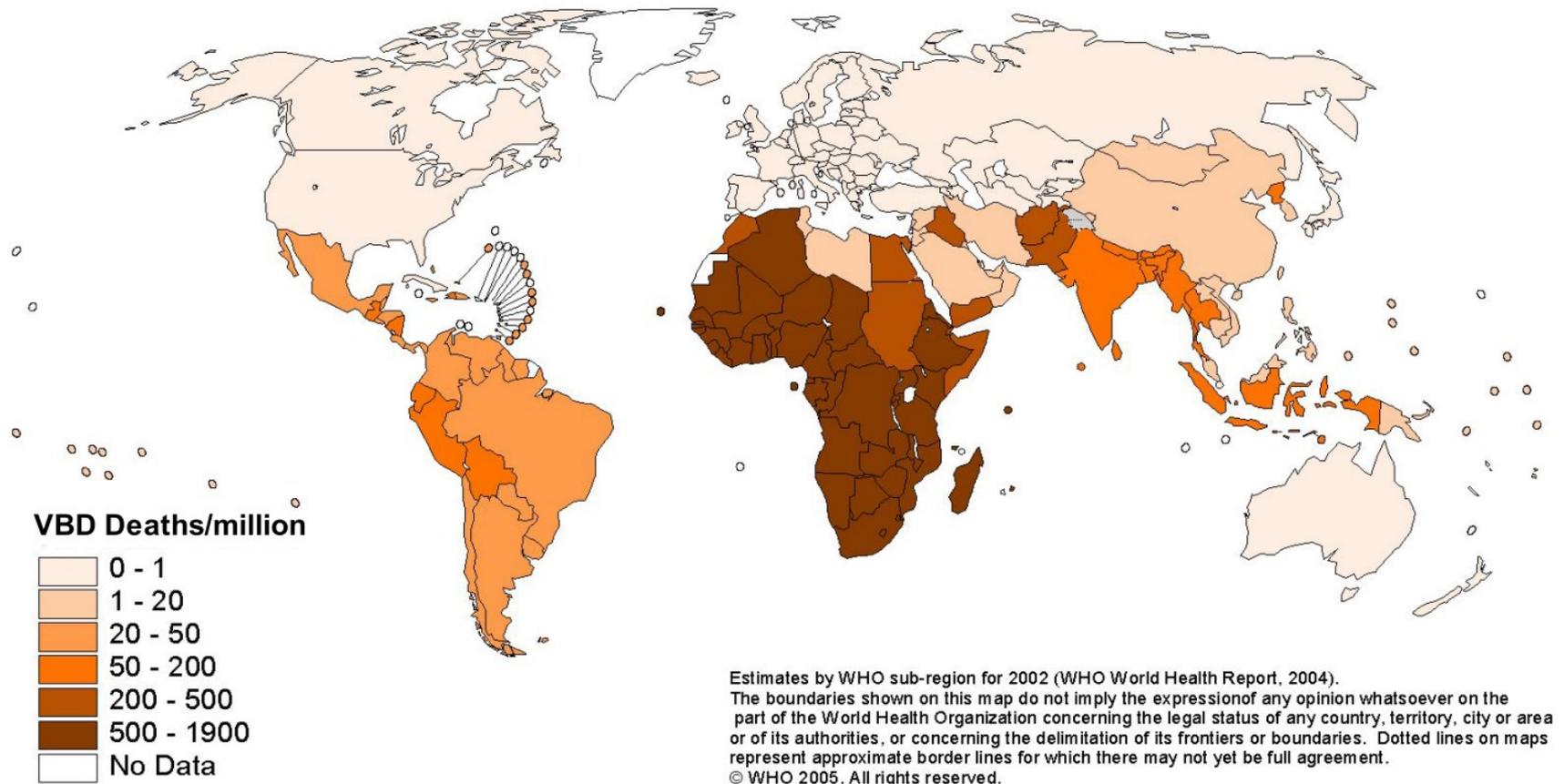
Vector-borne diseases

- Diseases that result from an infection transmitted to humans and animals by blood-sucking arthropods and some aquatic snails
- Many of these diseases are found in tropical and sub-tropical regions, as well as locations where there is minimal access to potable water and proper sanitation systems
- Various types of vectors:
 - Mosquitoes
 - Ticks
 - Fleas
 - Reduviids
 - Black flies
 - Tsetse flies
 - Sandflies
 - Aquatic snails



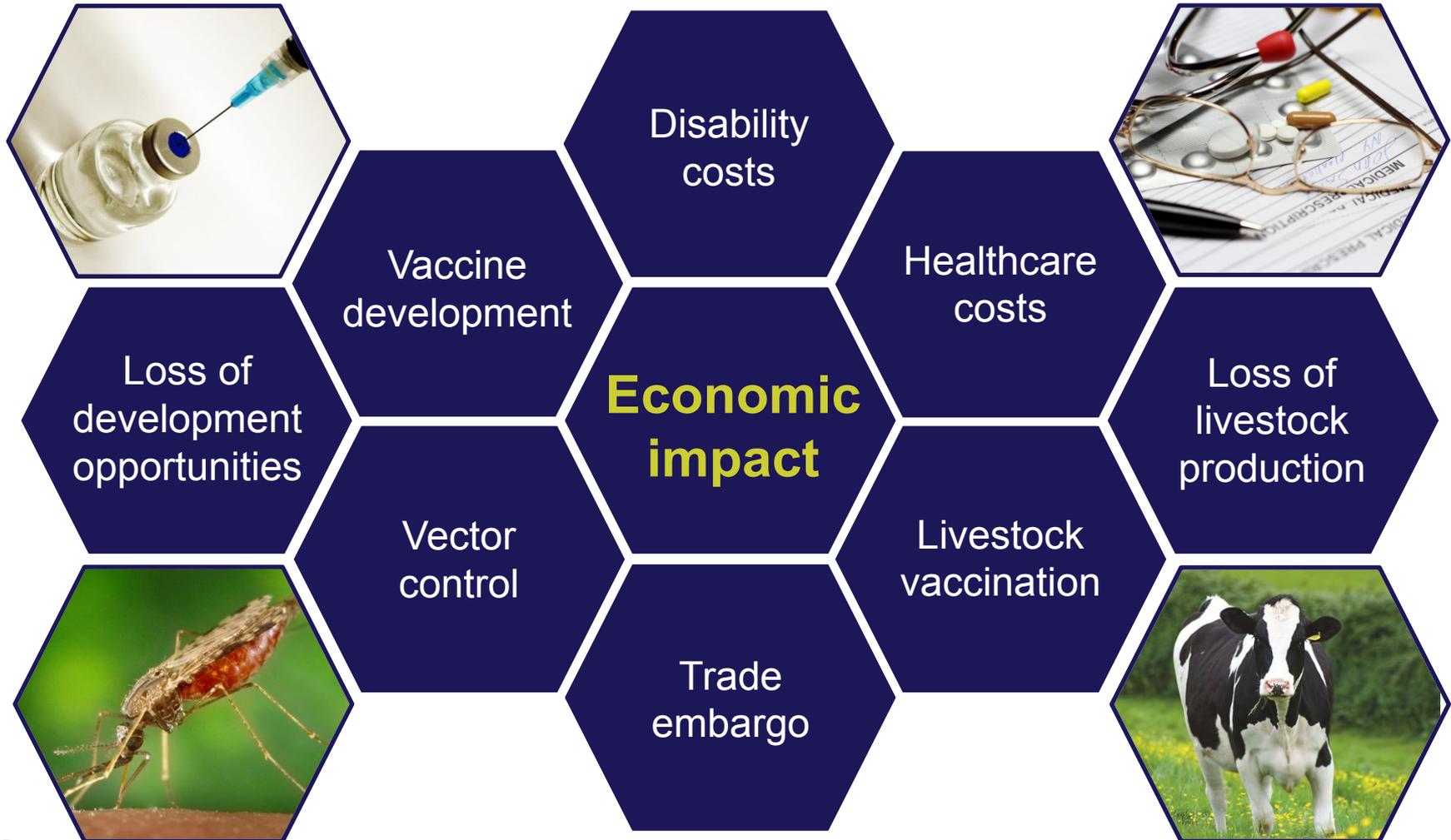
The World Health Organization estimates that vector-borne diseases account for 17% of the estimated global burden of all infectious diseases, resulting in more than 1 million deaths annually.

Vector-borne diseases



Deaths from vector-borne disease for 2002
WHO World Health Report, 2004

Vector-borne diseases



Vector-borne diseases



Mosquitoes

Chikungunya, West Nile, Eastern equine encephalitis, St. Louis encephalitis, Dengue, Malaria



Black flies

Filariasis, Onchocerciasis



Ticks

Lyme disease, Ehrlichiosis, Rocky Mountain spotted fever, Babesiosis, Bartonellosis, Anaplasmosis



Tsetse flies

Sleeping sickness



Fleas

Plague, Bartonellosis



Sand flies

Leishmaniasis



Reduviids

Chagas disease



Aquatic snails

Schistosomiasis



Vector-borne diseases



Mosquitoes

Chikungunya, West Nile, Eastern equine encephalitis, St. Louis encephalitis, Dengue, Malaria



Ticks

Lyme disease, Ehrlichiosis, Rocky Mountain spotted fever, Babesiosis, Bartonellosis, Anaplasmosis



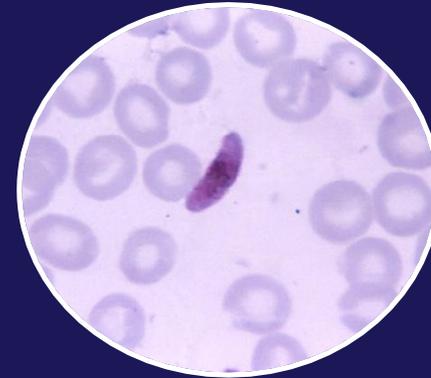
Fleas

Plague, Bartonellosis



Reduviids

Chagas disease



Malaria

- Transmitted by *Anopheles* mosquitoes
- Caused by *Plasmodium* spp.
- Considered to be the most deadly vector-borne disease
- Results in approximately 600,000 deaths annually
- Resistance to antimalarial drugs is a recurring problem



Vector-borne diseases



Mosquitoes

Chikungunya, West Nile, Eastern equine encephalitis, St. Louis encephalitis, Dengue, Malaria



Ticks

Lyme disease, Ehrlichiosis, Rocky Mountain spotted fever, Babesiosis, Bartonellosis, Anaplasmosis



Fleas

Plague, Bartonellosis



Reduviids

Chagas disease



Dengue

- Transmitted by *Aedes aegypti*
- Caused by Dengue virus serotypes 1, 2, 3, and 4
- Fastest growing vector-borne disease
- Over 40% of the world's population is at risk
- Estimated 50-100 million infections annually



Vector-borne diseases



Mosquitoes

Chikungunya, West Nile, Eastern equine encephalitis, St. Louis encephalitis, Dengue, Malaria



Ticks

Lyme disease, Ehrlichiosis, Rocky Mountain spotted fever, Babesiosis, Bartonellosis, Anaplasmosis



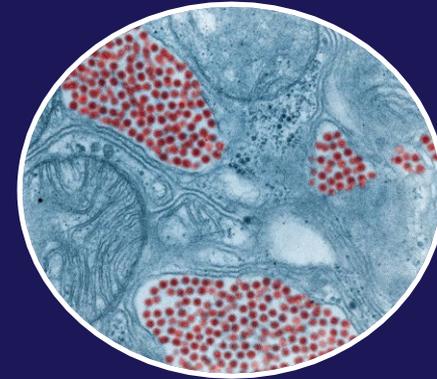
Fleas

Plague, Bartonellosis



Reduviids

Chagas disease



Eastern equine encephalitis

- Transmitted by *Culiseta melanura*, and some *Aedes* and *Culex* spp.
- Caused by Eastern equine encephalitis virus (EEEV)
- Estimated 30-70% mortality rate in the United States
- Most survivors exhibit significant brain damage



Vector-borne diseases



Mosquitoes

Chikungunya, West Nile, Eastern equine encephalitis, St. Louis encephalitis, Dengue, Malaria



Ticks

Lyme disease, Ehrlichiosis, Rocky Mountain spotted fever, Babesiosis, Bartonellosis, Anaplasmosis



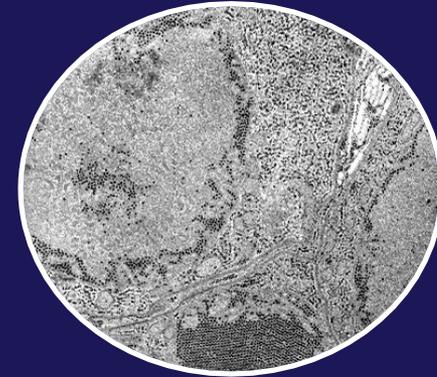
Fleas

Plague, Bartonellosis



Reduviids

Chagas disease



St. Louis encephalitis

- Transmitted by *Culex* spp.
- Caused by the Saint Louis encephalitis virus (SLEV)
- The principal reservoirs of SLEV include wild birds and domestic fowl
- The mortality rate ranges from 5-30%, with higher rates among the elderly



Vector-borne diseases



Mosquitoes

Chikungunya, West Nile, Eastern equine encephalitis, St. Louis encephalitis, Dengue, Malaria



Ticks

Lyme disease, Ehrlichiosis, Rocky Mountain spotted fever, Babesiosis, Bartonellosis, Anaplasmosis



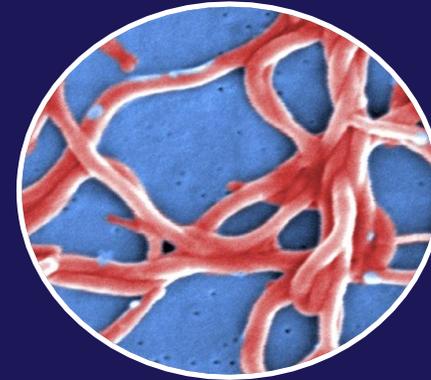
Fleas

Plague, Bartonellosis



Reduviids

Chagas disease



Lyme disease

- Transmitted by *Ixodes scapularis* and *Ixodes ricinus*
- Caused by *Borrelia burgdorferi*, *Borrelia afzelii*, and *Borrelia garinii*
- Reported in 80+ countries worldwide
- Estimated 300,000 infections annually in the United States alone



Vector-borne diseases



Mosquitoes

Chikungunya, West Nile, Eastern equine encephalitis, St. Louis encephalitis, Dengue, Malaria



Ticks

Lyme disease, Ehrlichiosis, Rocky Mountain spotted fever, Babesiosis, Bartonellosis, Anaplasmosis



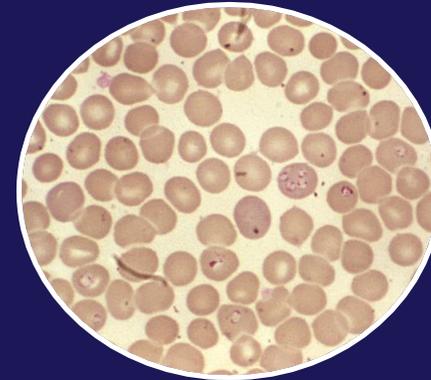
Fleas

Plague, Bartonellosis



Reduviids

Chagas disease



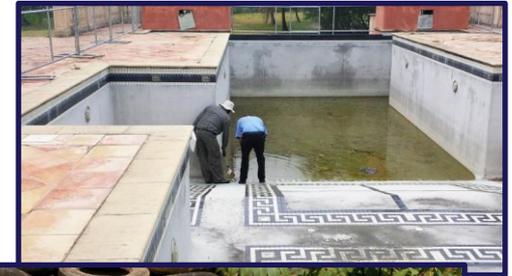
Babesiosis

- Transmitted by *Ixodes scapularis*
- Caused by *Babesia microti*
- Parasites infect red blood cells
- May be asymptomatic, or result in fatigue, fever, and hemolytic anemia
- Can be transmitted via blood transfusion and from mother to fetus



Factors contributing to the spread of disease

- Environmental changes
- Global transportation and trade
- Demographic and societal changes
- Poor waste disposal and water storage
- Changes in agricultural practices
- Pathogen evolution



Prevention



Personal protection/prevention strategies

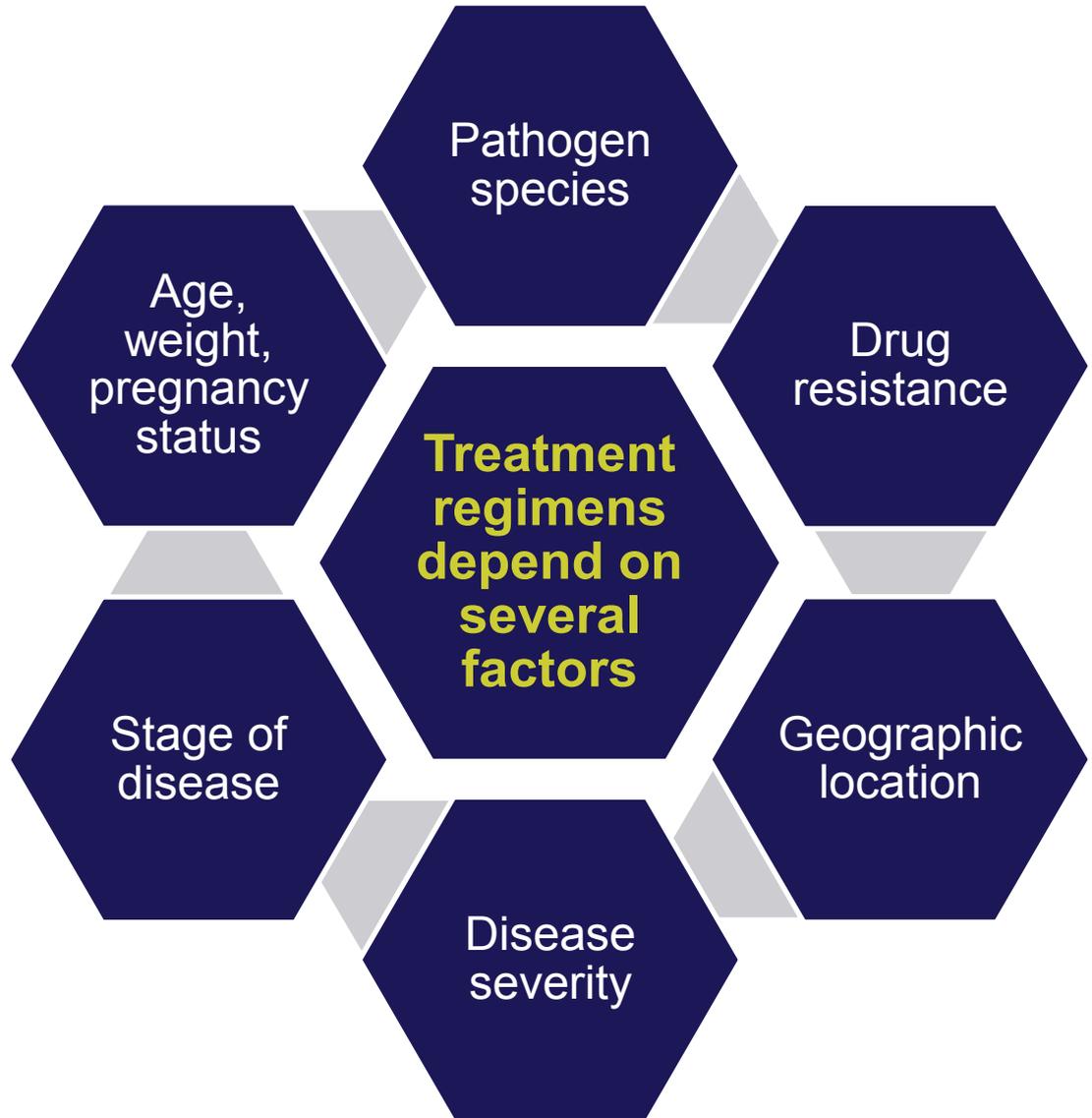
- Keep skin covered
- Mosquito nets
- Insecticides
- Avoid insect vector habitats
- Preventing access to egg-laying habitats



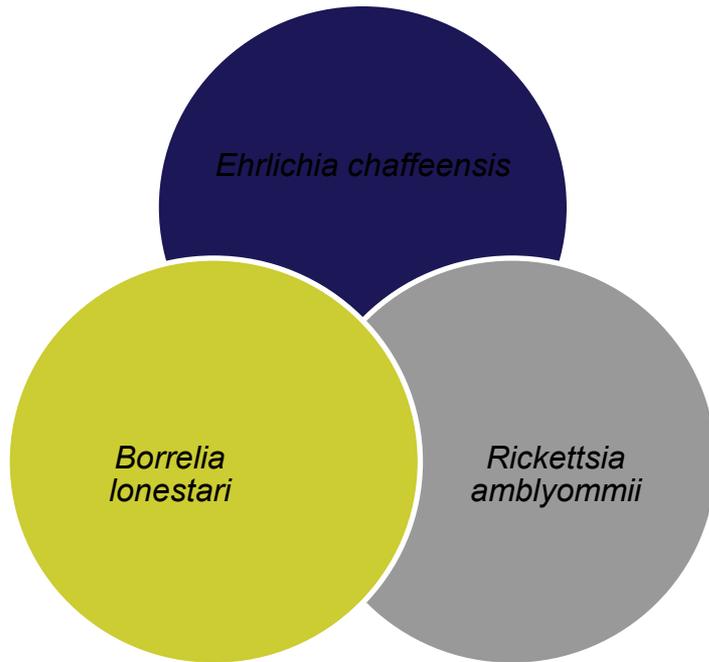
Environmental management strategies

- Active monitoring and surveillance of vectors and vector environments
- Improved design or operation of water resources development projects
- Biological controls that target vector larvae

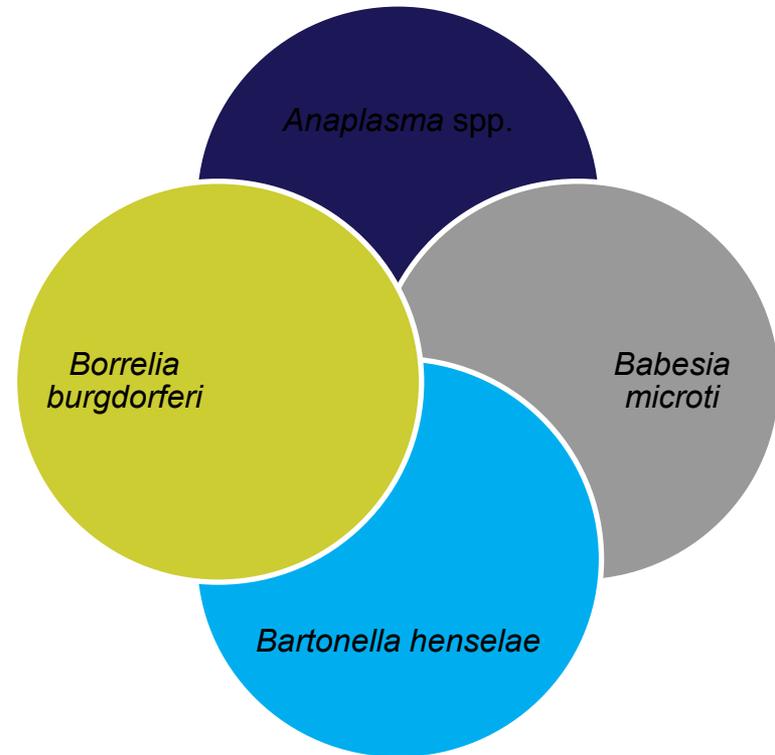
Treatment



Vector-borne pathogen co-infection



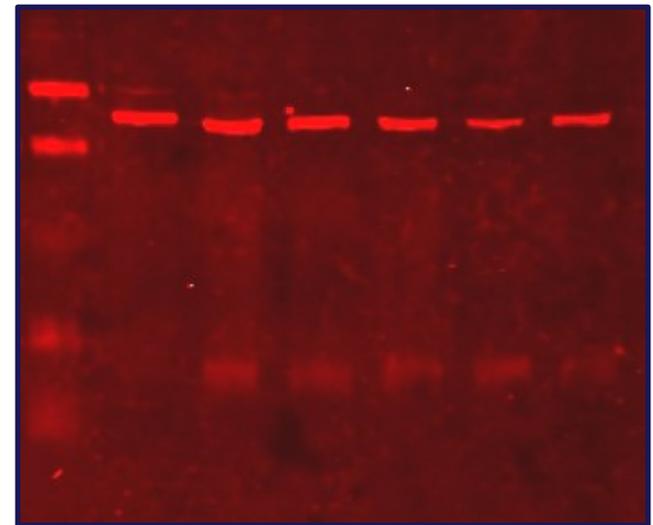
Amblyomma americanum tick vector



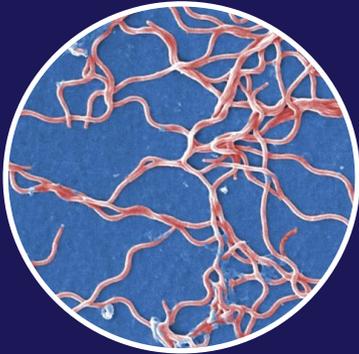
Ixodes scapularis tick vector

Detection of vector-borne diseases

- Detection Methods:
 - PCR
 - RT-PCR
 - ELISA
 - Western blot
 - Histopathology
 - Blood smear
- What is needed:
 - Field survey kits
 - Fast, accurate, inexpensive laboratory-based surveillance

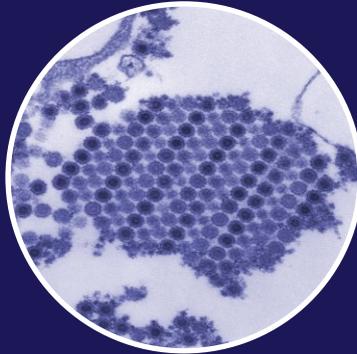


ATCC Genuine Cultures®



Bacteria

Anaplasma
Borrelia
Ehrlichia
Rickettsia



Viruses

Chikungunya virus
Dengue virus
Western equine
encephalitis virus
West Nile virus



Protozoa

Babesia
Leishmania
Plasmodium
Trypanosoma

Visit us online at www.atcc.org/vectorborne



ATCC® Genuine Nucleics



Genomic Nucleic Acids

Babesia

Borrelia

Leishmania

Plasmodium

Trypanosoma

Yersina pestis



Synthetic Nucleic Acids

Dengue virus I-IV

Eastern equine encephalitis virus

St. Louis encephalitis virus

West Nile virus

Visit us online at www.atcc.org/vectorborne

ATCC[®] Synthetic Molecular Standards

Can be used as a positive control for:

- Difficult to culture or unculturable strains
- Strains requiring BSL-3 containment
- Strains on the commerce control list

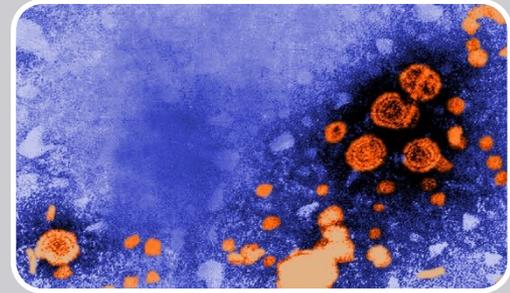
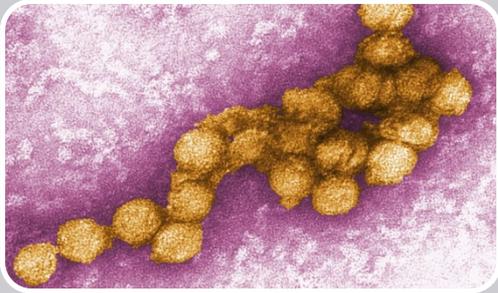
Advantages of synthetic nucleic acids:

- Eliminate the need to culture microorganisms
- Can be used in a BSL-1 facility
- No shipping restrictions
- Manufactured under ISO 13485:2003
- Quantified using Droplet Digital™ PCR
- Useful for monitoring assay-to-assay or lot-to-lot variation



ATCC® Synthetic Molecular Standards

The current collection



Vector-borne disease research

- West Nile virus
- Dengue virus serotypes 1-4
- Eastern equine encephalitis virus
- St. Louis encephalitis virus

Enteric research

- Norovirus GI & GII
- Sapovirus

STI & bloodborne pathogen research

- *Mycoplasma genitalium*
- *Treponema pallidum*
- Hepatitis B
- Hepatitis C
- HPV 16 & 18

ATCC[®] Synthetic Molecular Standards

ATCC [®] No.	Synthetic Genome	Genetic Target
VR-3228SD	Dengue virus type 1	Fragments from the capsid, membrane, and envelope regions
VR-3229SD	Dengue virus type 2	Fragments from the capsid, membrane, and envelope regions
VR-3230SD	Dengue virus type 3	Fragments from the capsid, membrane, and envelope regions
VR-3231SD	Dengue virus type 4	Fragments from the capsid, membrane, and envelope regions
VR-3239SD	Eastern equine encephalitis virus	Fragments from the capsid, NSP1, NSP3, 3' UTR, and the E1 and E2 envelope glycoproteins
VR-3236SD	St. Louis encephalitis virus	Fragments from the NS1 gene, pre-membrane, envelope, NS5 gene, and the 3' UTR regions
VR-3198SD	West Nile virus	Fragments from the 5' UTR, capsid protein C, membrane glycoprotein precursor prM, envelope protein E, nonstructural proteins NS1, NS2A, NS3, NS5, and the 3' UTR regions



Dengue virus

Problem

- Dengue virus is the most common and clinically important arbovirus disease, with an estimated 50-100 million cases annually.
- Surveillance and rapid detection of Dengue infections is important as there are no vaccines or antivirals available.

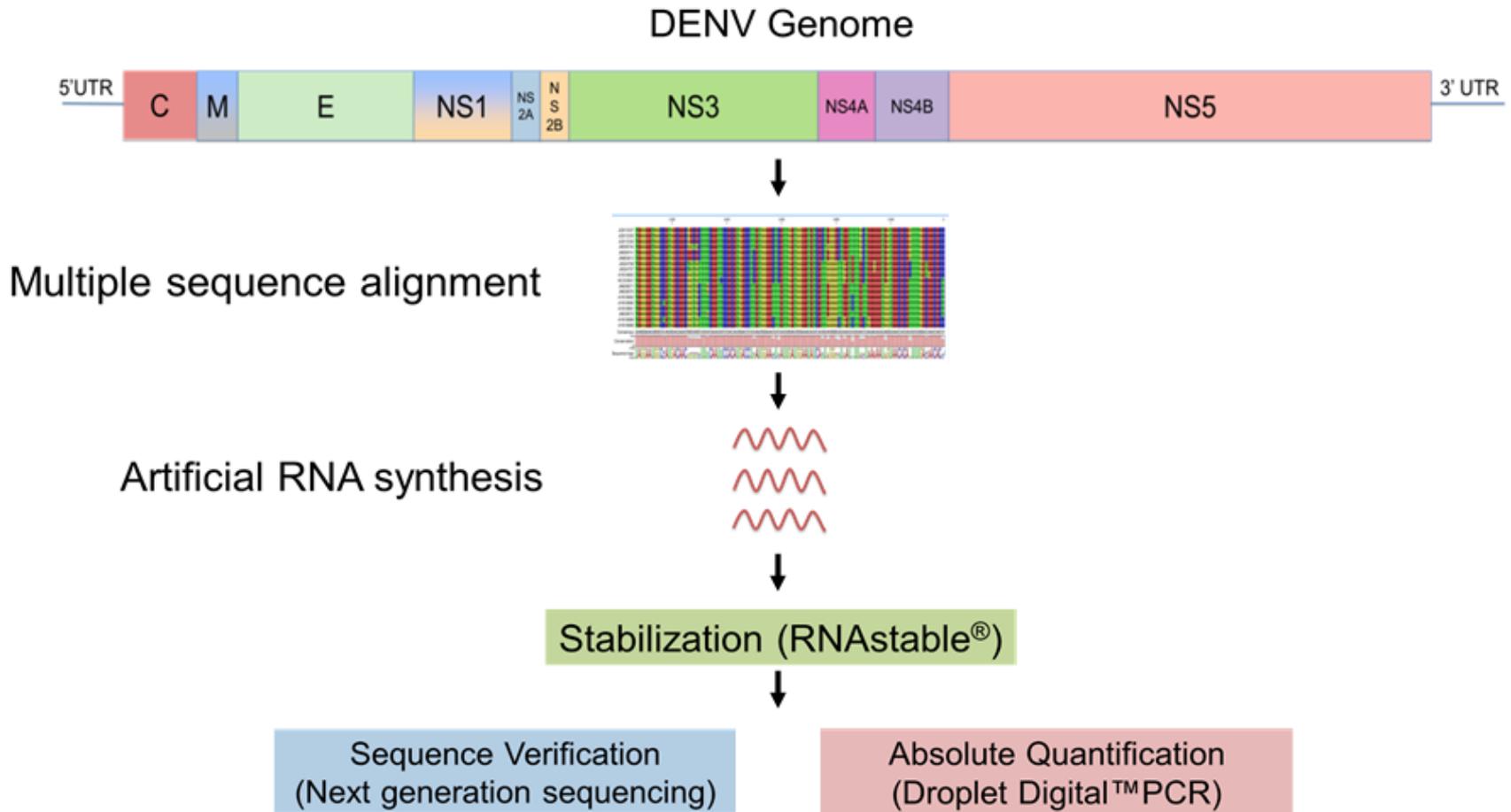
Concerns

- qRT-PCR is the preferred method for the detection and quantification of Dengue virus in clinical diagnostics and epidemiological surveillance.
- However, full-length RNA on the Commerce Control List, and cannot be shipped internationally without a permit.

Solution

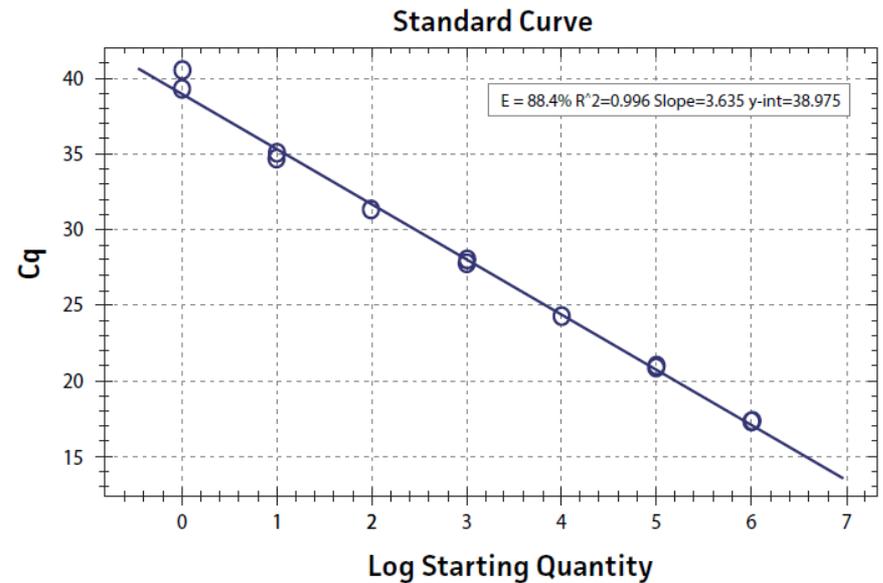
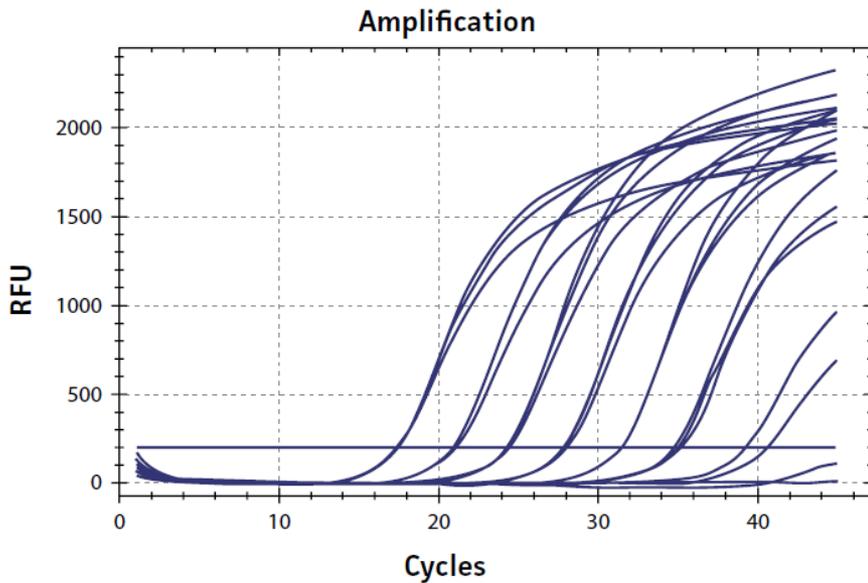
- ATCC Synthetic Dengue virus RNA is not on the Commerce Control List, and can be used in a BSL-1 facility.
- The standards were developed to contain short fragments from the capsid, membrane, and envelope genes, as well as the target regions encompassing the primer sequences from numerous published RT-PCR assays.

Synthetic Dengue virus RNA



Synthetic Dengue virus RNA

Generation standard curves using the DENV-4 Molecular Standard

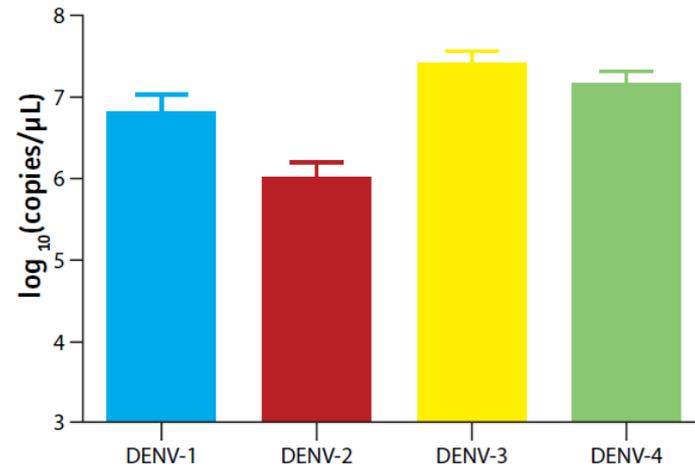
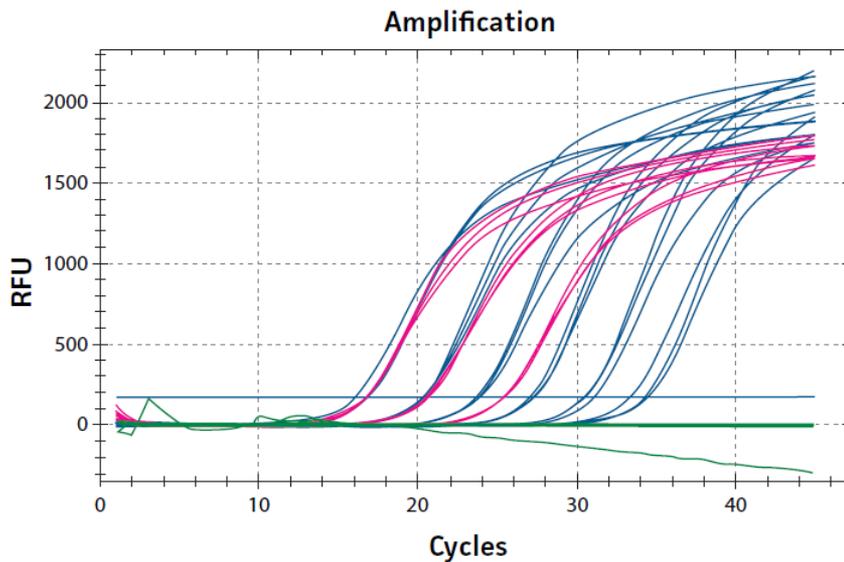


Primer and Probe		DENV-1	DENV-2	DENV-3	DENV-4
CDC Assay	Slope	-3.244	-3.277	-3.315	-3.642
	R ²	0.990	0.996	0.987	0.996
Waggoner Assay	Slope	-3.536	-3.535	-3.705	-3.775
	R ²	0.991	0.997	0.989	0.996



Synthetic Dengue virus RNA

Quantification of native DENV RNA



Titers of DENV-1, -2, -3, and -4 samples were determined using the qRT-PCR standard curves generated by the DENV 1-4 molecular standards

DENV 1 = TH-S-man (ATCC® VR-1586™)

DENV 2 = New Guinea C (NR-84, BEI Resources)

DENV 3 = Philippines/H87/1956 (NR-80, BEI Resources)

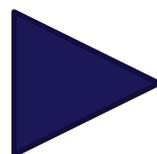
DENV 4 = H241 (ATCC® VR-1257™)



Synthetic Dengue virus RNA

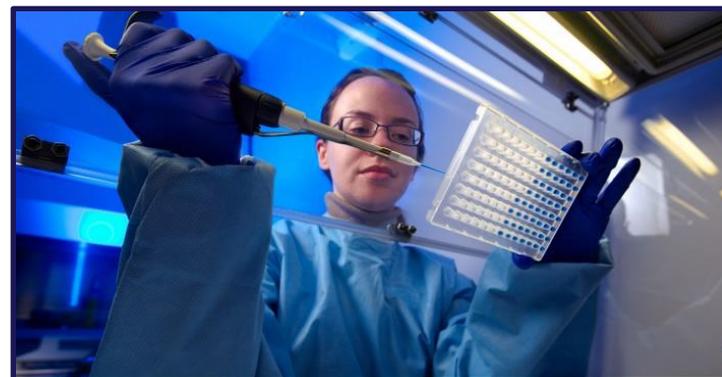
Characteristics

- Fully authenticated
- Quantitated and stable
- Can be handled in BSL-1
- No special permits required
- Exhibit minimal variability
- Compatible with published real-time PCR assays



Applications

- Positive controls in qRT-PCR assays
- Pathogen detection
- Quantify viral load
- Technology can be expanded to other strains





Conclusion

- Vector-borne diseases are a major cause of morbidity and mortality
- Due to limited preventative and therapeutic treatment options, accurate detection methods are critical
- ATCC offers authenticated strains and nucleic acids that support the development, validation, and employment of novel detection methods
- ATCC Synthetic Molecular Standards
 - Represent key target regions for difficult-to-culture, unculturable, and high containment microorganisms
 - Authenticated, stable, and quantitative
 - Safe to use in BSL-1 facilities
 - Used to detect and quantify bacterial and viral load
 - Generate standard curves
 - Function as positive controls in molecular-based applications

Thank you!

Register for more webinars in the ATCC “*Excellence in Research*” webinar series at www.atcc.org/webinars.



March 19, 2015

10:00 AM, 3:00 PM EST

Scott Sutton, Ph.D., *Principal*, Microbiology Network and
Liz Kerrigan, *Direct, New Product Development, Sales & Marketing*, ATCC
Microbiological quality control of pharmaceutical products



May 21, 2015

10:00 AM, 3:00 PM EST

Jodie Lee, M.S.

Seeing is believing – Reporter-labeled microbial control strains

Thank you for joining today!
Please send additional questions to tech@atcc.org