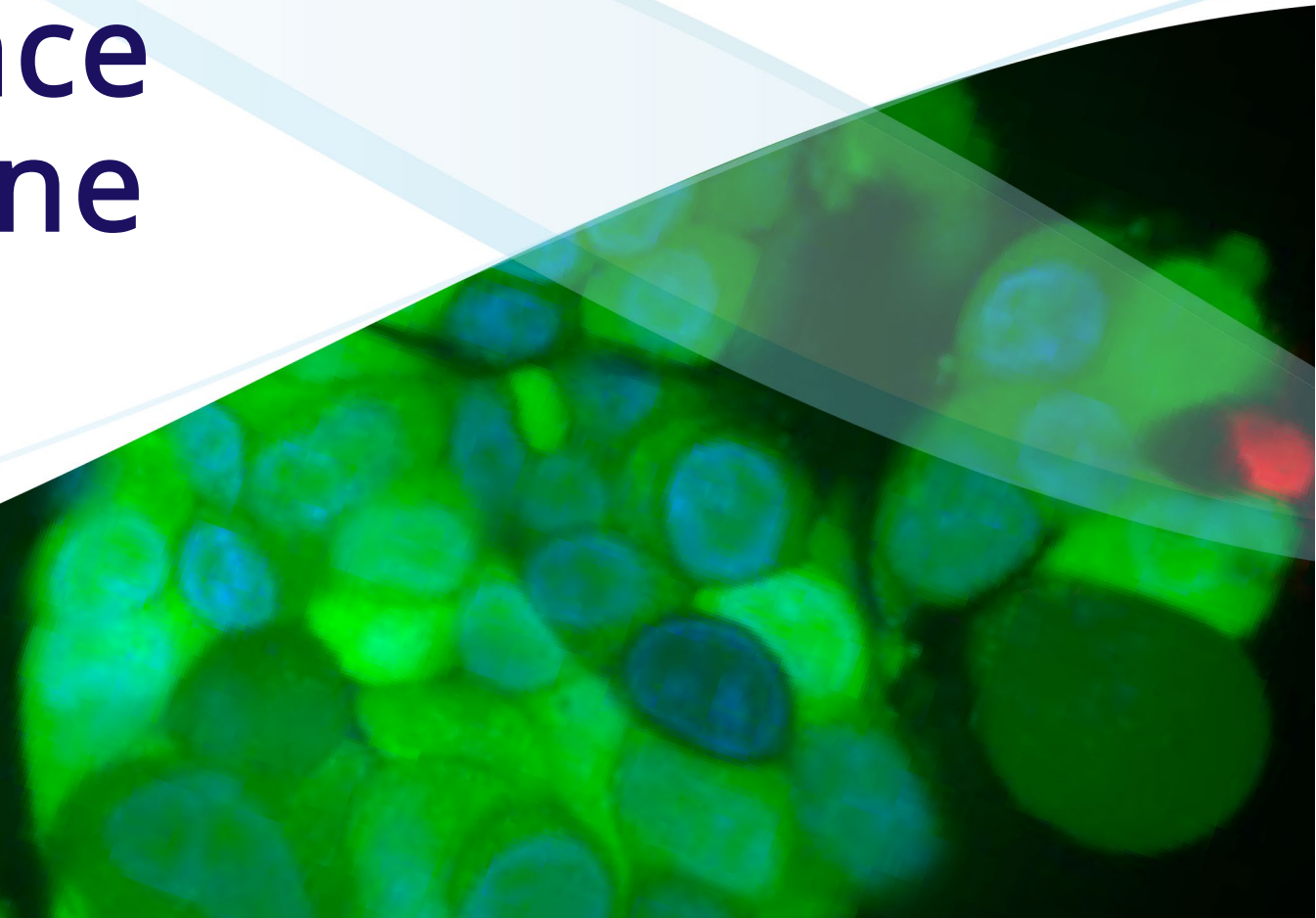


# Building Better Cancer Models to Advance Precision Medicine

AACR Spotlight 2026



# Agenda

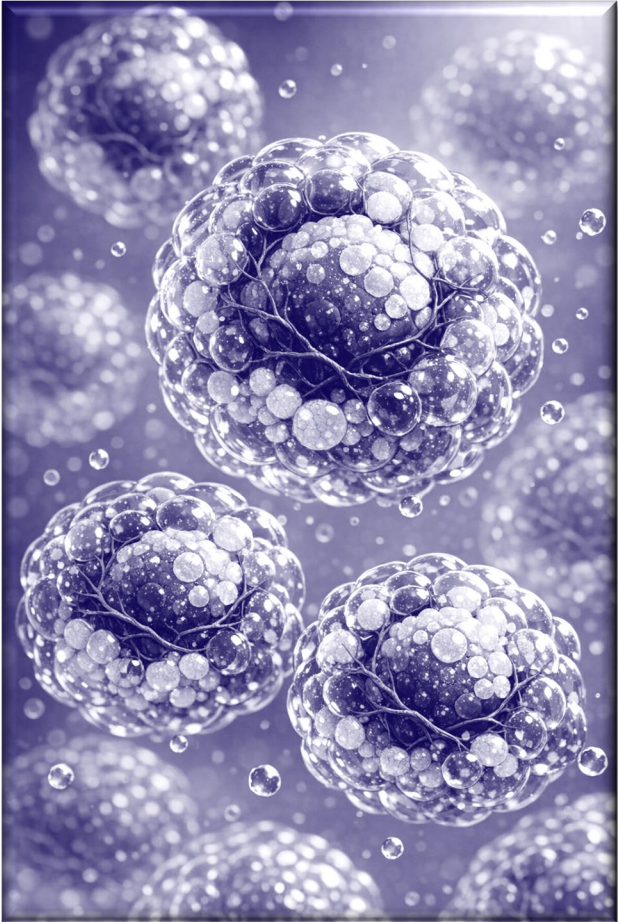


Image generated by Microsoft Copilot

- Carolina Lucchesi  
*Building Better Cancer Models to Advance Precision Medicine*
- Claudia K. Petritsch, PhD:  
*Cell Plasticity-Driven Immune Evasion in Patient-Derived Glioma Models*
- Benjamin D. Hopkins, PhD:  
*Patient Derived Tumor Organoid for Therapeutic Modeling in Cancer*

# Presenters



**Carolina Lucchesi, PhD**  
Principal Scientist, Head of  
Microphysiological Systems, ATCC



**Claudia K. Petritsch, PhD**  
Associate Professor in Research,  
Director Pediatric Cancer Model  
Development Center, Sr. Scientist in  
Neuroscience, Stanford University



**Benjamin David Hopkins, PhD**  
Assistant Professor of Research in  
Systems and Computational  
Biomedicine, Weill Cornell Medical  
College

# About Us



ATCC is a global leader in providing authenticated, high-quality biological resources and standards for industry, academia, and government.

- Founded in 1925, ATCC is a private, nonprofit, global biological resource center and standards organization that provides scientists with the biomaterials and resources they need to conduct critical life science research.
- World's trusted, premier biological materials resource and standards development organization:
  - 4,000+ cell lines
  - 80,000+ microorganisms
  - Genomic and synthetic nucleic acids
  - Media, sera, and reagents
  - Advanced cell models
  - Standards



# Building Better Cancer Models to Advance Precision Medicine

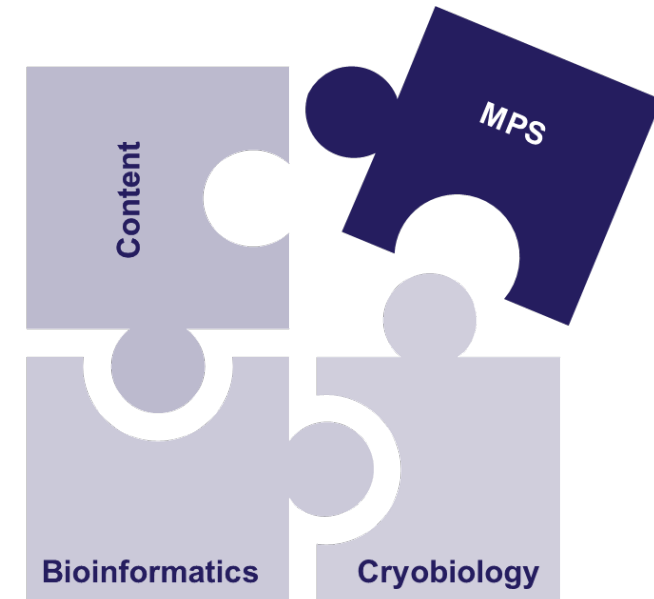
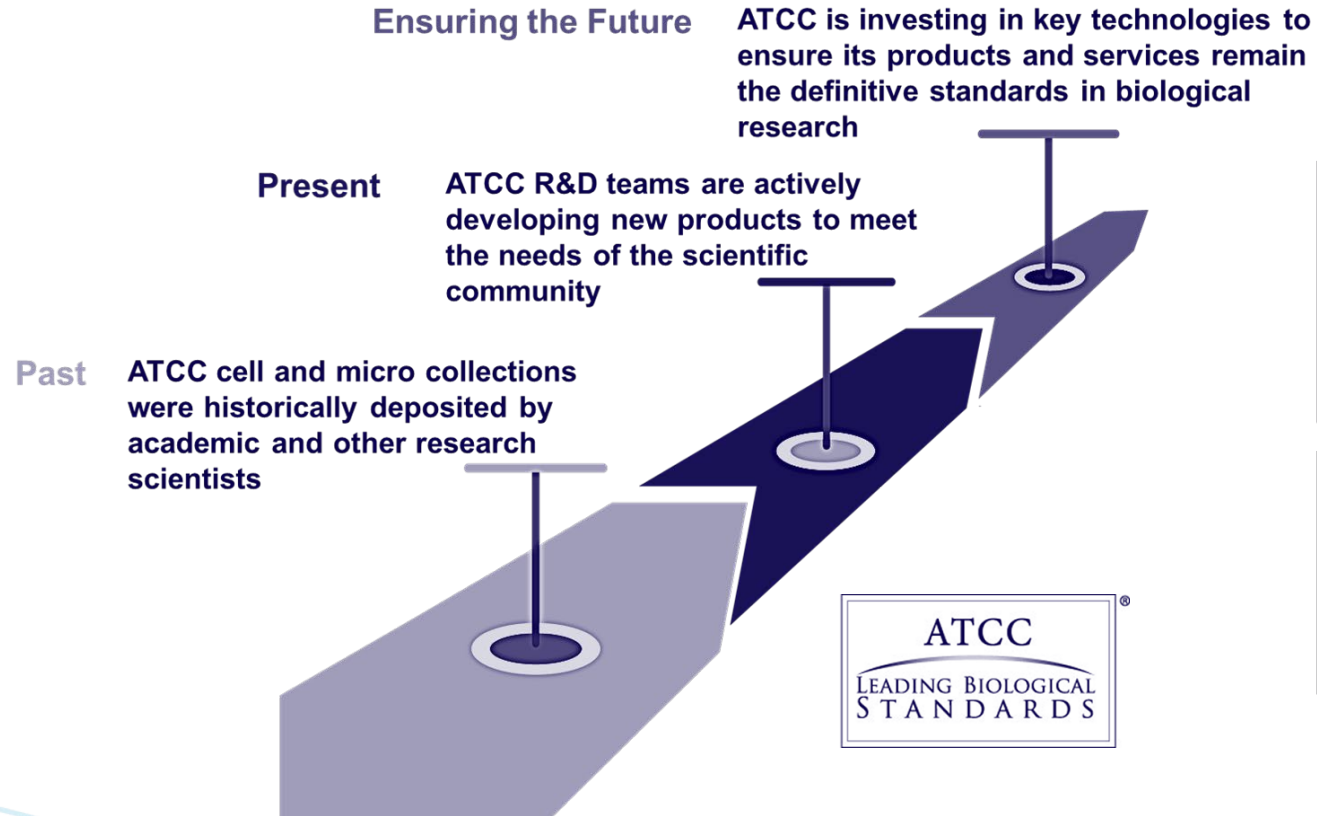
AACR Spotlight 2026

# Modernization of the ATCC in vitro cell model portfolio



Established partner to global researchers

Image generated by Microsoft Copilot



Research & Development

# ATCC's Human Cancer Models Initiative

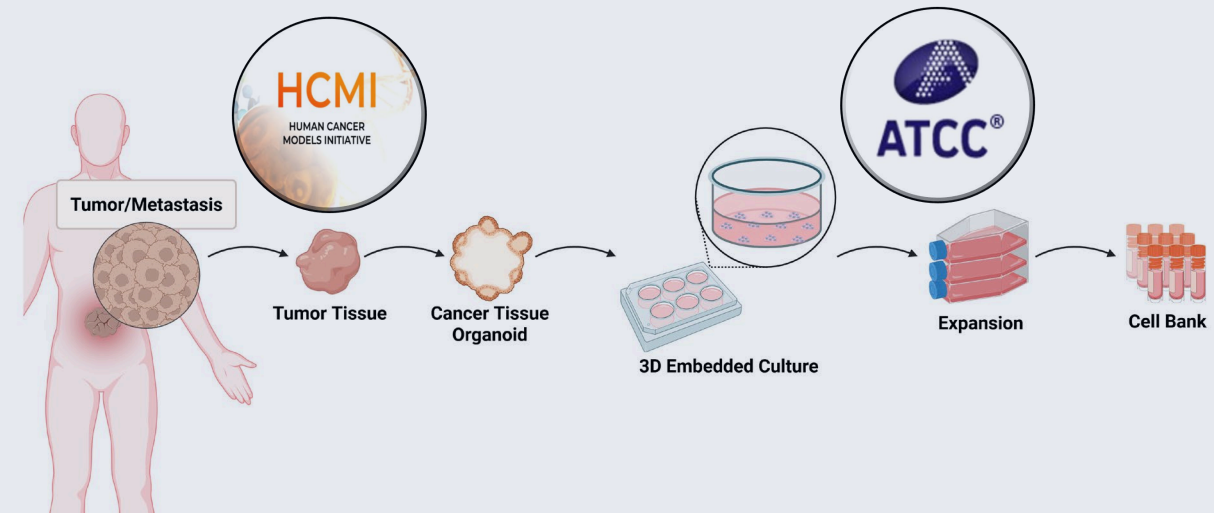


- The Human Cancer Models Initiative (HCMI) is an international consortium dedicated to generating patient-derived cancer models to facilitate cancer research.
- **ATCC is the sole distributor of HCMI models**

## Founders and Members

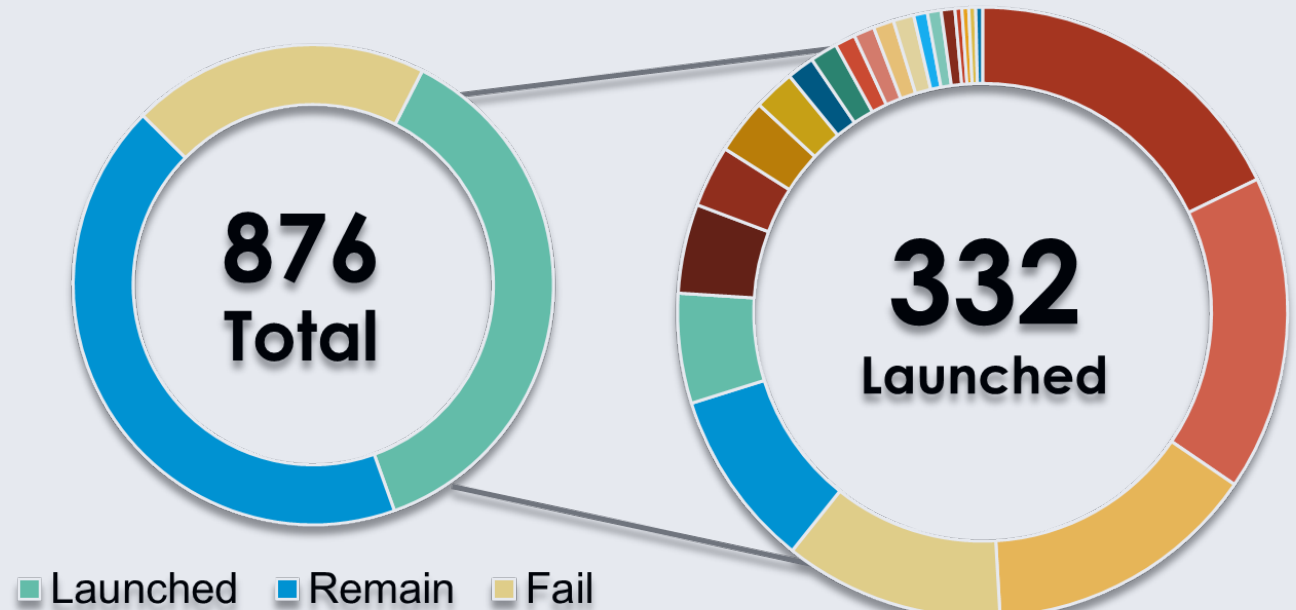
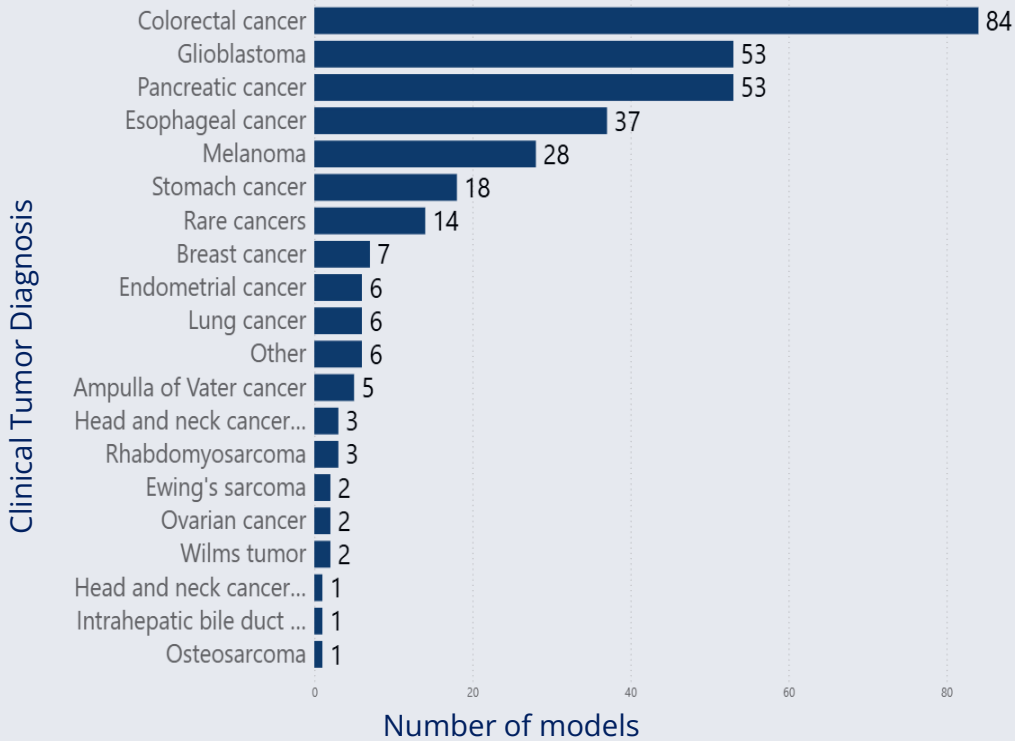
- Broad Institute
- Cancer Research UK
- Cold Spring Harbor Laboratories
- Cornell University
- Hubrecht Organoid Technology Foundation
- National Cancer Institute
- Stanford University
- Wellcome Sanger Institute and others

## ATCC's HCMI Workflow



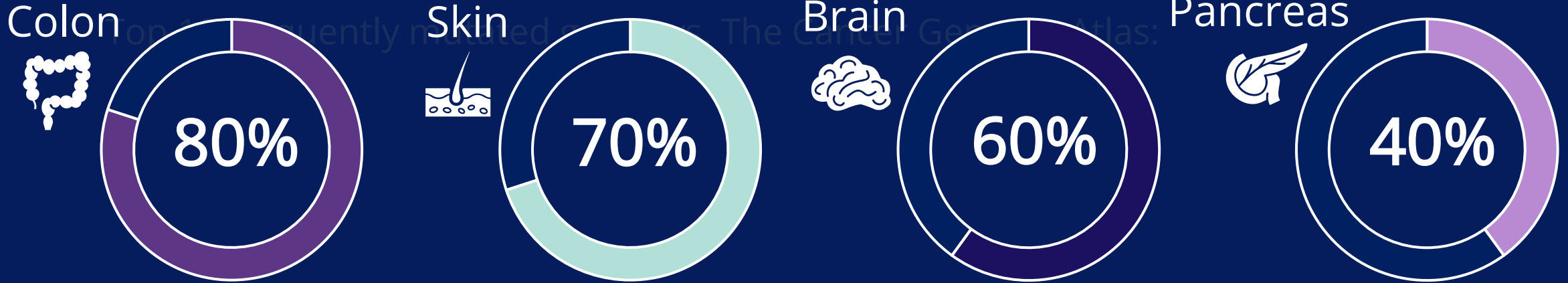
Created with BioRender.com

# HCMI Portfolio Model Diversity



Collection includes models derived from rare adult and pediatric cancer such as rhabdomyosarcoma, leiomyosarcoma, Ewing sarcoma and Will tumor

# Model Relevance



Top 10 genes captured in the HCMI models:

|       |       |
|-------|-------|
| APC   | LRP1B |
| TP53  | ZFHX4 |
| KRAS  | CSMD3 |
| MUC16 | ROBO2 |
| FAT4  | ALK   |

|                       |        |
|-----------------------|--------|
| TP53, BRAF, NRAS, NF1 |        |
| MUC16                 | GRIN2A |
| LRP1B                 | MECOM  |
| CSMD3                 | FAT4   |
| FAT3                  | DCC    |
| FAM135B               | COL1A1 |

|         |        |
|---------|--------|
| PTEN    | PIK3CA |
| TP53    | CSMD3  |
| EGFR    | BCOR   |
| CNTNAP2 | NF1    |
| PIK3CR1 | FAT1   |

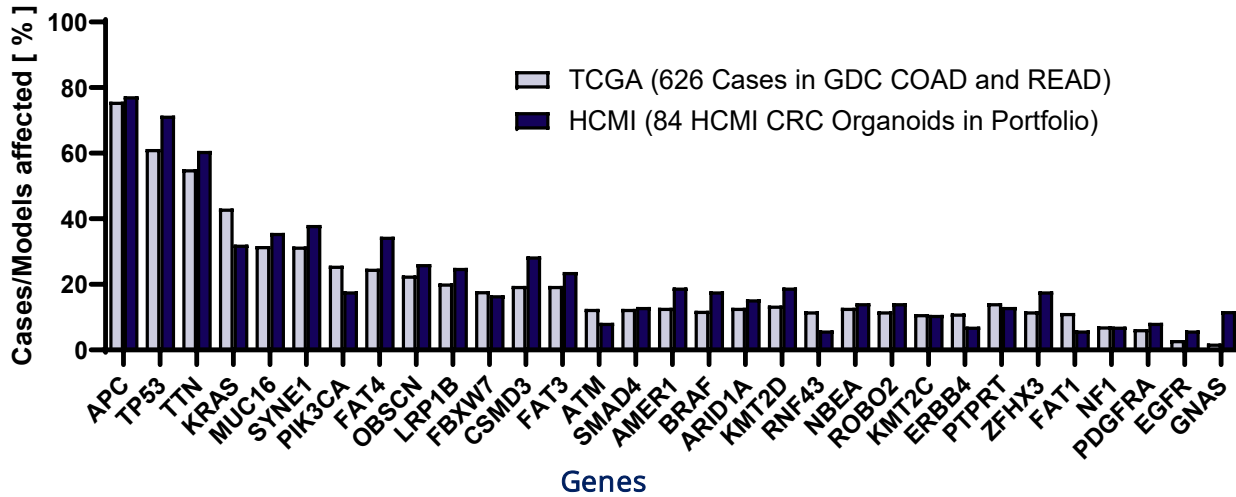
|        |       |
|--------|-------|
| KRAS   | MUC4  |
| TP53   | MUC16 |
| SMAD4  | KMT2D |
| CDKN2A | FAT3  |
| ACVR2A | RHOH  |

Canonical mutation      Matching mutation

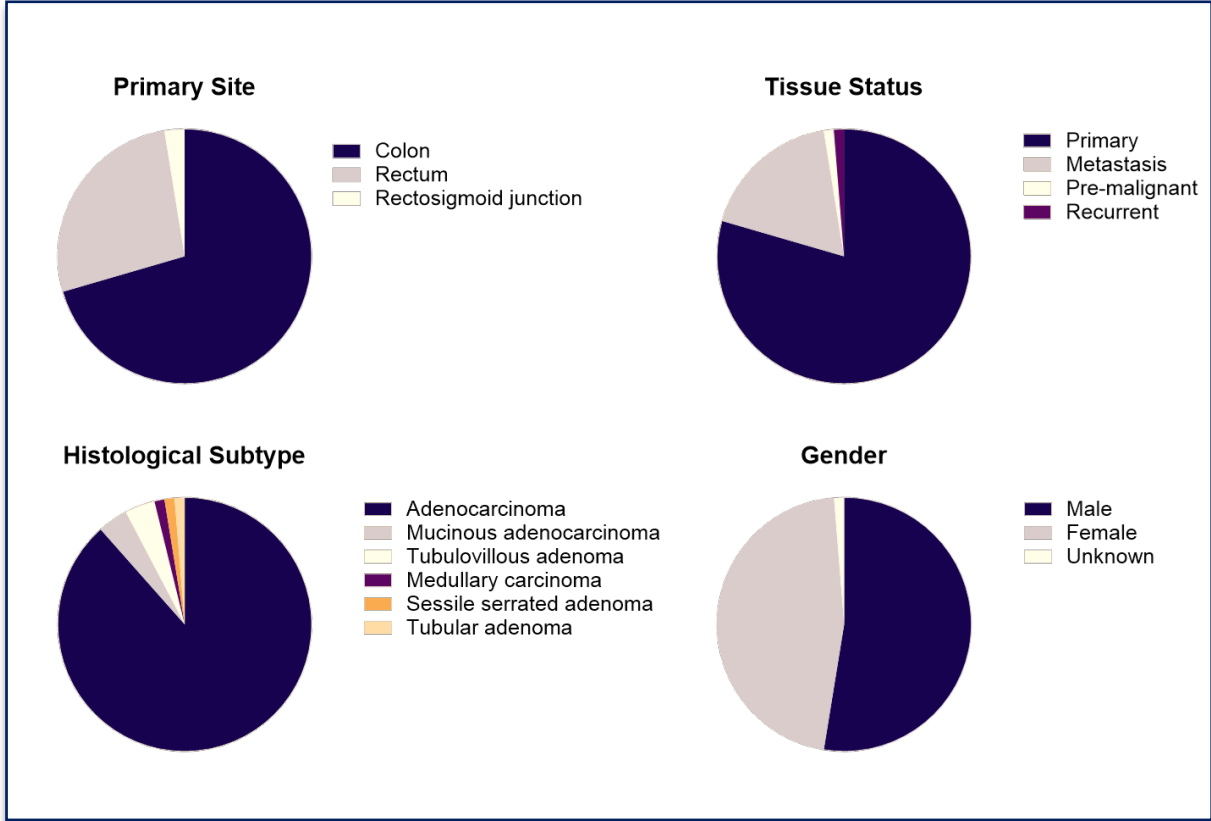
# HCFI colorectal cancer model cohort



Most Frequently Mutated Genes in Colorectal Cancer



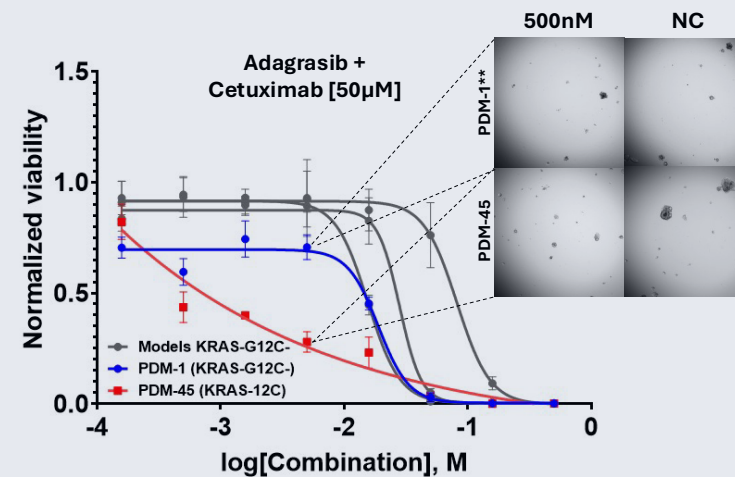
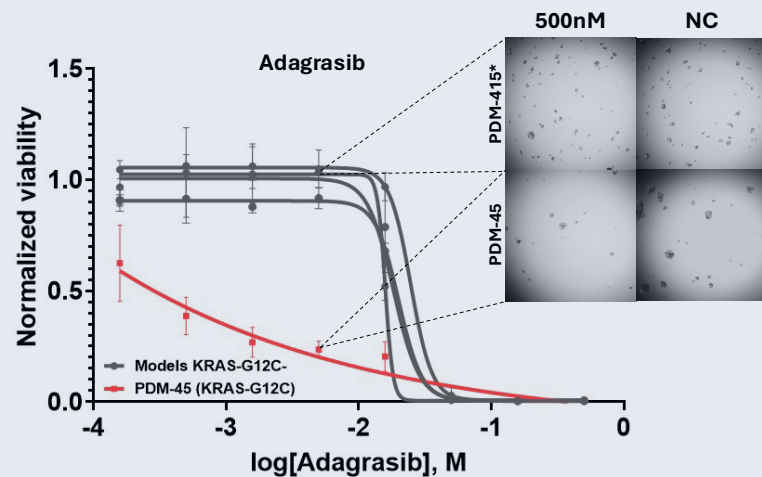
Diverse population dynamics and gene mutations in our colorectal cancer model



# Response to Adagrasib or Adagrasib+Cetumimab

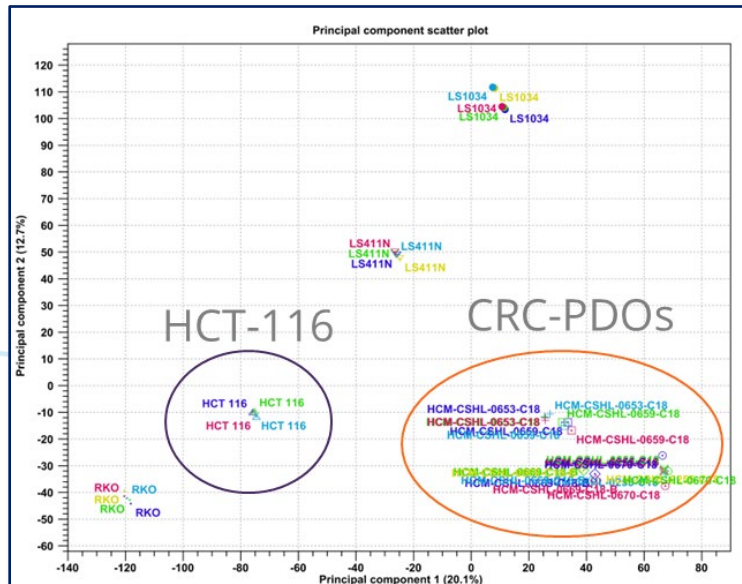
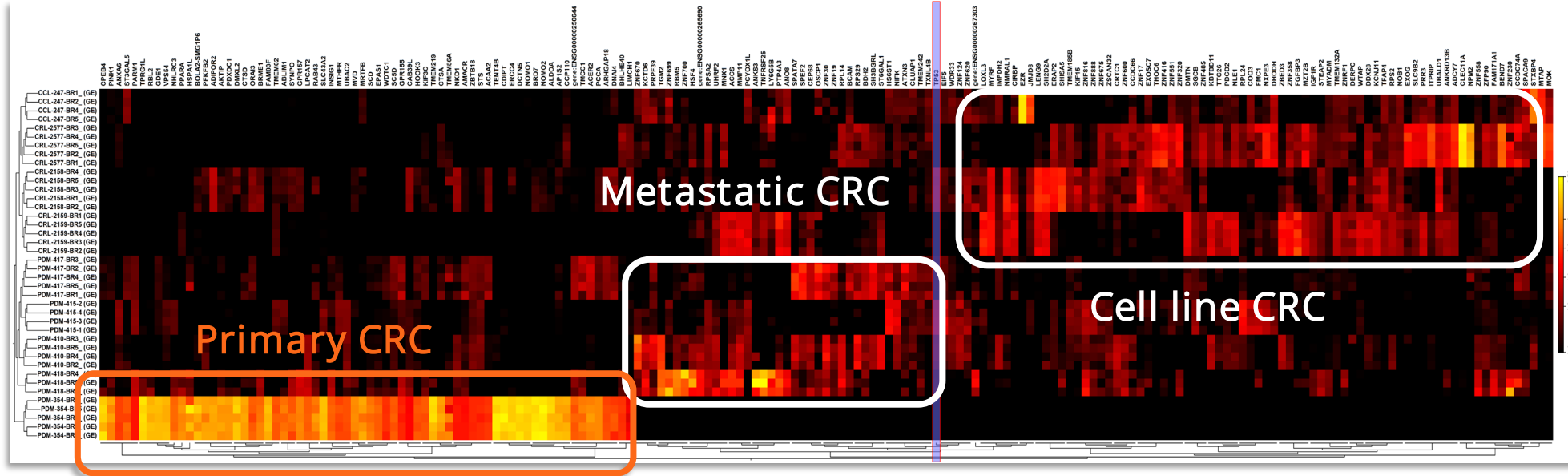


| ATCC® Model | Cancer Type | Histological Subtype | Type       | Acquisition Site | Gender | Race  | Age | Clinical Stage | KRAS Status      |
|-------------|-------------|----------------------|------------|------------------|--------|-------|-----|----------------|------------------|
| PDM-1™      | Colorectal  | Adenocarcinoma       | Primary    | Cecum            | Male   | White | 75  | Stage I        | KRAS-G12A        |
| PDM-45™     | Colorectal  | Adenocarcinoma       | Primary    | Transverse colon | Male   | --    | 80  | Stage IIIB     | KRAS-G12C        |
| PDM-354™    | Colorectal  | Adenocarcinoma       | Primary    | Sigmoid colon    | Female | --    | 70  | --             | No KRAS Mutation |
| PDM-410™    | Colorectal  | Adenocarcinoma       | Metastatic | Liver            | Female | Black | 56  | Stage IVA      | No KRAS Mutation |
| PDM-415™    | Colorectal  | Adenocarcinoma       | Metastatic | Peritoneum       | Female | Black | 48  | Stage IIIB     | KRAS-G13D        |



The KRAS-G12C model shows clear sensitivity to Adagrasib as a single agent, consistent with mutation-specific targeting.

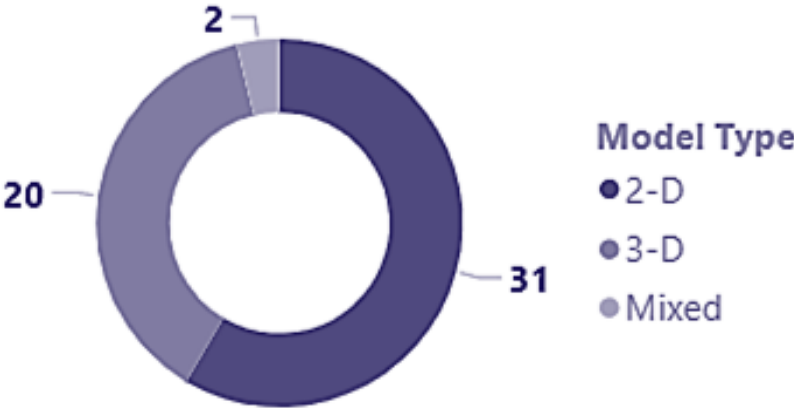
# Transcriptomic Clustering Distinguishes Organoids from Conventional Cell Lines



Patient-derived colorectal cancer organoids cluster by disease state (primary vs. metastatic) and exhibit gene expression profiles that are markedly distinct from standard cell lines (e.g., HCT116), demonstrating superior preservation of patient-specific tumor biology

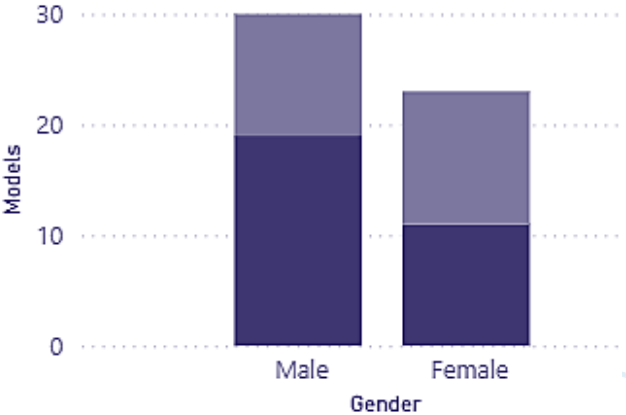
# Diversity of Glioblastoma Models

Glioblastoma Model Types

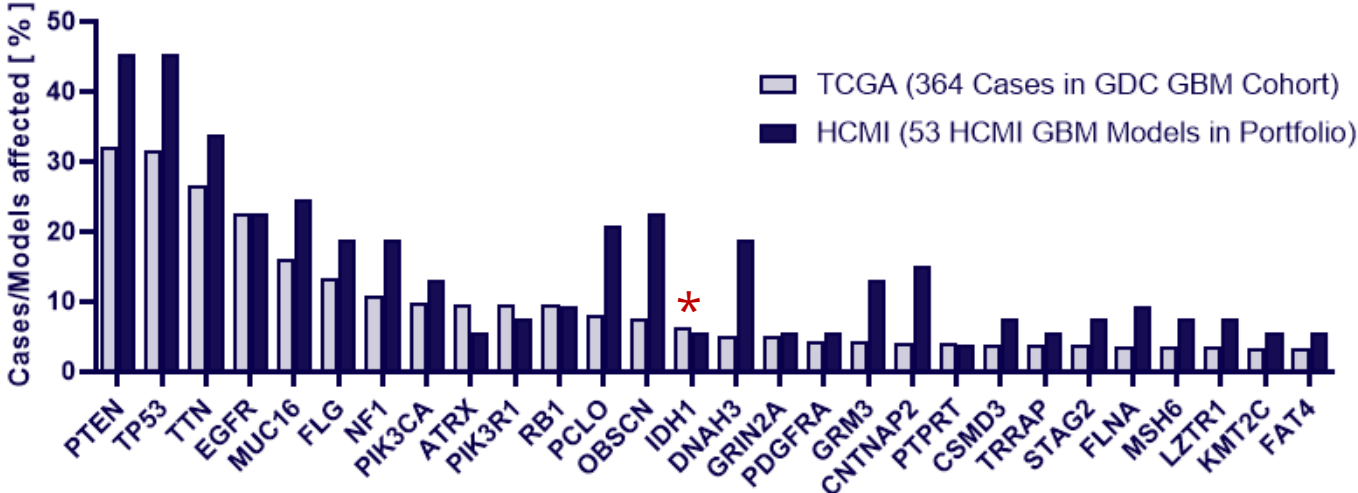


Gender and Tissue Status

Tissue Status ● Primary ● Recurrent



Most Frequently Mutated Genes in Glioblastoma Cancer

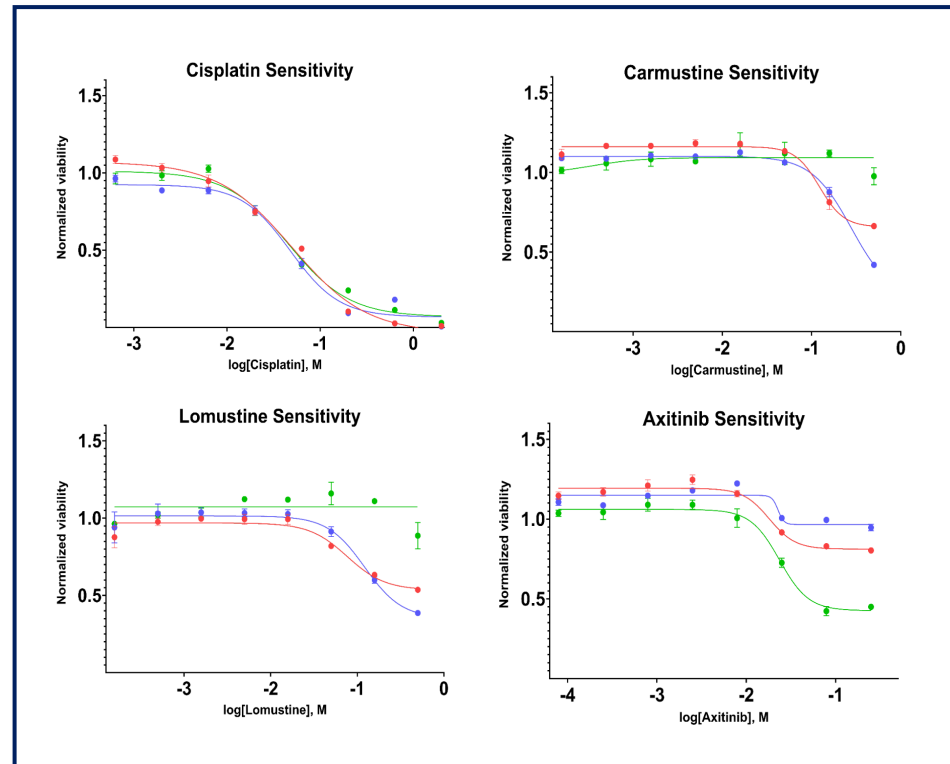
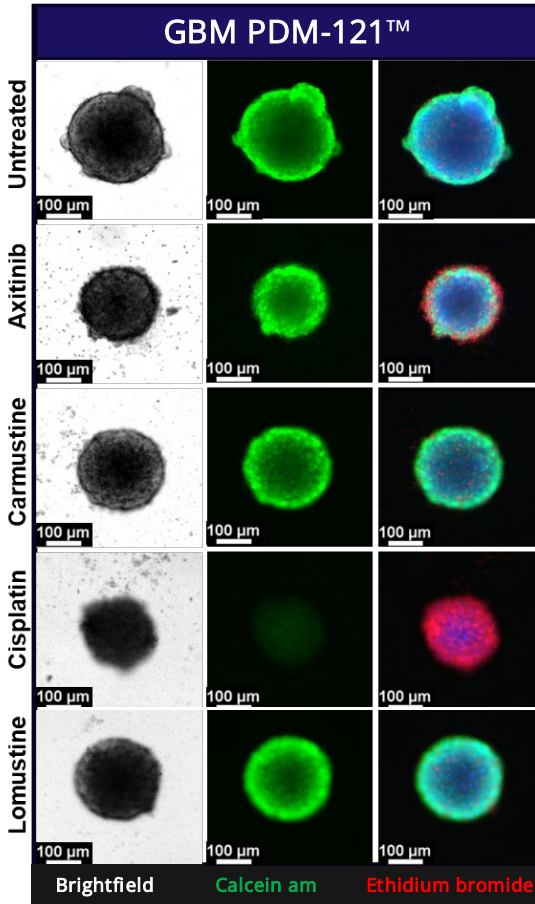


Diverse population dynamics and gene mutations in our glioblastoma model

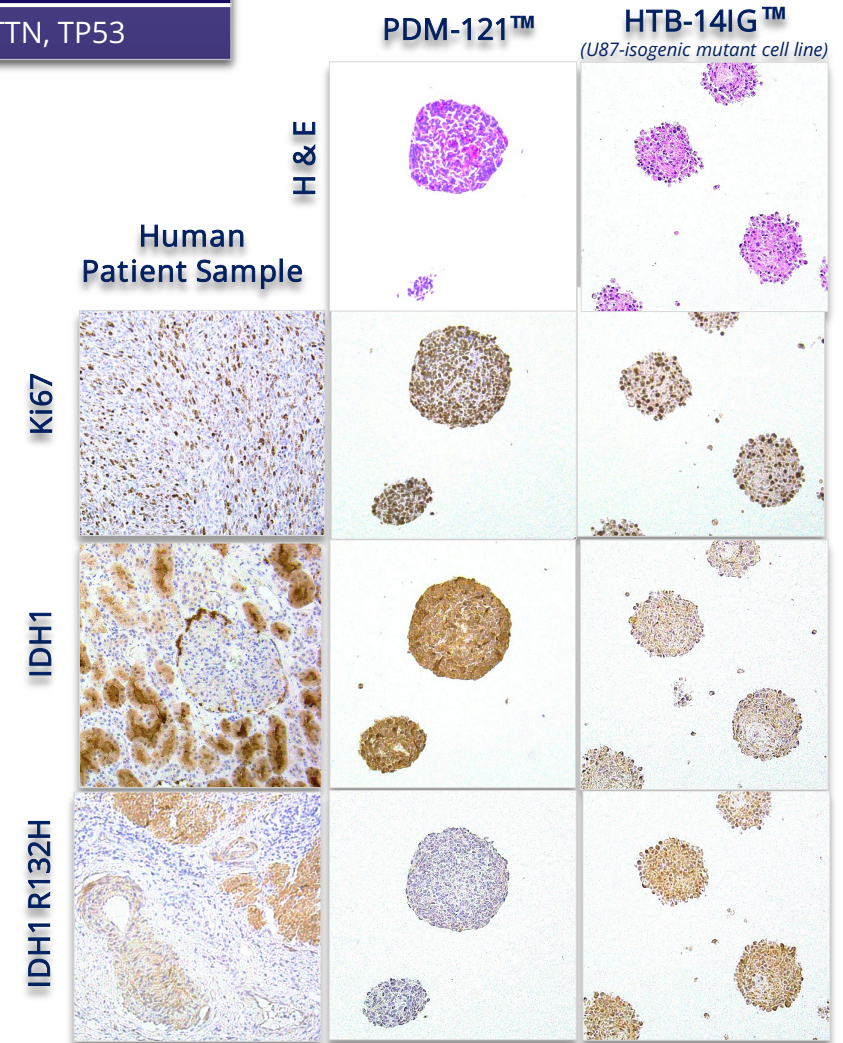
\*ACR Poster focused on IDH1

# Pediatric Glioblastoma

| ATCC Part # | Cancer Type  | Disease Status | Type        | Acquisition Site | Gender | Race  | Age | Tissue Status | Key Mutations   |
|-------------|--------------|----------------|-------------|------------------|--------|-------|-----|---------------|-----------------|
| PDM-121     | Glioblastoma | Progressive    | 3D Spheroid | Brain            | Female | White | 11  | Primary       | IDH1, TTN, TP53 |



**IDH1-driven drug resistance tested in pediatric primary neurospheres**



# Presenters



**Carolina Lucchesi, PhD**  
Principal Scientist, Head of  
Microphysiological Systems, ATCC



**Claudia K. Petritsch, PhD**  
Associate Professor in Research,  
Director Pediatric Cancer Model  
Development Center, Sr. Scientist in  
Neuroscience, Stanford University



**Benjamin David Hopkins, PhD**  
Assistant Professor of Research in  
Systems and Computational Biomedicine,  
Weill Cornell Medical College

The background of the slide features several clusters of cells, likely glioma models, stained with various fluorescent dyes. The colors include bright yellow, orange, red, and blue, set against a dark background. The cells are arranged in irregular, somewhat spherical clusters, with some showing a more organized, lattice-like structure.

# Cell Plasticity-Driven Immune Evasion in Patient-Derived Glioma Models

Claudia K Petritsch

Assoc Professor (Research), Neurosurgery

Director, Pediatric Cancer Model Development Center

& Organoid Shared Resources

Sr. Scientist, Neurology and Neurosciences



**Stanford**  
MEDICINE

# Disclosures

- No Disclosures

# Topics covered

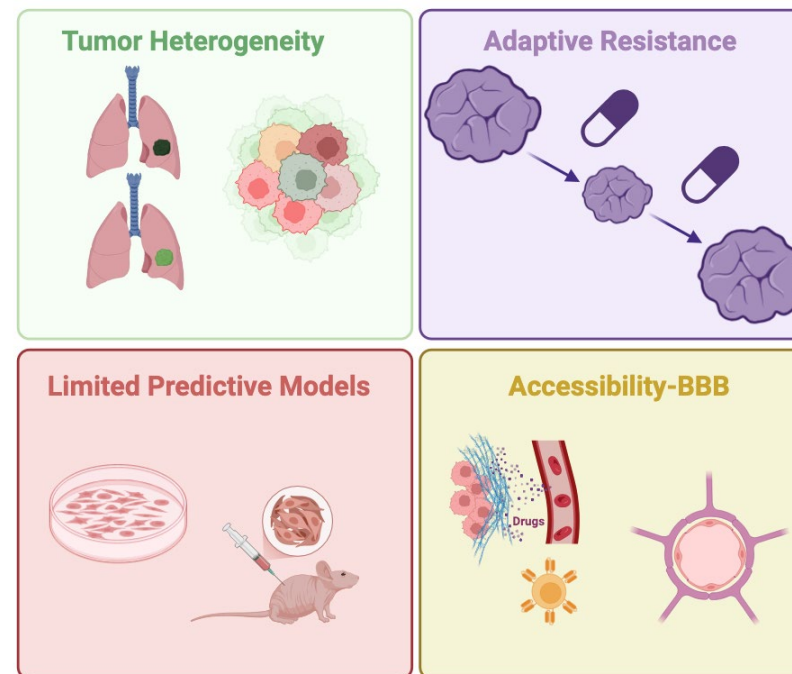
- Why do oncology drugs fail in the clinic?
- Why do we need better cell culture models, especially for childhood brain tumors?
- What are better (next-generation) cell culture models?
- BRAF V600E-mutant high-grade patient-derived models: a paradigm for precision medicine – insights achieved by next-generation cell culture models
- The Stanford Pediatric Cancer Model Development Center

# Low Prediction Models: Barrier for Oncology Drug Development

## 1 Oncology drug development attrition rates



## 2 Lack of efficacy causes for oncology drugs



**~50-60%** of drugs fail in the clinic due to lack of efficacy  
FDA Approval can take up to 13 years and cost up to \$2.5b

# Advantages of Human Next-Generation Cell Culture Models

## 1 Conventional Cell Culture Models

Tumor/Healthy tissue



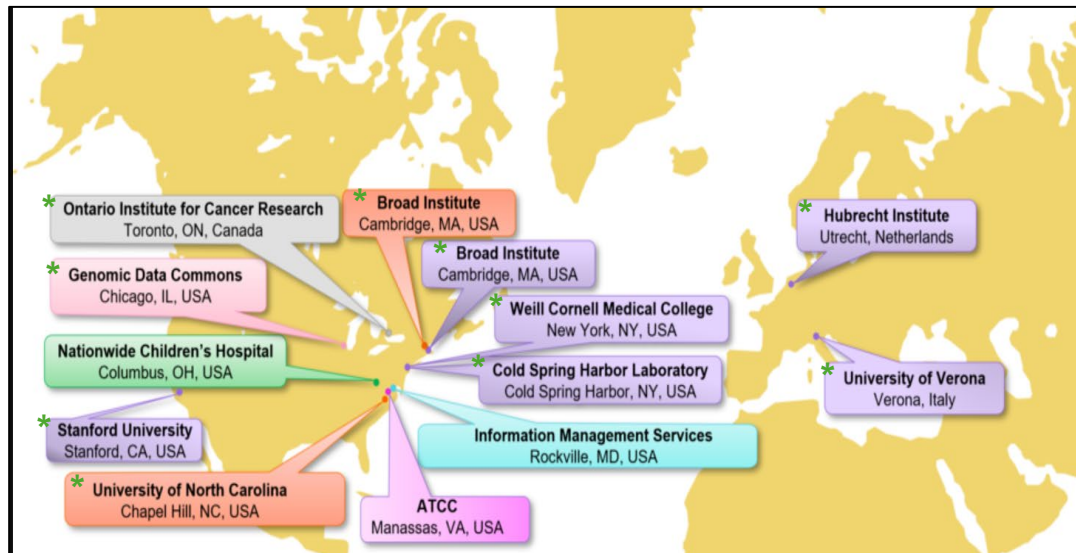
*(monolayer, 2-dimensional (2D) cultures on plastic)*

- Cells plated on flat **plastic surfaces**
- Cells grown in high levels of animal **serum**
- Genetic and transcriptomic **drift**
- Lack stemness, **tumor heterogeneity and plasticity**
- **Overpredict drug efficacy**
- Often lacking clinical information of **parental tumor origin, molecular characterization missing**

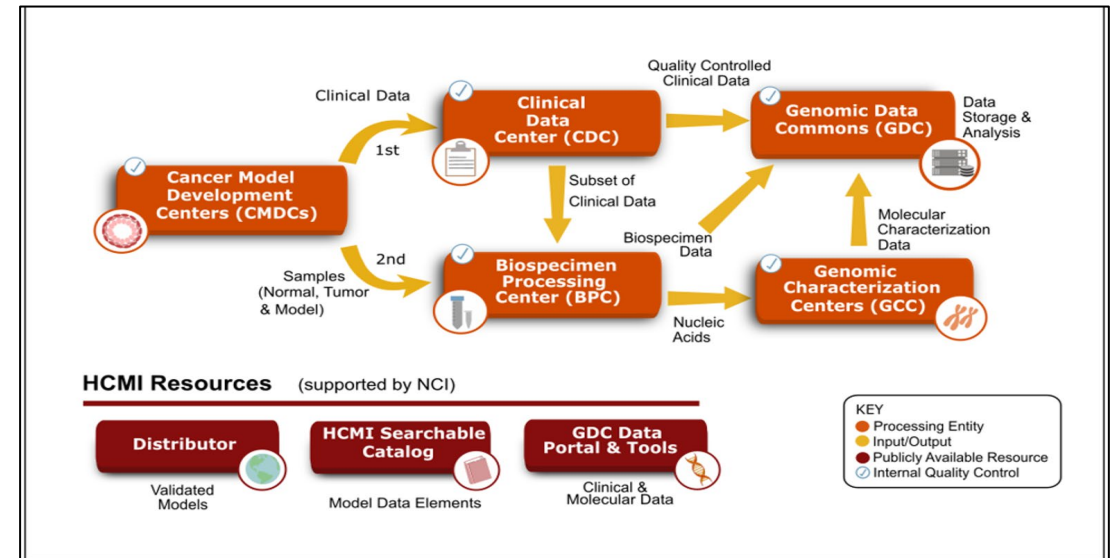
→ Simple, scalable, **low physiological relevance**  
**application: e.g., early stages of drug screening**

# The Human Cancer Model Initiative (HCMI)

The HCMI is an international consortium founded by the **National Cancer Institute** and dedicated to generating next-generation, patient-derived cancer models as a community resource to facilitate cancer research



(Source: <https://www.cancer.gov/ccg/research/functional-genomics/hcmi/about/cancer-model-development>)

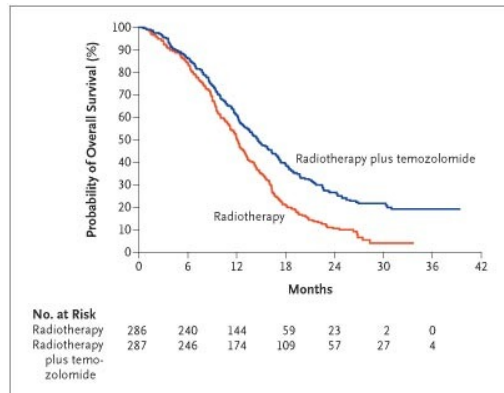


(Source: <https://ocg.cancer.gov/programs/hcmi/nci-cancer-model-development>)

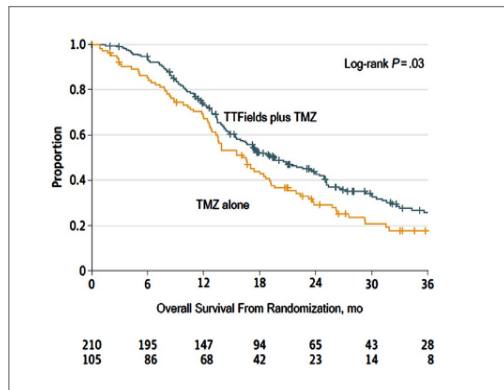
(\* Managed by the Frederick National Laboratory for Cancer Research (FNLCR), Leidos Biomedical Research, Inc.)

# Glioblastoma: Survival with SoC and Drug Approvals

## 1 Survival rates with SoC

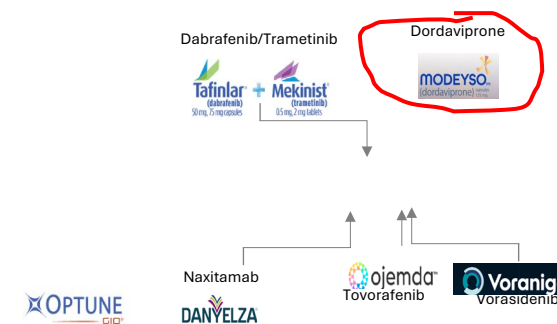


Stupp *et al*, Lancet 2005, PMID15758009



## 2 Drug Approvals Lung Cancer vs. Gliomas

Carmustine  
Lomustine



Glioblastoma/HGG SoC has minimal effect on survival (med surv = 15.4 mos) with >90% recurrence rate  
 One drug approval for high-grade gliomas in 10 years  
 Clinical need for new therapeutics is high

# Challenges for Development of Therapeutics for High-Grade Glioma

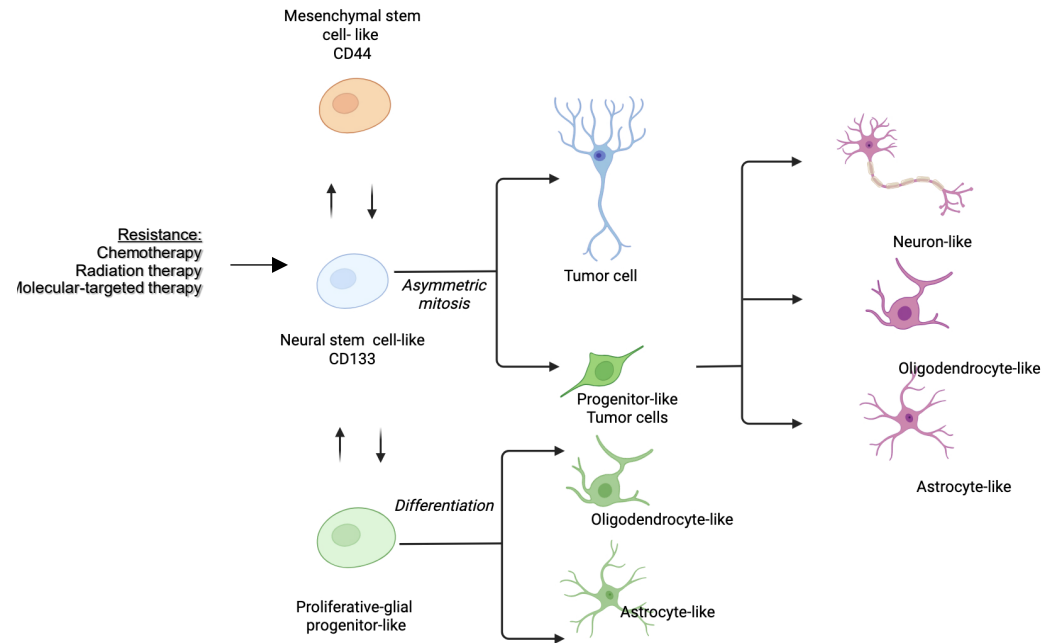
## 1 Key challenges malignant brain tumor treatment

### Tumor Heterogeneity

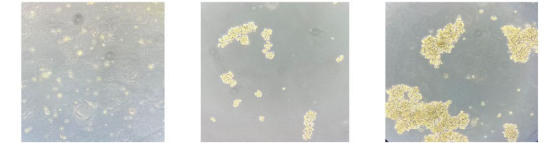
HGG contains multiple genetically distinct clones. Targeting one subpopulation allows resistant clones to repopulate.

### Therapy Resistance, Plasticity, Glioma Stem Cells

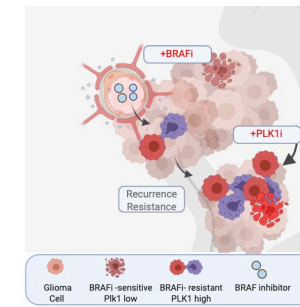
A subpopulation of cells is resistant to radiation and chemotherapy, showing plasticity, driving tumor re-initiation after treatment.



## 2 Stem cell-like features of malignant brain tumors



Morphology of an epitheloid glioblastoma tumor 3D spheroid cell line HCM-STAN-1297-C71. Morphology: left. 2 h after thawing; center. at low density; right. at high density.



Lerner *et al.* Cancer Res 2015, PMID:PMC4698003

Several key features of malignant brain tumors contribute to failed efficacy

Next-generation models recapitulate key features of malignant brain tumors

Malignant brain tumors are very diverse and have many subtypes – model panels

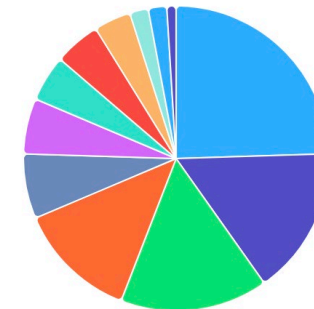
# Majority of Current HCMI Models are for Adult Cancers

## 1 Human Cancer Models Initiative Representation

332

Only 12 pediatric cancer models in the HCMI collection

Childhood Cancer Incidence, Age 0-19, 2017-2021



## 2 Models for Brain Malignancies in the HCMI catalogue

**Human Cancer Models Initiative Searchable Catalog**  
Model: HCM-BROD-0106-C71

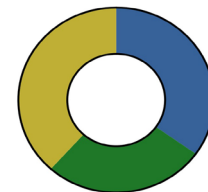
**MODEL DETAILS**  
3-D: Other (e.g., neurosphere, air-liquid interface, etc.)  
Split Ratio: 1:2  
Time to Split: N/A  
Doubling Time: N/A  
Tissue Status: Recurrent

**PATIENT DETAILS**  
Gender: Male  
Race: White  
Age At Diagnosis (Years): 52  
Age At Acquisition (Years): 54  
Disease Status: Progressive disease  
Vital Status: Deceased  
Neoadjuvant Therapy: No  
Therapy: -Surgery  
Chemotherapeutic Drug List Available: No  
Clinical Tumor Diagnosis: Glioblastoma  
Histological Subtype: NOS  
Primary Site: Brain  
Acquisition Site: Brain  
Tissue Status: Recurrent  
TNM Stage: N/A  
Clinical Stage Grouping: N/A  
Histological Grade: N/A

**MODEL IMAGES (2)**  
Magnification: x

**REPOSITORY STATUS**  
Data Updated: May 03, 2024  
Date Of Availability: September 26, 2019  
Licensing Required For Commercial Use: Yes  
Date Created: September 27, 2019

**EXTERNAL RESOURCES**  
SEQUENCING FILES CASE METADATA MASKED SCHEMATIC MAP  
DOI: 10.1001/2019.12345



Total=86

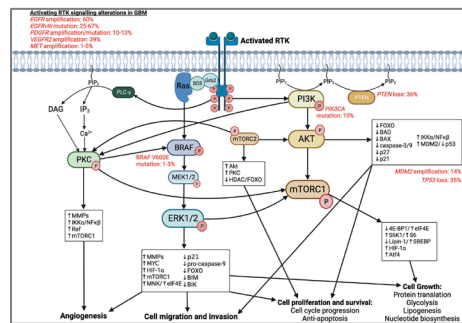


# Pediatric and Adult Brain Cancers Are Fundamentally Different

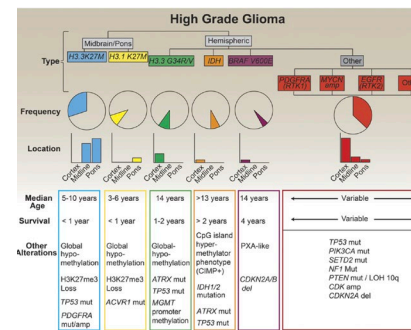
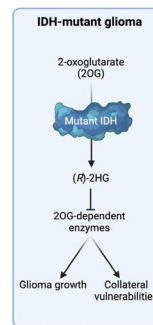
## 1 Pediatric brain cancer characteristics

- Rare (4-5 x lower incidences than in adults)
- Different treatment strategies (minimize long-term sequelae)
- Different pathogenetic mechanisms (developmental, Gliomas: MAPK pathway vs. RTK, IDH1 mut vs BRAF mutations)

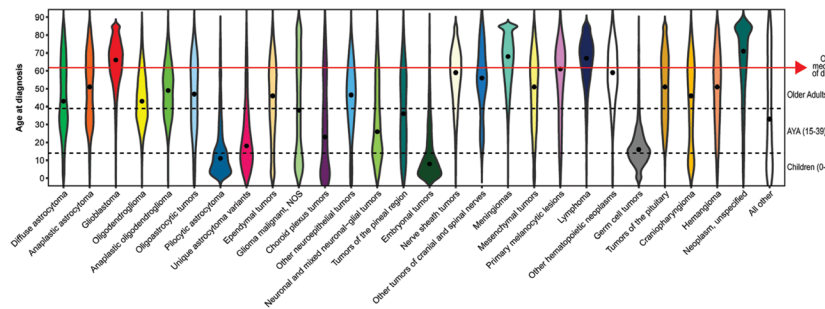
## 2 Common oncogenic pathways of adult glioma



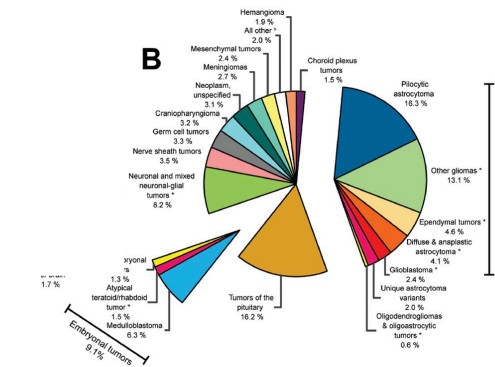
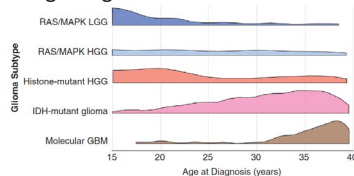
Dewdney et al. Signal Transduct Target Ther 2023  
 PMID:PMC10587102



## 3 Distribution of age at diagnosis by selected primary brain tumors



Low-grade glioma are the most common

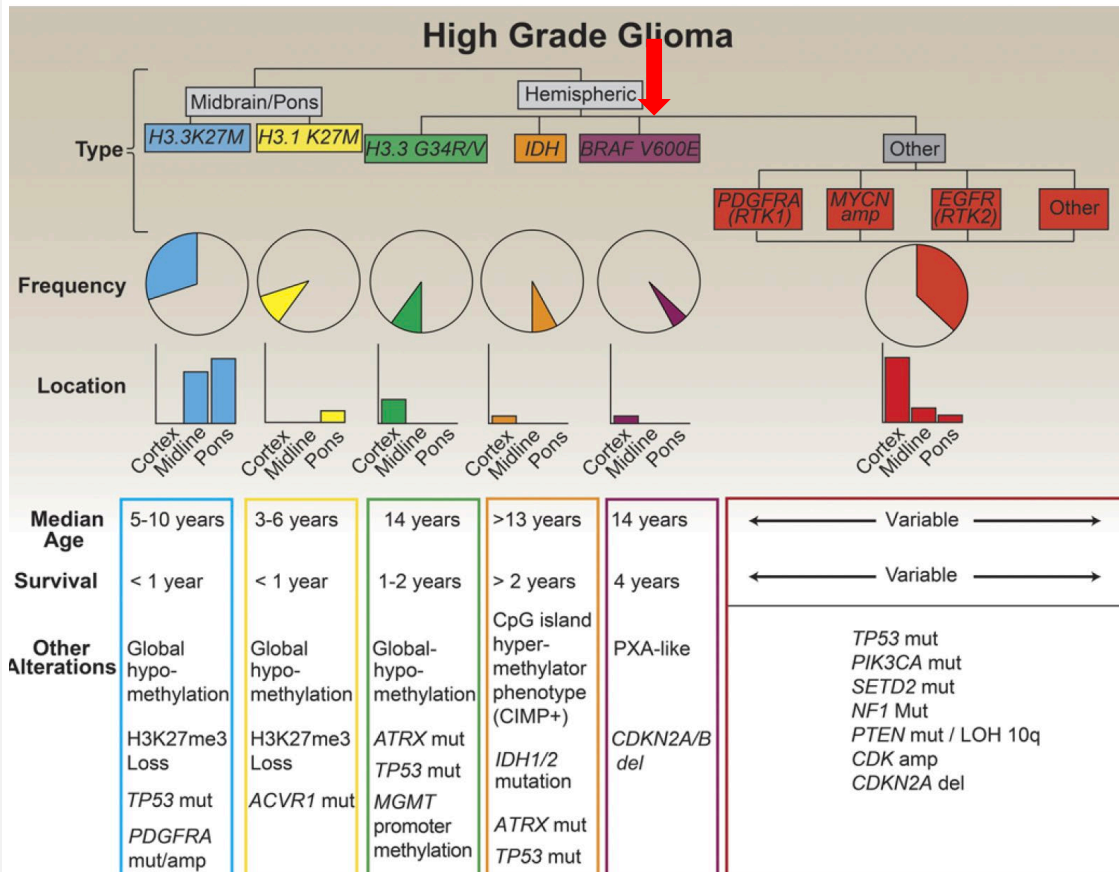


Price et al; Neuro-oncology 2024; <https://doi.org/10.1093/neuonc/noae145>

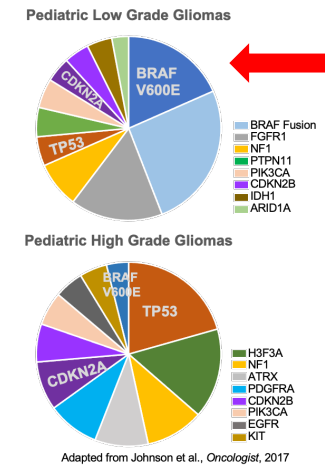
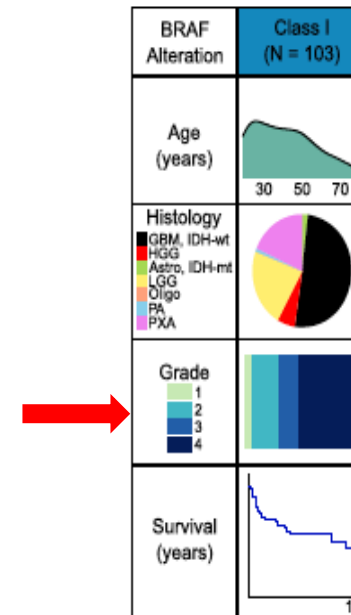
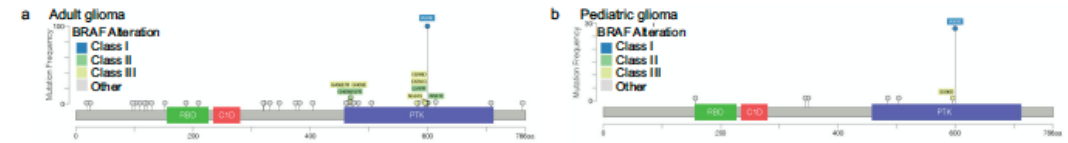
Lim-Fat et al; Neuro-oncology 2025, PMID:PMC11726256

# BRAF V600E-mutant glioblastoma as a paradigm for precision medicine in brain tumors

## 1 BRAF V600E-altered glioma occur in pediatric + adult patients but in different types



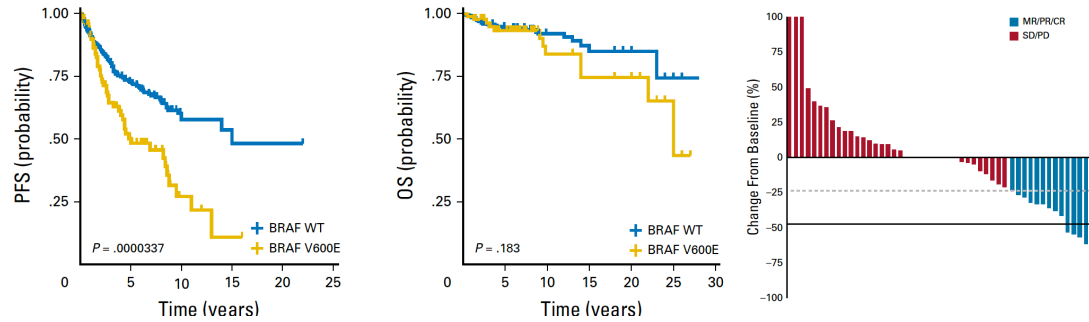
Pollack et al, J Neurosurg Pediatr 2019, PMID:PMC6823600



Schreck et al, NPJ Precis Oncol 2023, PMID:PMC9975216

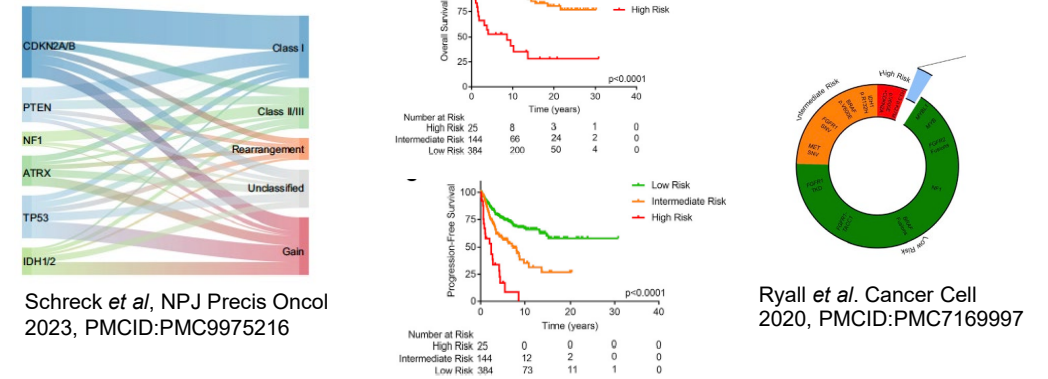
# The BRAF V600E-mutant glioma frequency, types, and prognosis

## 1 BRAF V600E-altered glioma are chemotherapy-resistant



Lassaletta *et al.*, JCO 2017, PMID:PMC5791837

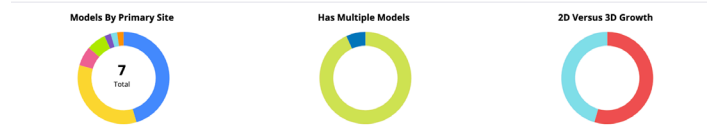
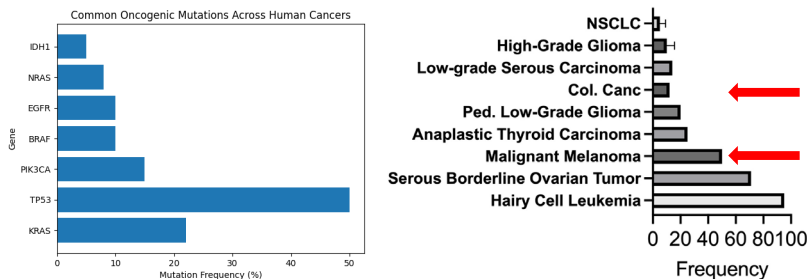
## 2 BRAF V600E co-mutations affect risk for progression



Schreck *et al.*, NPJ Precis Oncol 2023, PMID:PMC9975216

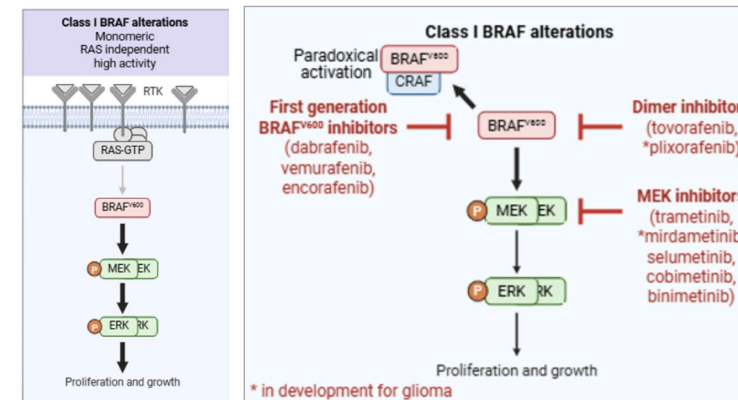
Ryall *et al.* Cancer Cell 2020, PMID:PMC7169997

## 3 BRAF V600E mutations found in various cancer



BRAF V600E models in HCM1 catalogue

## 4 BRAF/MEK inhibitors in the clinical arena



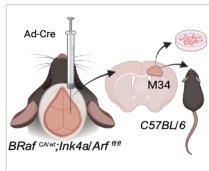
Courtesy of Karisa Schreck



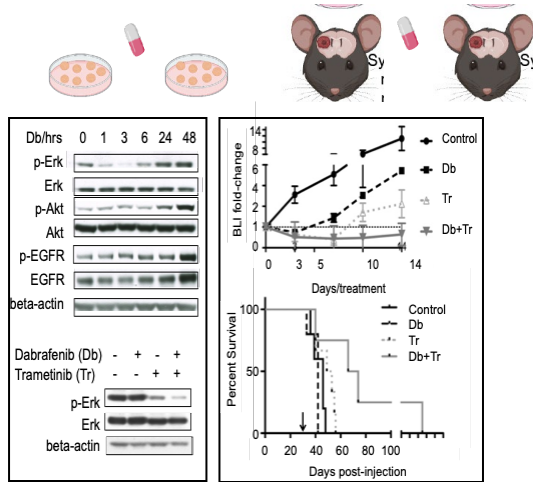
# Research Objectives Based on Low-Response Rates

## 1 MAPK pathway reactivation after BRAF inhibition

Syngeneic Mouse Models For BRAF V600E-mutant High-Grade Glioma:



Grossauer *et al.* Oncotarget, 2016, PMID:PMC5342782



Dabrafenib (DB)=BRAF inhibitor  
Trametinib (Tr)=MEK inhibitor

Understand how BRAF V600E mutated high-grade gliomas respond to clinically relevant, molecular-targeted inhibition (BRAF V600E and MEK inhibition)

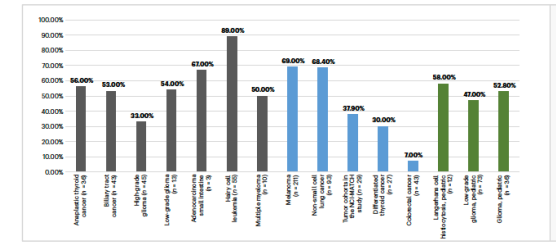
Identified potential mechanisms of therapy escape

Find novel therapeutic opportunities to combine with BRAF V600E- targeted therapy to overcome resistance

## 2 Combined BRAF and MAPK inhibition response rates

|   | Grade III (n=13) | Glioblastoma (n=31) | Age 18-39 years (n=22) | Age >40 years (n=23) |
|---|------------------|---------------------|------------------------|----------------------|
| Objective response rate by investigator, % (95% CI)                     | 38 (13.9-68.4)   | 32 (16.7-51.4)      | 50 (28.2-71.8)         | 17 (5.0-38.8)        |
| Patients responding at 12 months by investigator assessment, % (95% CI) | 100              | 67 (28.2-87.8)      | 89 (43.3-98.4)         | 50 (5.8-64.5)        |
| Median progression-free survival by investigator, months (95% CI)       | 3.8 (1.7-NR)     | 2.8 (1.8-13.7)      | 18.5 (5.5-41.4)        | 1.7 (0.9-2.5)        |
| Median overall survival, months (95% CI)                                | 45.2 (6.3-NR)*   | 13.7 (8.4-25.6)     | 45.2 (17.9-NR)†        | 8.7 (3.7-11.7)       |

Wen *et al.*, The Lancet Oncology 2022, PMID:34838156



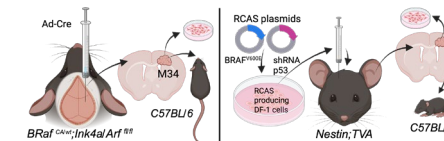
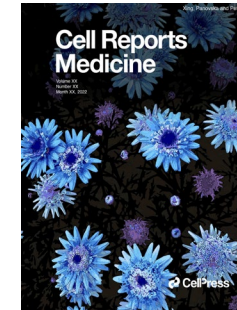
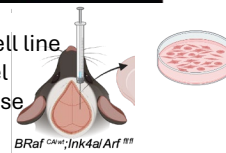
Subbiah *et al.*, Nat Med 2023, PMID:PMC10202803

## 3 Model Development for BRAF V600E-mutant high-grade gliomas

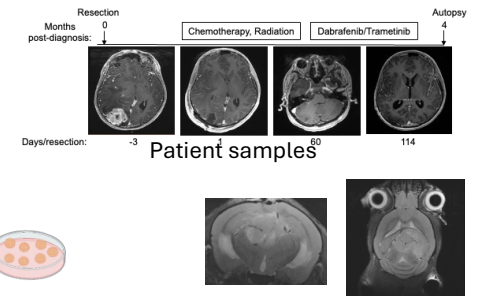
Lerner *et al.* Cancer Res 2015, PMID:PMC4698003



Patient samples  
1 conventional cell line  
1 xenograft model  
1 orthotopic mouse model

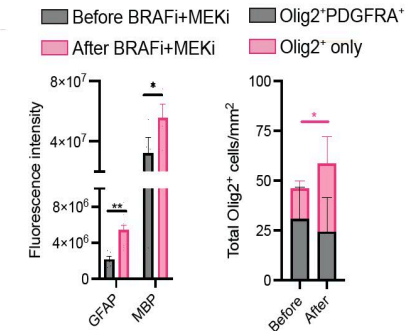
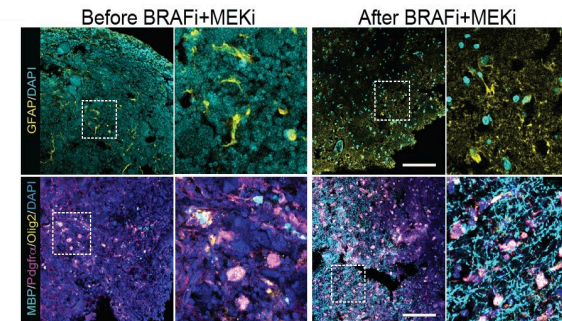
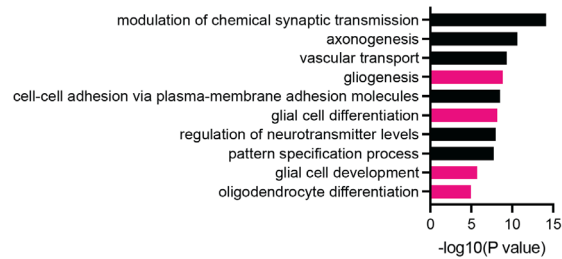
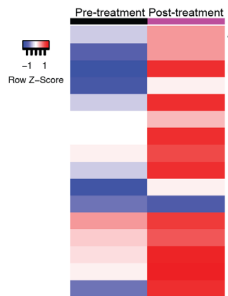
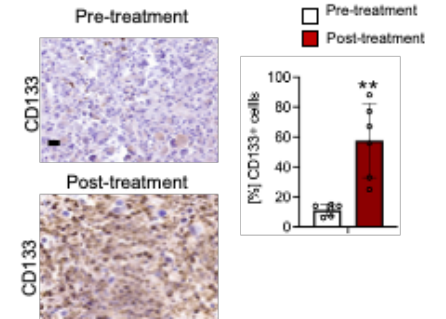
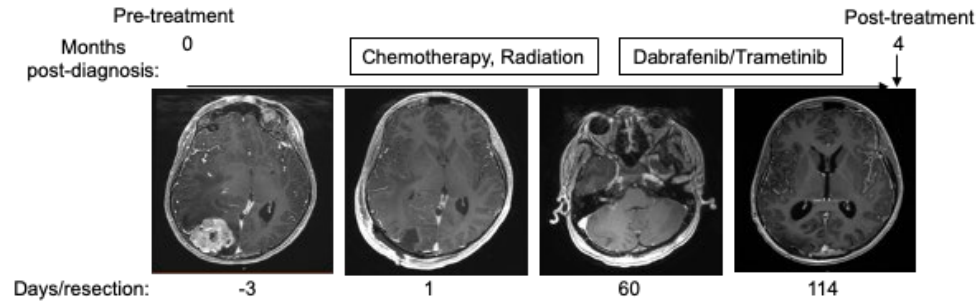
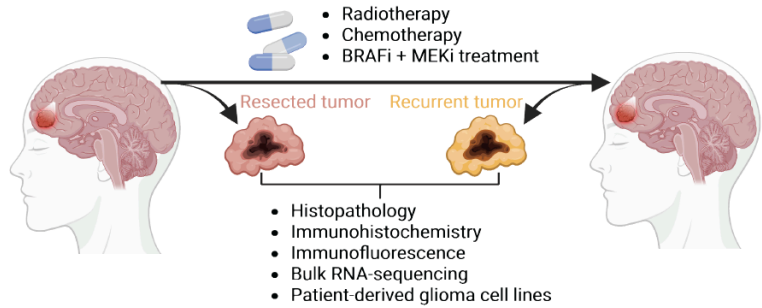


Xing, Panovska *et al.*, Cell Reports Med 2025, PMID:PMC12208339



3 patient-derived spheroid lines  
+ matched patient-derived xenografts

# Analyses of Pre- and Post-treatment BRAF V600E-mutant Glioblastoma Samples

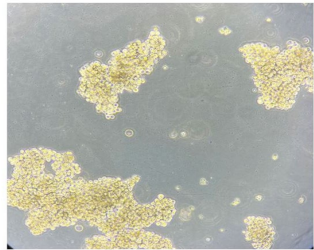


**BRAF/MEK inhibition in patients upregulated not only stem-cell markers but also glial differentiation, indicative of therapy-induced increases in cell plasticity and differentiation**

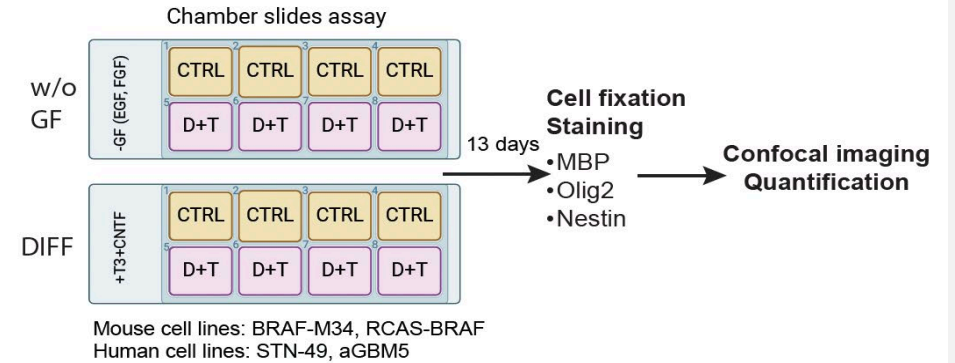
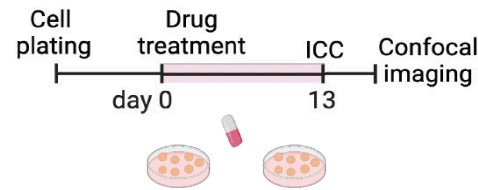
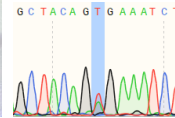
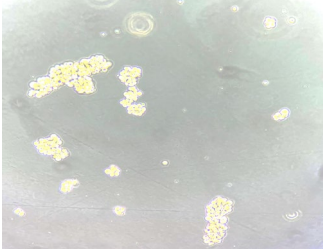
Xing, Panovska et al, Cell Reports Med 2025, PMID:PMC12208339

# Patient-Derived 3D Models Recapitulate Therapy-Induced Cell State Changes

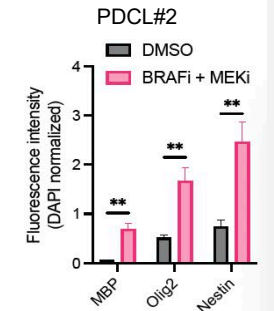
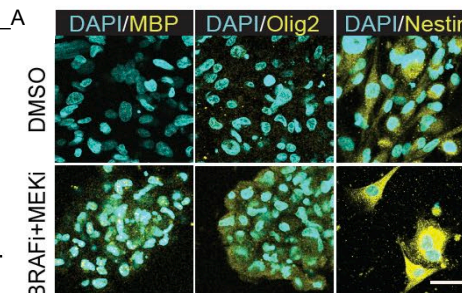
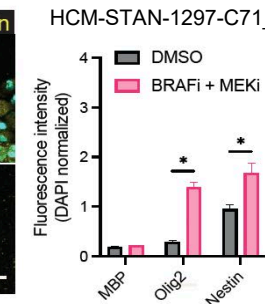
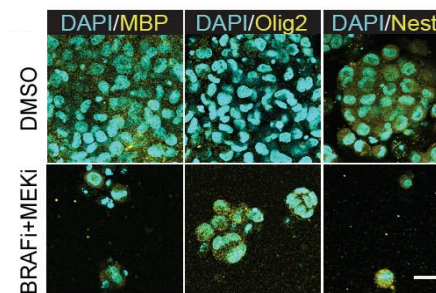
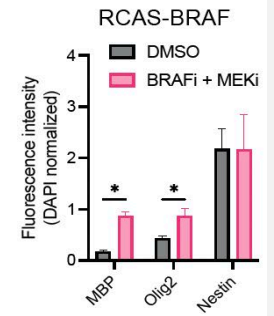
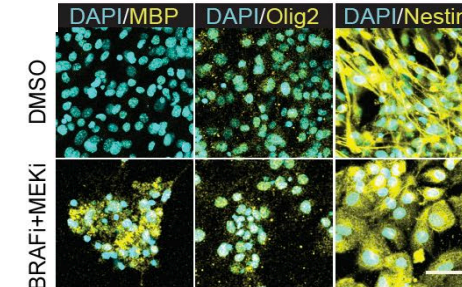
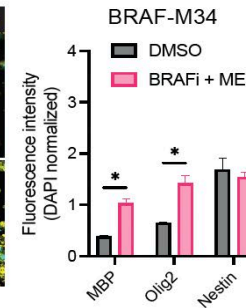
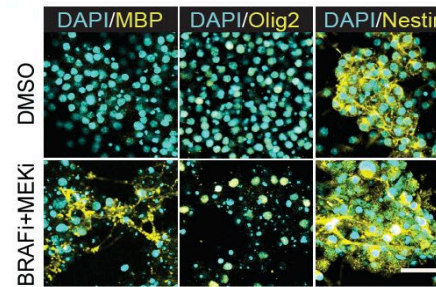
HCM-STAN-1297-C71\_A  
BRAF V600E



Patient-derived cell line  
(PDCL) BRAF V600E #2

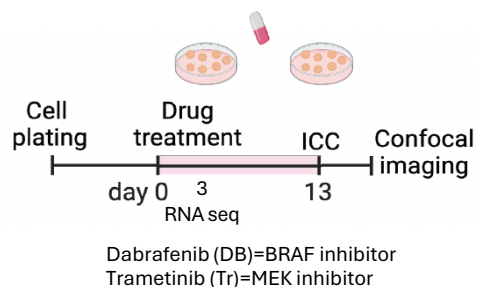


**BRAFi+MEKi  
Induces  
Stem Cells and  
Glial Differentiation  
States**

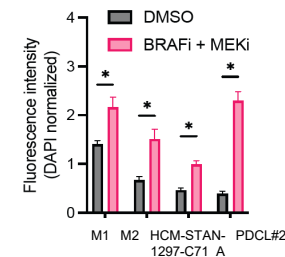
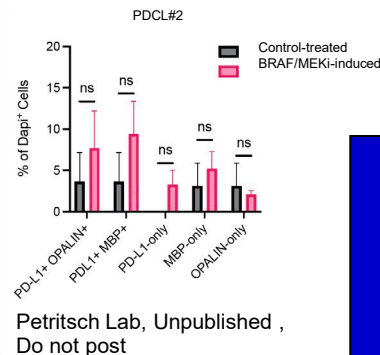
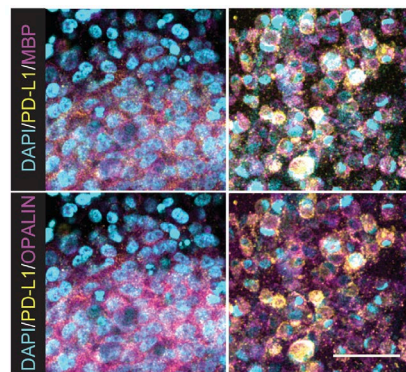
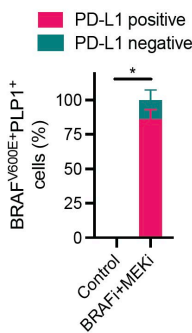
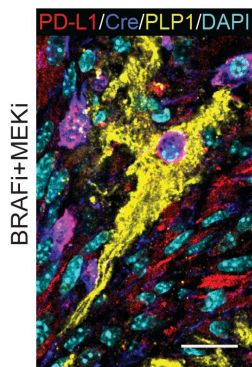
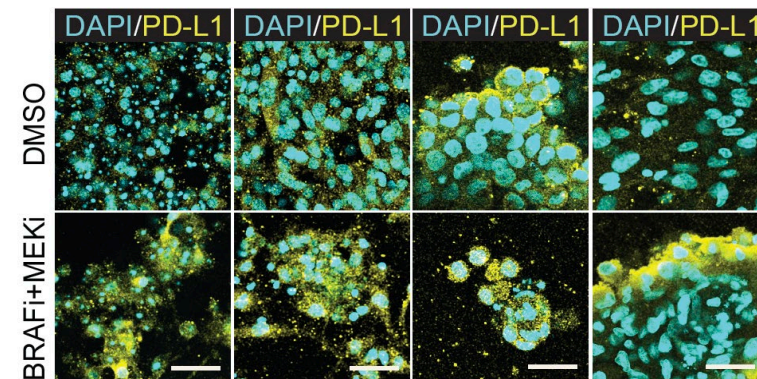
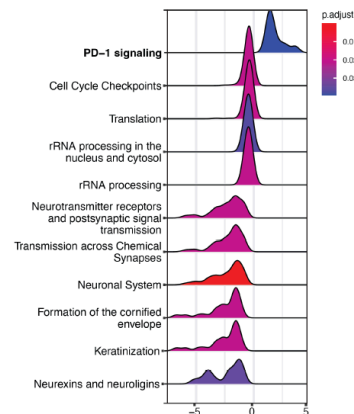
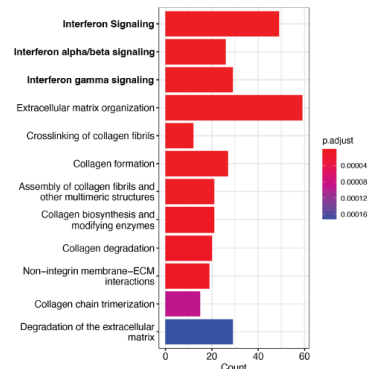


BRAF/MEK inhibition upregulated not only stem-cell markers but also glial differentiation, in cell culture-based assays, indicative of direct effects on tumor cell plasticity and differentiation

# Immune-modulatory Effects of BRAF/MEK inhibitors?



Xing, Panovska et al, Cell Reports Med 2025, PMID:PMC12208339

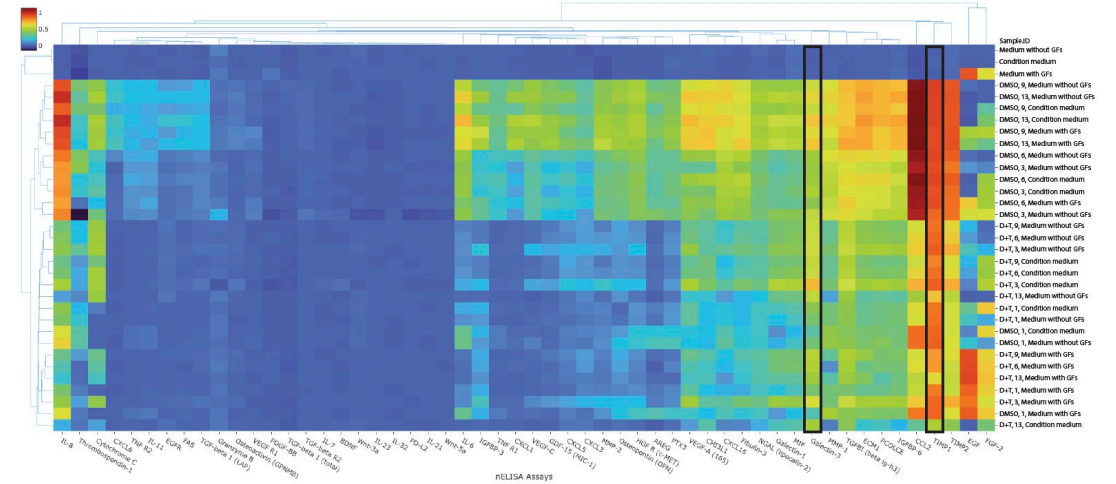
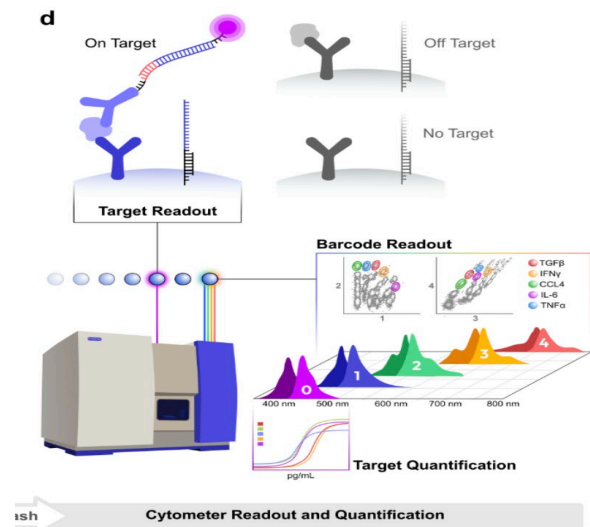
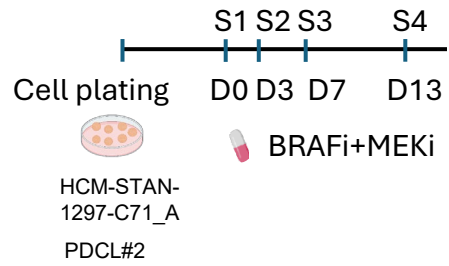


**BRAF+MEK inhibitors induced glioma cell differentiation along with upregulated antigen presentation & up-expression of immune checkpoint inhibitors**

Xing, Panovska et al, Cell Reports Med 2025, PMID:PMC12208339

# Multi-plex ELISA to Investigate BRAF/MEK inhibitor-Induced Immunoregulation

## Nomic - ELISA



Xing, Panovska et al, Cell Reports Med 2025, PMID:PMC12208339

## Nomic - ELISA

Rel. Factor Expression

HCM-STAN-1297-C71\_A

Rel. Factor Expression

PDCL#2

Factor Expression

HCM-STAN-1297-C71\_A

## RNA-seq

Factor Expression

PDCL#2

BRAF+MEK inhibitors elevate secretion of pro-inflammatory cytokines and T cell inhibitory factors

# Summary

## MAIN FINDINGS

- BRAF/MEK inhibition induces **glioma cell state** increases differentiation and simultaneously immune evasion
- BRAFi+MEKi activates the interferon response and anti-tumor immunity, while simultaneously **suppressing T cells via PD-L1 upregulation in glial cells**
- Glial differentiation and immune evasion could be mediated by therapy-induced **immune modulatory secretome**
- High PD-L1 expression in BRAF-mutant GBM provides a criterion for anti-PD-1 therapy
- **Concurrent BRAF/MEK and checkpoint inhibition** enhances anti-tumor immunity and survival

## CLINICAL IMPLICATION

Our preclinical findings highlight the potential of integrating BRAFi+MEKi treatment with ICI, with emphasis on concurrent treatment

# Common Pediatric Brain Tumor Subtypes

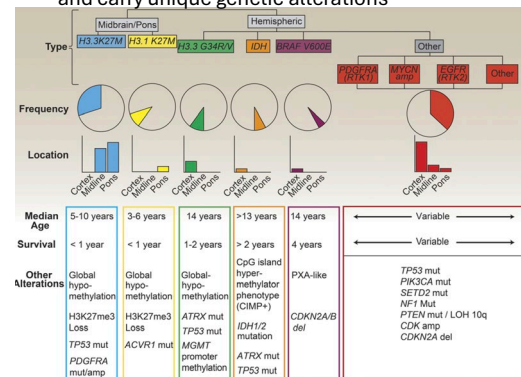
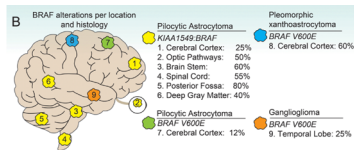
>100 subtypes of pediatric solid tumors

Pfister, SM et al; Cancer Discov. 2022; PMID:PMC9401511

Subtypes of most common primary pediatric brain tumors

## Glioma

Pediatric high-grade glioma are rare but devastating and carry unique genetic alterations

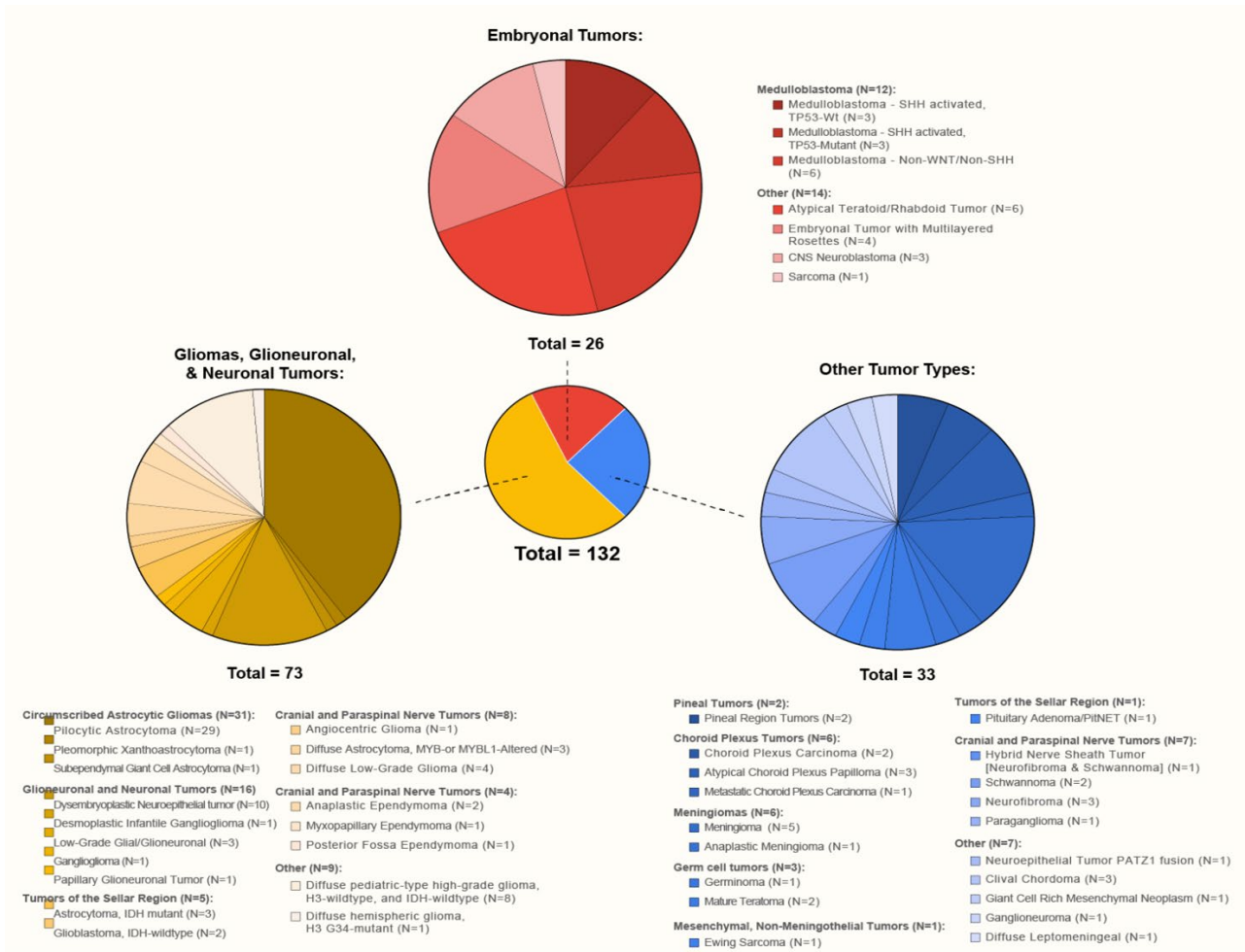


## Embryonal Tumors

### Medulloblastoma

| Subgroup            | WNT                | SHH                                | Group 3  | Group 4  |
|---------------------|--------------------|------------------------------------|--|--|
| Incidence           | 10%                | 30%                                | 25%  | 35%  |
| Subtype             | WNT α              | WNT β, SHH α, SHH β, SHH γ, SHH δ  | Group 3a, Group 3b, Group 3γ, Group 4a, Group 4b | Group 4c, Group 4d                                   |
| Gender              | ♂:♀                | ♂:♀                                | ♂:♀  | ♂:♀  |
| Subtype proportion  | α, β               | α, β, γ, δ                         | 3a, 3b, 3γ                                       | 4c, 4d   |
| Age                 | 3-17               | >10, 3-17, 0-3, 0-3                | >17, 0-10  | 3-17, 0-10, 3-17, 3-17                               |
| Metastases          | 9%                 | 21%, 20%, 33%, 9%                  | 9%, 43%  | 20%, 40%, 40%, 40%                                   |
| 5 year survival     | 97%                | 100%, 70%, 70%, 90%                | 90%, 65%   | 55%, 40%, 65%, 75%, 80%                              |
| Copy Number Changes | 6                  | MYCN amp, GLI3 amp, VAF amp        | PTEN Loss  | Balanced genome, 10q22, 7p, 8, 10, 11q23.3, 11, 11q7 |
| Other events        | TP53 mutations     | TP53 mutations                     | High OPF1/1B expression                          | MYC amp, MYCN amp, CDK8 amp, SHC4P dup, CDK8 amp     |
| Histology           | Classic, LCA(rare) | Desmoplastic, Nodular Classic, LCA | Extensive nodularity                             | Classic, LCA   |
| Microscopy          | 40x                | 20x                                | 10x  | 20x  |

# Representative Cohort of Pediatric Brain Cancer Types at Stanford Pediatrics CMDC

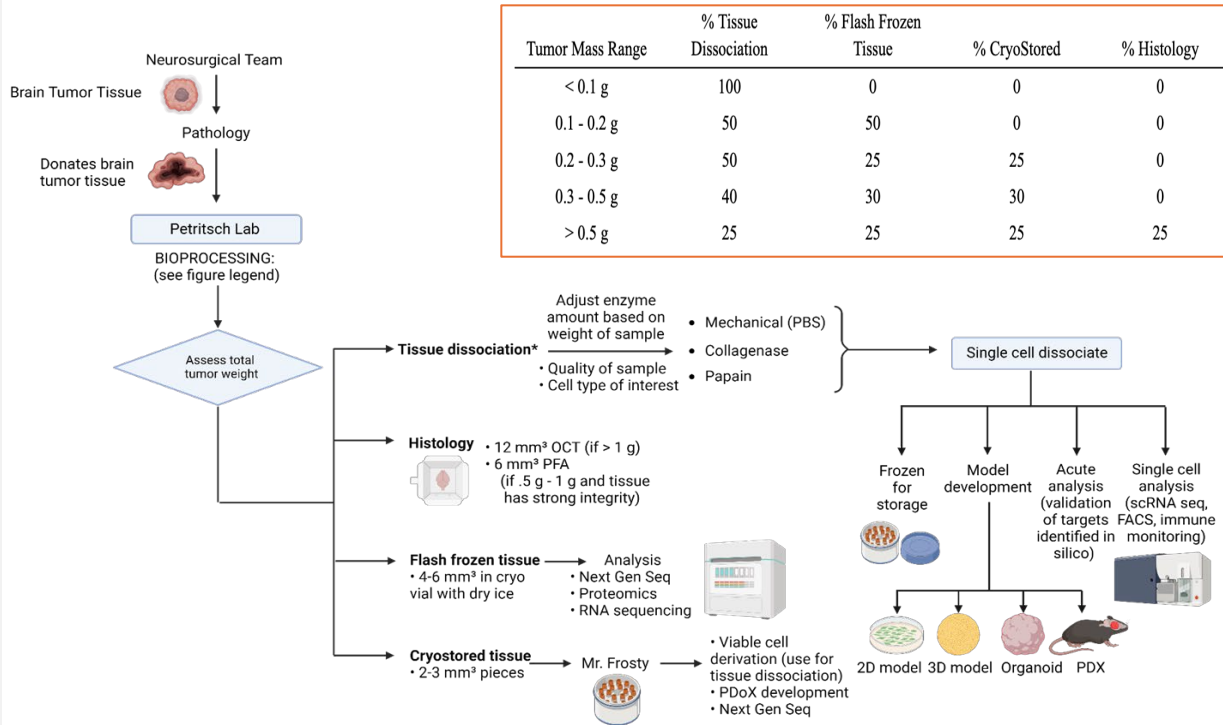


| No. of Banked Cases | Tumor Type   | Genes/Molecular Profiles Characteristically Altered  |
|---------------------|--|--|
| 3                   | Astrocytoma, IDH-mutant  | IDH1, IDH2, ATRX, TP53, CDKN2A/B   |
| 2                   | Glioblastoma, IDH-wildtype   | IDH-wildtype, TERT promoter, chromosomes 7/10, EGFR  |
| 2                   | Diffuse astrocytoma, MYB-or MYBL1-altered                              | MYB, MYBL1   |
| 1                   | Angiocentric glioma  | MYB  |
| 4                   | Diffuse low-grade glioma, MAPK pathway-altered                         | FGFR1, BRAF  |
| 1                   | Diffuse hemispheric glioma, H3 G34-mutant                              | H3 G34, TP53, ATRX   |
| 9                   | Diffuse pediatric-type high-grade glioma, H3-wildtype and IDH-wildtype | IDH-wildtype, H3-wildtype, PDGFRA, MYCN, EGFR (methylome)  |
| 30                  | Pilocytic astrocytoma  | KIAA1549-BRAF, BRAF, NF1   |
| 1                   | Pleomorphic xanthoastrocytoma  | BRAF, CDKN2A/B   |
| 1                   | Subependymal giant cell astrocytoma                                    | TSC1, TSC2   |
| 2                   | Chordoid glioma  | PRKCA  |
| 1                   | Ganglion cell tumors   | BRAF   |
| 10                  | Dysembryoplastic neuroepithelial tumor                                 | FGFR1  |
| 1                   | Papillary glioneuronal tumor   | PRKCA  |
| 1                   | Diffuse leptomeningeal glioneuronal tumor                              | KIAA1549-BRAF fusion, 1p (methylome)   |
| 3                   | Supratentorial ependymomas   | ZFTA, RELA, YAP1, MAML2  |
| 1                   | Posterior fossa ependymomas  | H3 K27me3, EZHIP (methylome)   |
| 6                   | Medulloblastoma, SHH-activated   | TP53, PTCH1, SUFU, SMO, MYCN, GLI2 (methylome)   |
| 6                   | Medulloblastoma, non-WNT/non-SHH                                       | MYC, MYCN, PRDM6, KDM6A (methylome)  |
| 6                   | Atypical teratoid/rhabdoid tumor                                       | SMARCB1, SMARCA4   |
| 5                   | Embryonal tumor with multilayered rosettes                             | C19MC, DICER1  |
| 3                   | CNS neuroblastoma, FOXR2-activated                                     | FOXR2  |
| 6                   | Meningiomas  | NF2, AKT1, TRAF7, SMO, PIK3CA; KLF4, SMARCE1, BAP1 in subtypes; H3K27me3; TERT promoter, CDKN2A/B in CNS WHO grade 3 |

- ~90% of cases yield tissue for research
- 45 subtypes of brain cancer and 12 subtypes of non-CNS cancers captured over a 4-year period
- Longitudinal collection is crucial to capture diverse, rare cancers

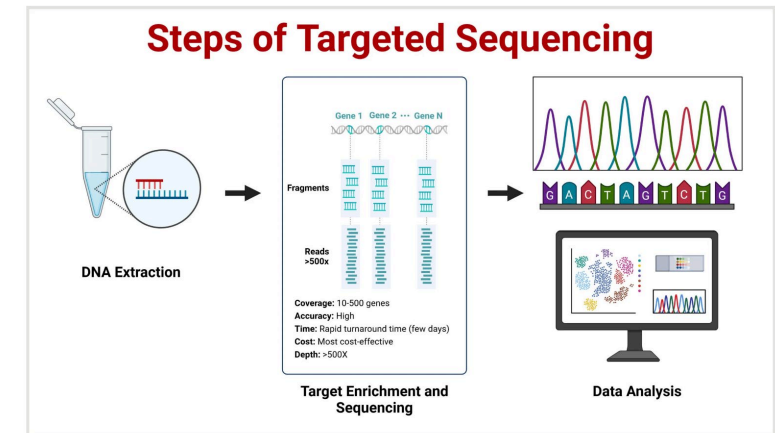
# Pediatric Cancer Model Development - Workflow

## Established Bioprocessing Workflow in our laboratory



| Tumor Mass Range | % Tissue     |        | % Flash Frozen |             |
|------------------|--------------|--------|----------------|-------------|
|                  | Dissociation | Tissue | % CryoStored   | % Histology |
| < 0.1 g          | 100          | 0      | 0              | 0           |
| 0.1 - 0.2 g      | 50           | 50     | 0              | 0           |
| 0.2 - 0.3 g      | 50           | 25     | 25             | 0           |
| 0.3 - 0.5 g      | 40           | 30     | 30             | 0           |
| > 0.5 g          | 25           | 25     | 25             | 25          |

## Established Mutation Identification



Stanford Actionable Mutation Panel ~250 genes, UCSF 500 Caris (WES)

An established biobanking workflow with standardized targeted sequencing of patient Tumors to identify recurrent mutations, including oncofusions

# Pediatric Cancer Types for HCMI

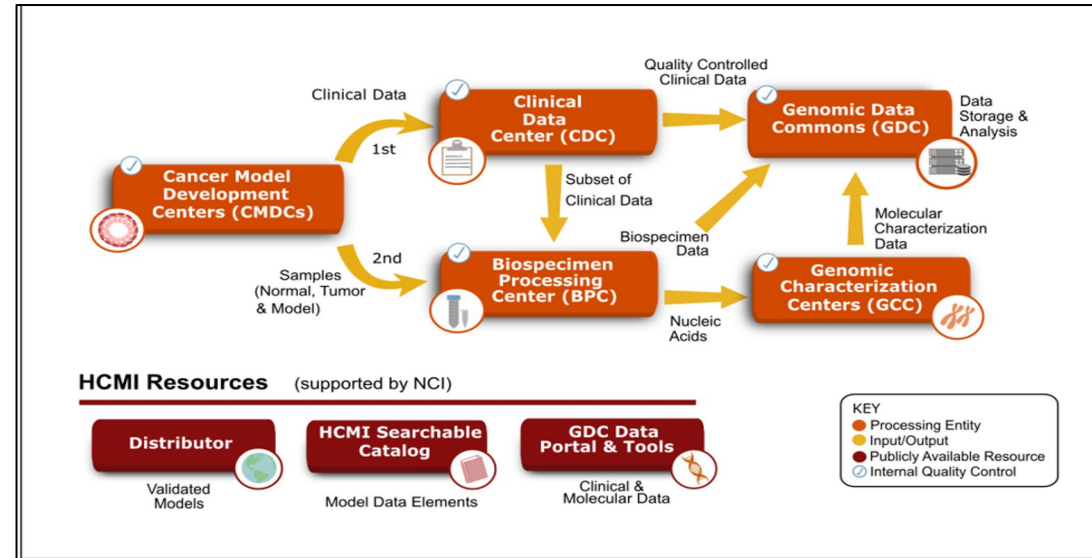
## ■ Pediatric central nervous system (CNS) solid tumors

- Diffuse Midline Glioma (DMG), H3-K27-mutant
- Diffuse High-Grade Glioma (HGG), MAPK activated
- HGG, H3 and IDH wildtype
- Glioblastoma
- Anaplastic Astrocytoma, BRAF V600E-mutant
- Astrocytoma, IDH-mutant
- Medulloblastoma, ATRT
- Ependymoma
- Ganglioneuroblastoma
- ETMR
- Choroid plexus carcinoma
- Pineoblastoma

## ■ Non-CNS pediatric solid tumors

- **Malignant Rare Soft Tissue and Bone sarcoma**
- **Wilms' Tumor (Primary/Metastasis Pair)**
- **Neuroblastoma**
- Hepatoblastoma

60-70% success rate for model development  
27 subtypes of brain cancer and 4 of non-CNS cancer  
were captured in models  
WES/WGS/RNAseq/DNA Methylation



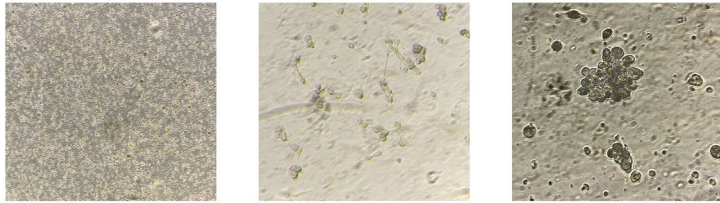
(Source: <https://ocg.cancer.gov/programs/hcmi/nci-cancer-model-development>)

Planning and management oversight by the team at Frederick National Laboratory for Cancer Research, Leidos Biomedical Research, Inc.

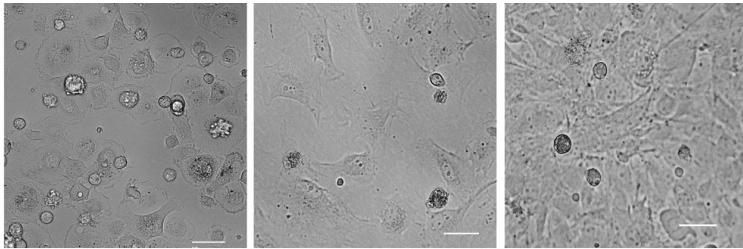
This project has been funded in part with federal funds from the Childhood Cancer Data Initiative (CCDI), National Cancer Institute, National Institutes of Health, Task Order numbers 75N91020F00035, under contract no. 75N91019D00024.



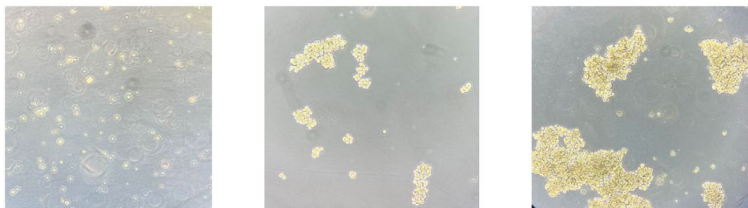
# Examples next-generation models of pediatric solid cancer



Morphology of a Giant cell tumor line HCM-STAN-1408-C71. Morphology. **left.** 2 h after thawing; **center.** at low density; **right.** at high-density.

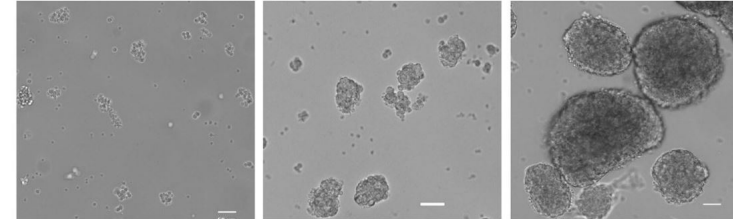


Morphology of a pediatric diffuse anaplastic Wilms tumor 2D adherent cell line HCM-STAN-1353-C64. (A). Morphology. **A.** 2 h after thawing; **B.** at low density; **C.** at high density. Scale bars are 50 μM.



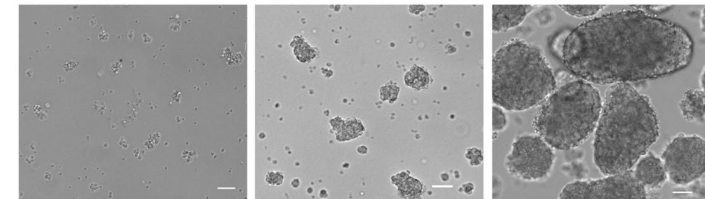
Morphology of an epithelioid glioblastoma tumor 3D spheroid cell line HCM-STAN-1297-C71. Morphology. **left.** 2 h after thawing; **center.** at low density; **right.** at high density.

HCM-STAN-1351-C71-A



Morphology of Neuroblastoma 3D spheroid cell lines generated from bone marrow infiltrated tumor cells of a pediatric MYCN-amplified neuroblastoma patient at diagnosis (HCM-STAN-1351-C71-A) **left.** 2 h after thawing; **center.** at low density; **right.** at high density.

HCM-STAN-1351-C71-B



Morphology of Neuroblastoma 3D spheroid cell lines generated from bone marrow infiltrated tumor cells of a pediatric MYCN-amplified neuroblastoma patient at relapse (HCM-STAN-1351-C71-B). **left.** 2 h after thaw; **center.** at low density; **right.** at high density. Scale bars are 50 μM.



Morphology of diffuse intrinsic pontine glioma 3D spheroid cell line HCM-STAN-1420-C71. **A.** 2 h after thawing; **B.** at low density; **C.** at high density. Scale bars are 5000 μM.

# Stanford Pediatric CMDC Team and Partners \_Acknowledgements

## Stanford Pediatric CMDC

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**CDC:** Megan Stine

**ATCC:** Carolina Lucchesi

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**Research Inc:** Rachana Agarwal, Conrado Soria

**The HCMD Network**



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Stanford  
Cancer Institute





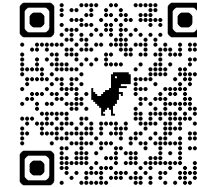
# Resources to learn more about ATCC and the HCMI



Browse and search unreleased HCMI models at ATCC

- Use the “Submit your Input” button on the HCMI Landing page

[www.atcc.org/hcmi-input](http://www.atcc.org/hcmi-input)



HCMI Landing page  
[atcc.org/hcmi](http://atcc.org/hcmi)

HCMI Searchable Catalog  
<https://hcmi-searchable.catalog.nci.nih.gov>

NCI Genomic Data Commons  
<https://portal.gdc.cancer.gov/projects/HCMI-CMDC>

# Presenters



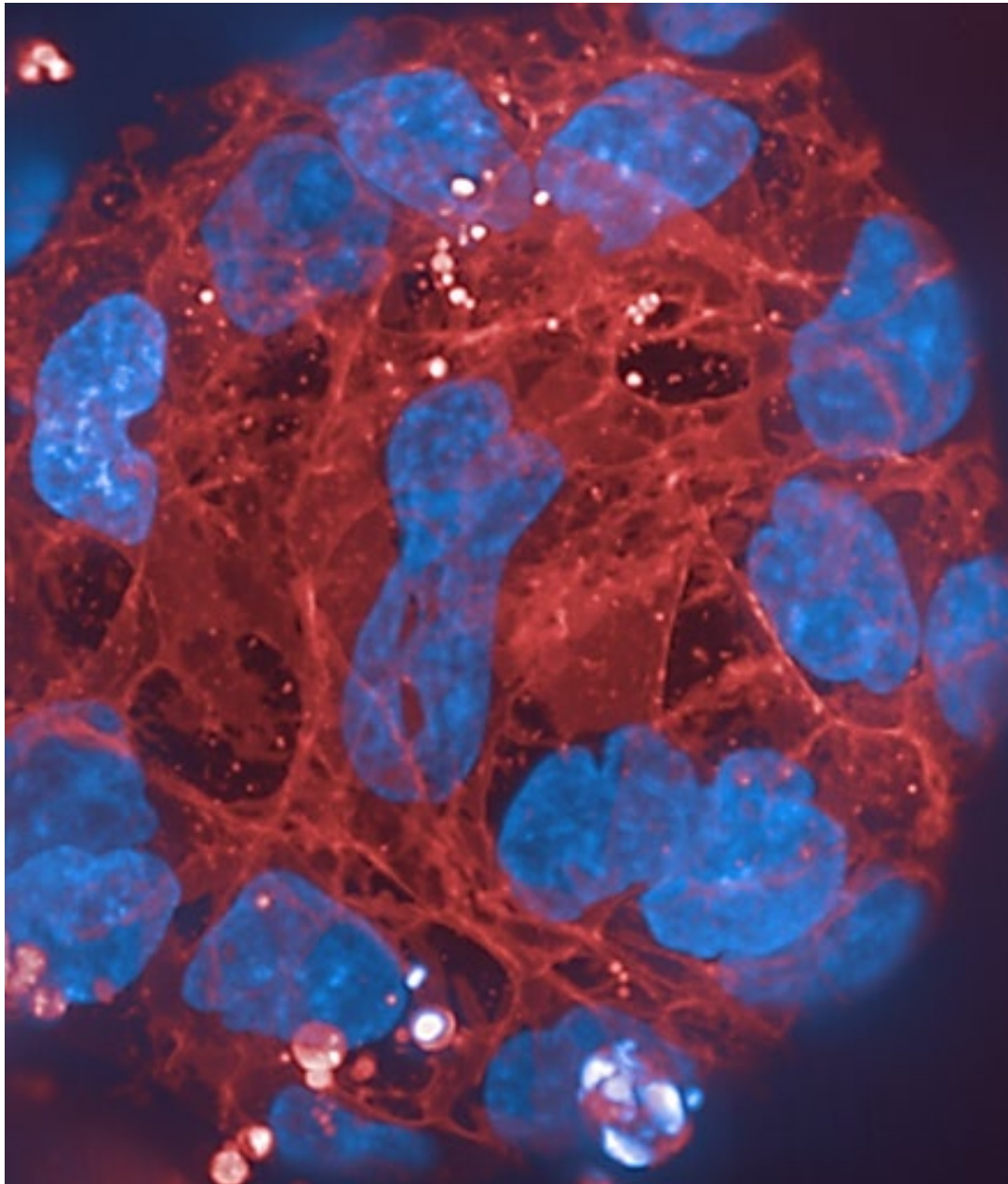
**Carolina Lucchesi, PhD**  
Principal Scientist, Head of  
Microphysiological Systems, ATCC



**Claudia K. Petritsch, PhD**  
Associate Professor in Research,  
Director Pediatric Cancer Model  
Development Center, Sr. Scientist in  
Neuroscience, Stanford University



**Benjamin David Hopkins, PhD**  
Assistant Professor of Research in  
Systems and Computational  
Biomedicine, Weill Cornell Medical  
College



## **Patient derived tumor organoid for therapeutic modeling in cancer**

*Benjamin D Hopkins, PhD*

*Director Tumor Organoid Platform  
Englander Institute  
for Precision Medicine*

# Disclosure Information

## Benjamin D. Hopkins

I have the following relevant financial relationships to disclose:

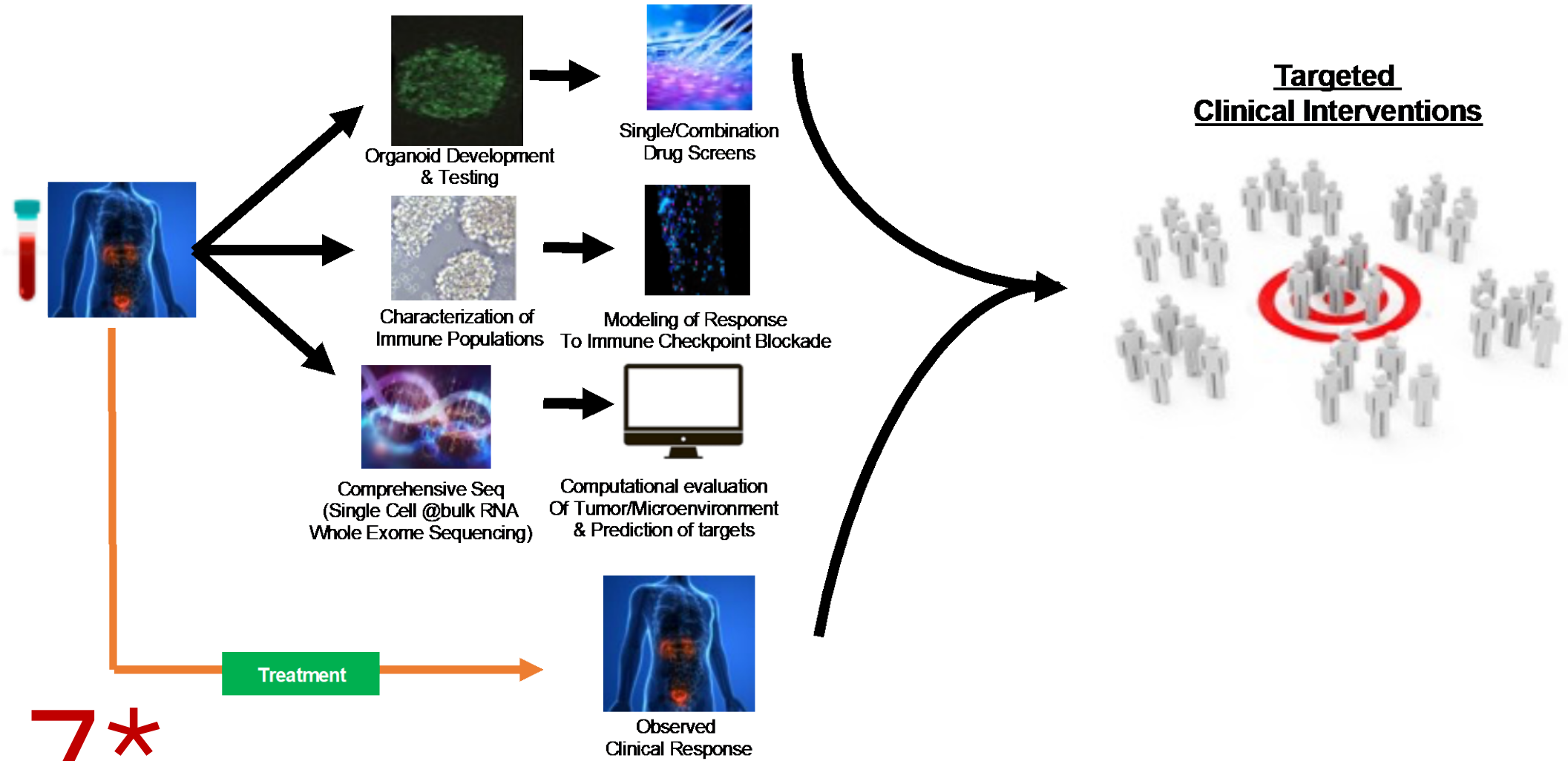
Consultant for: Faeth Therapeutics

Grant/Research support from: Faeth, Novartis, Astra Zeneca, Jazz, Sanofi, Amgen

Stockholder in: Faeth Therapeutics/Sensei Biotherapeutics

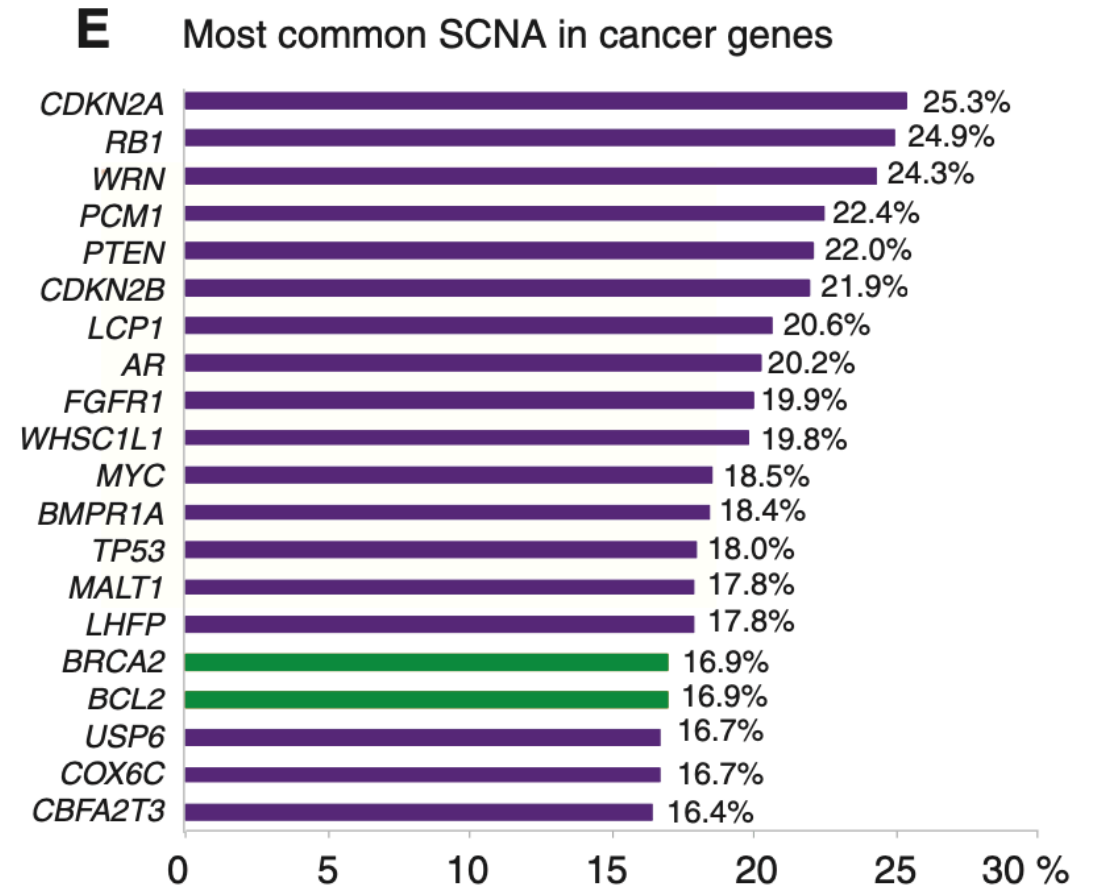
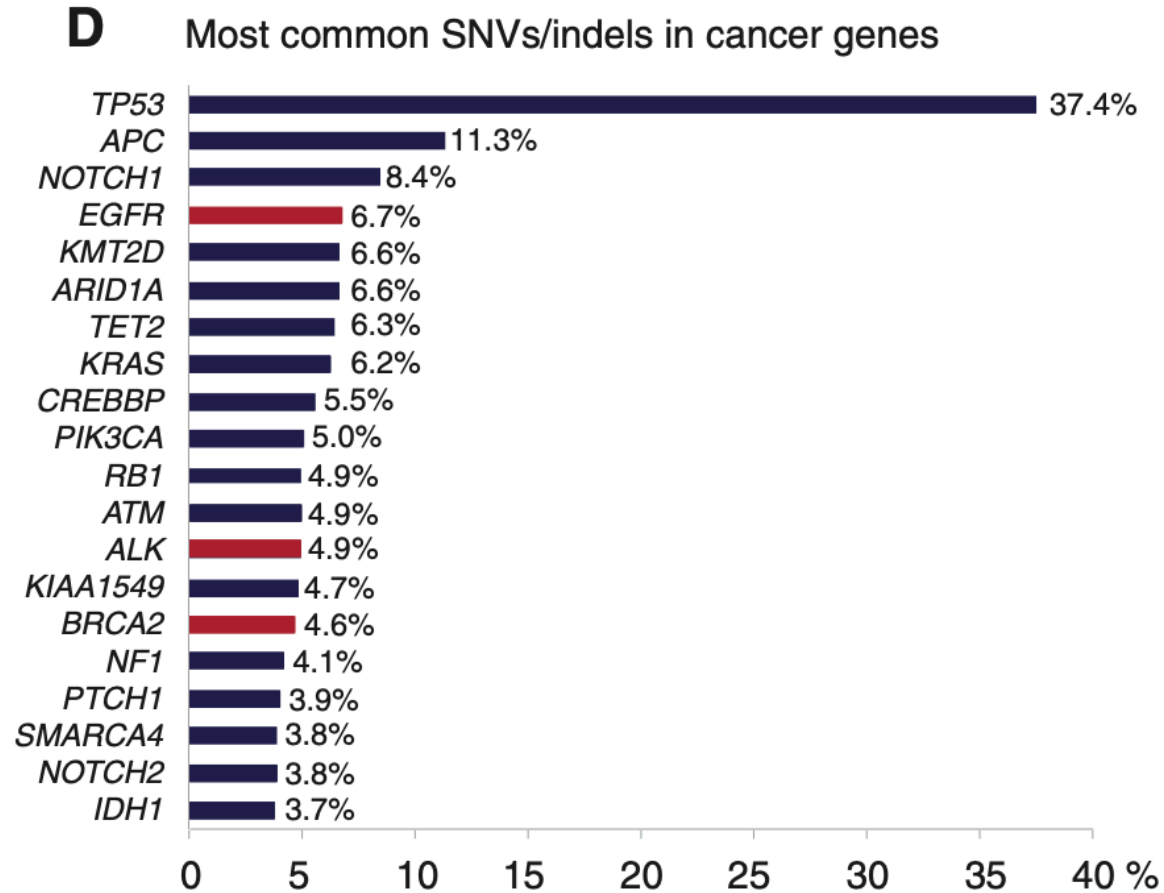
# Organoid Pipeline Entry Points

# EIPM Organoid Pipeline:



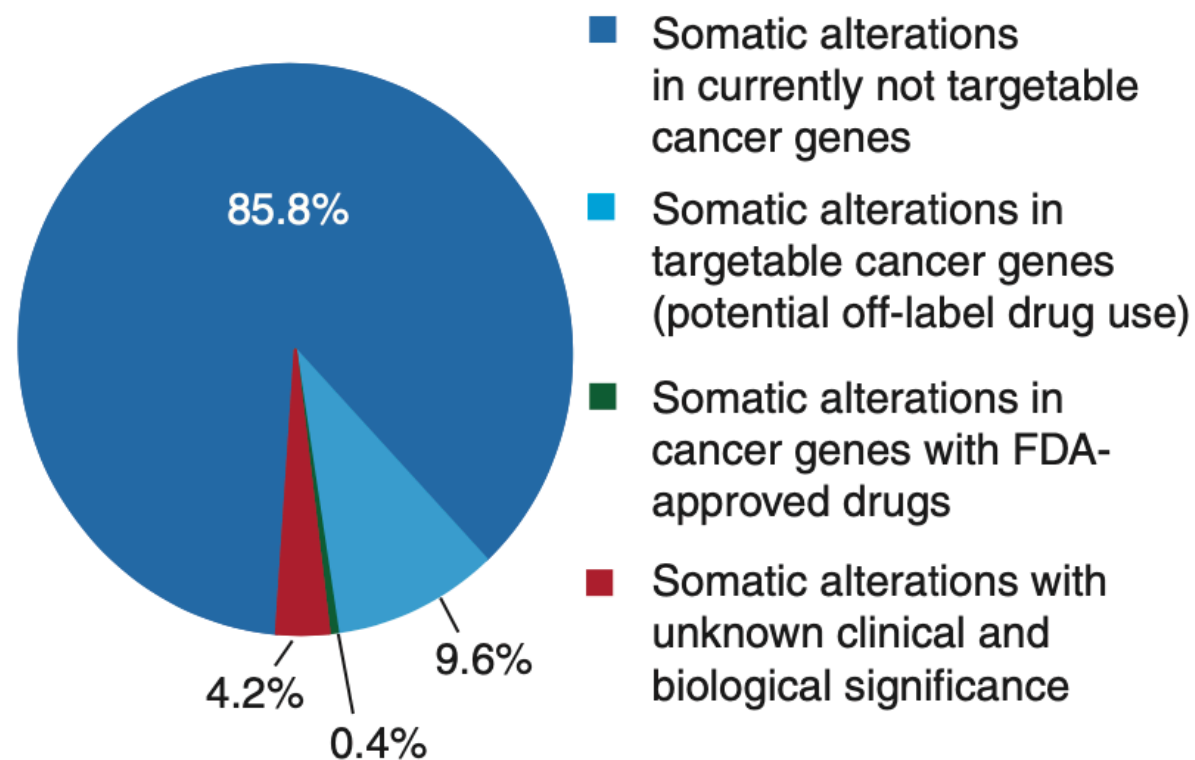
**~17\***  
people/case

# Starting Point: EXaCT1 (WES)



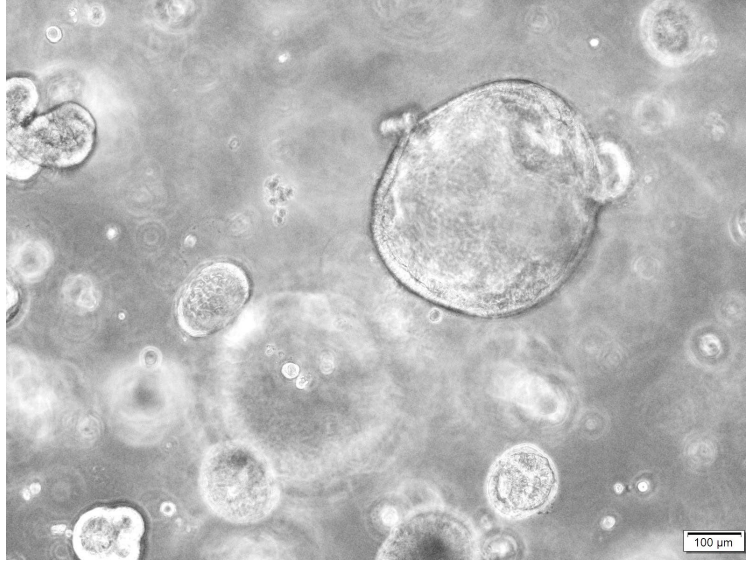
# Therapeutic Options based on WES

## **B** EXaCT-1 overview: detected genomic alterations

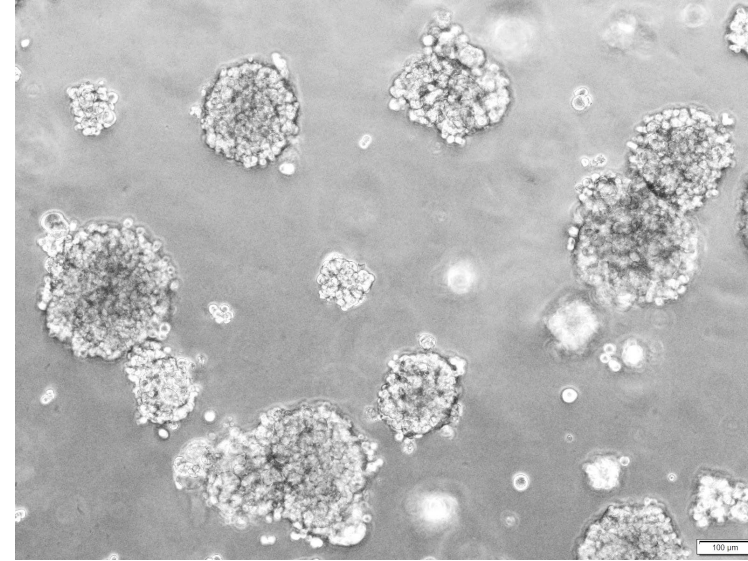


# Patient Derived Organoids

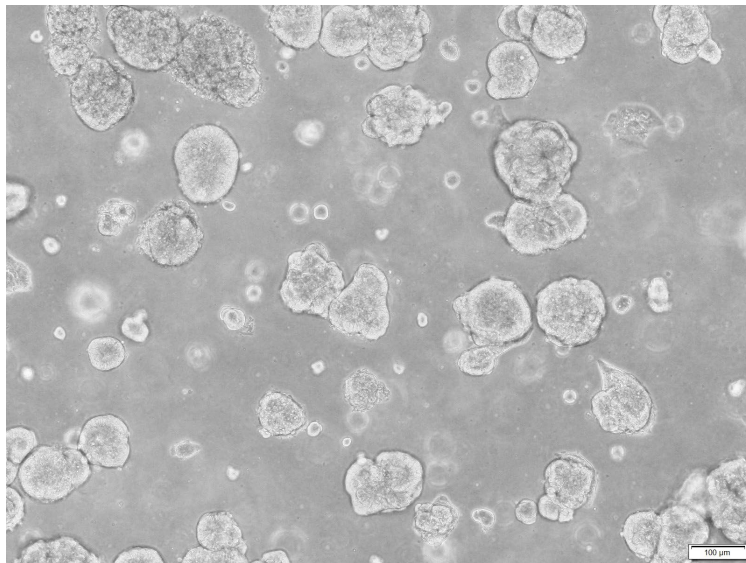
**Bladder Tumors**



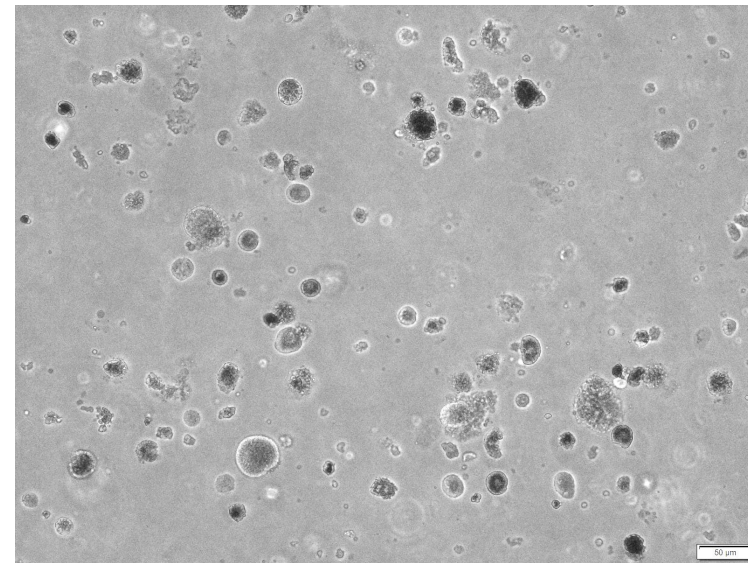
**Stomach Tumors**



**Kidney Tumors**

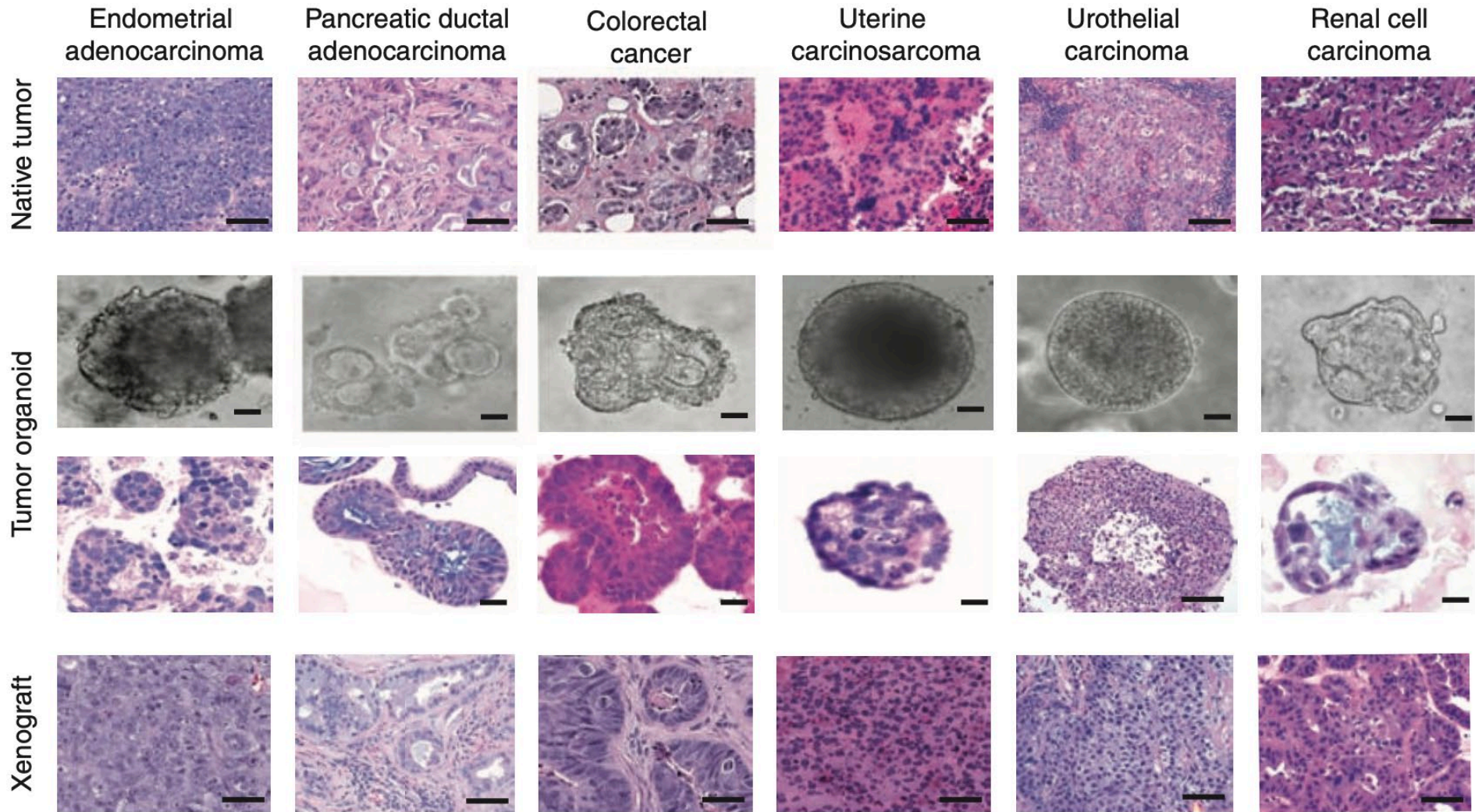


**Melanoma Tumors**

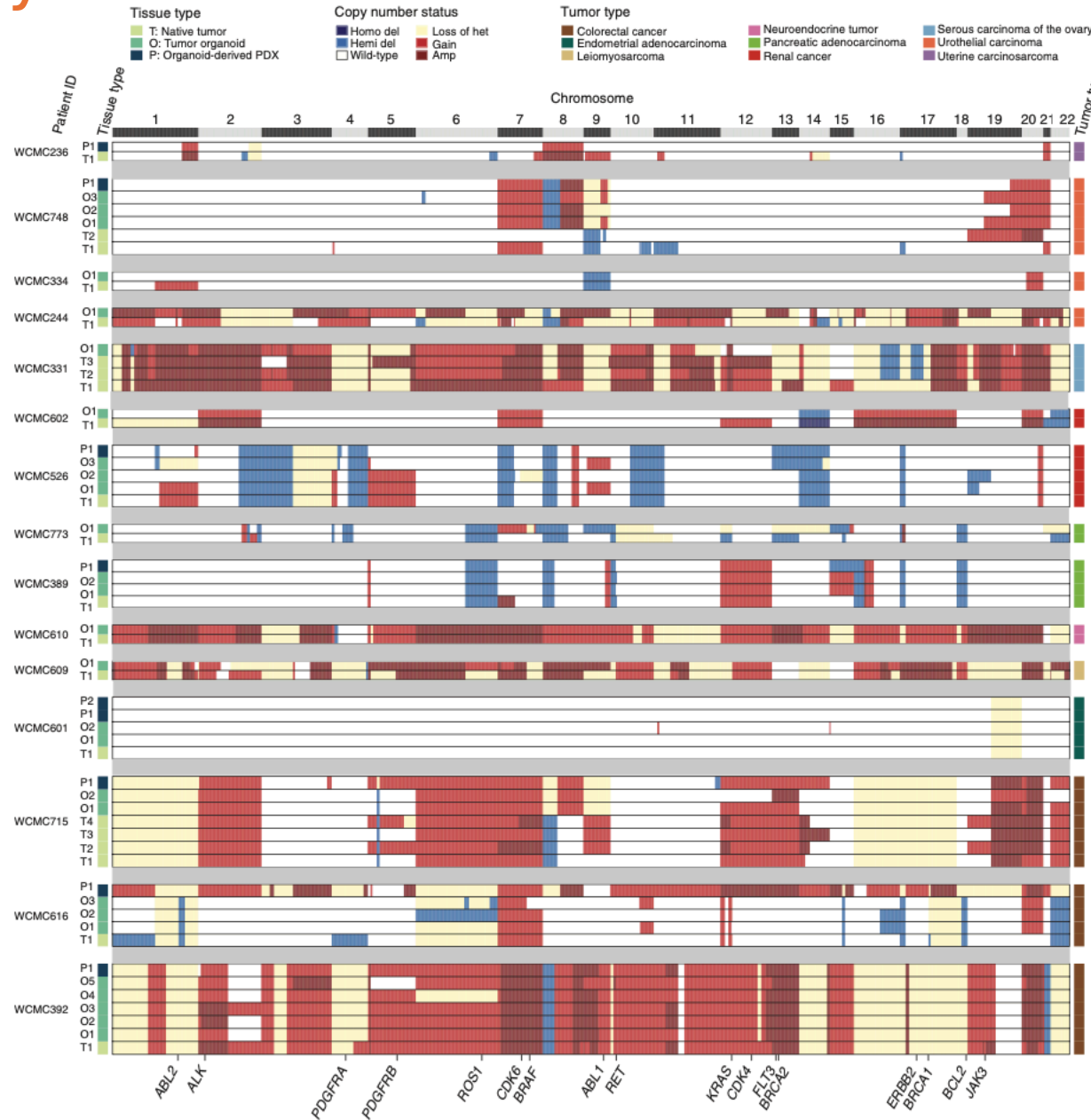


# Pathology of Organoid Models

## D Native tumor specimens and their derived tumor organoid and xenograft



# Genomic Fidelity of Models:

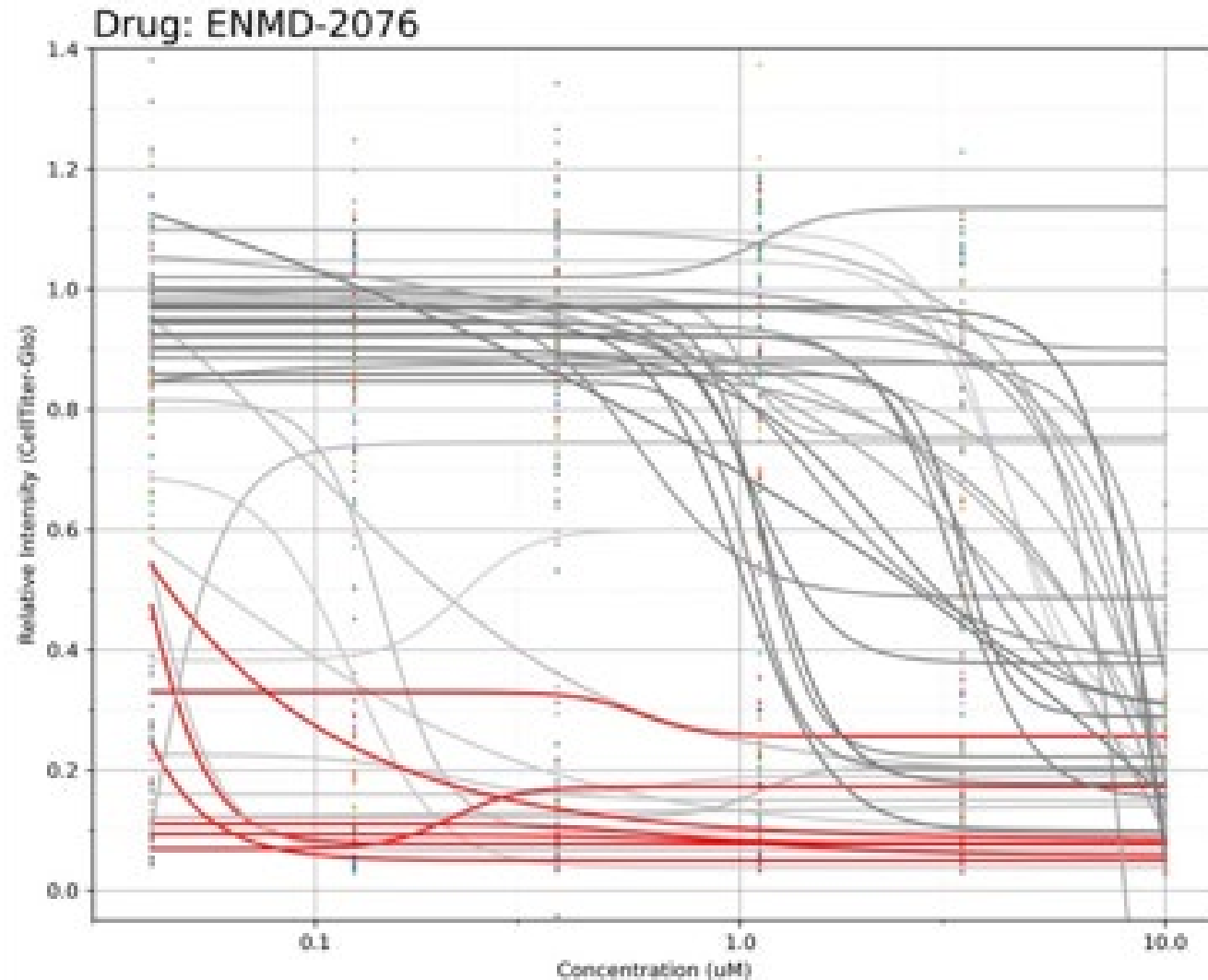


What is the best drug for the patient?

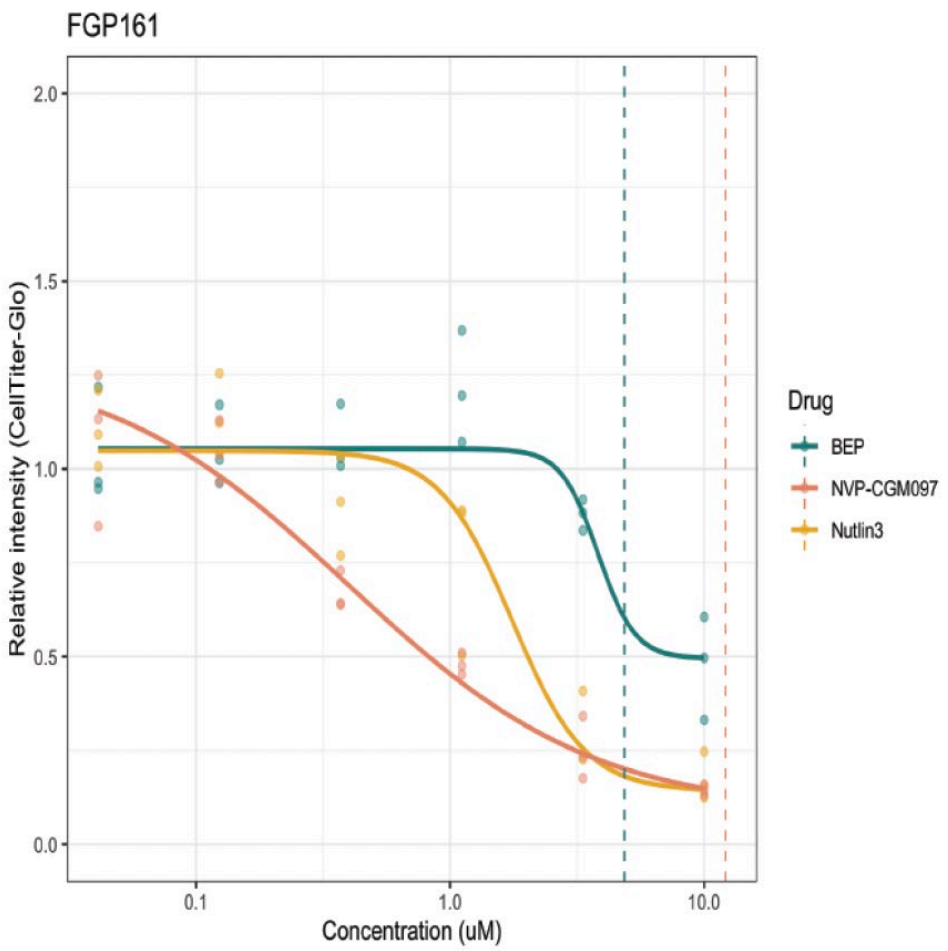
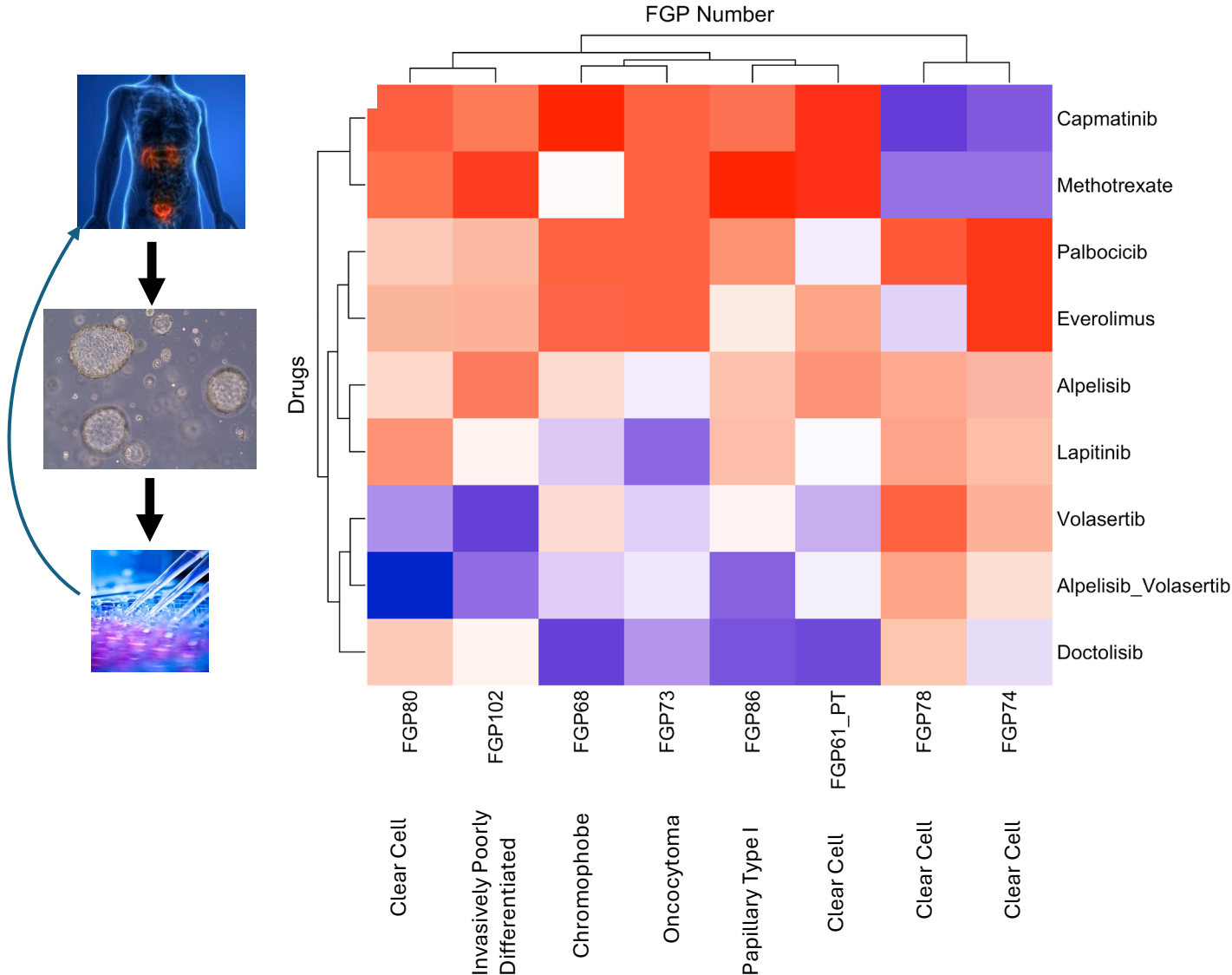
What is the best patient for each drug?



# Identification of Tumor Specific Drug Sensitivities

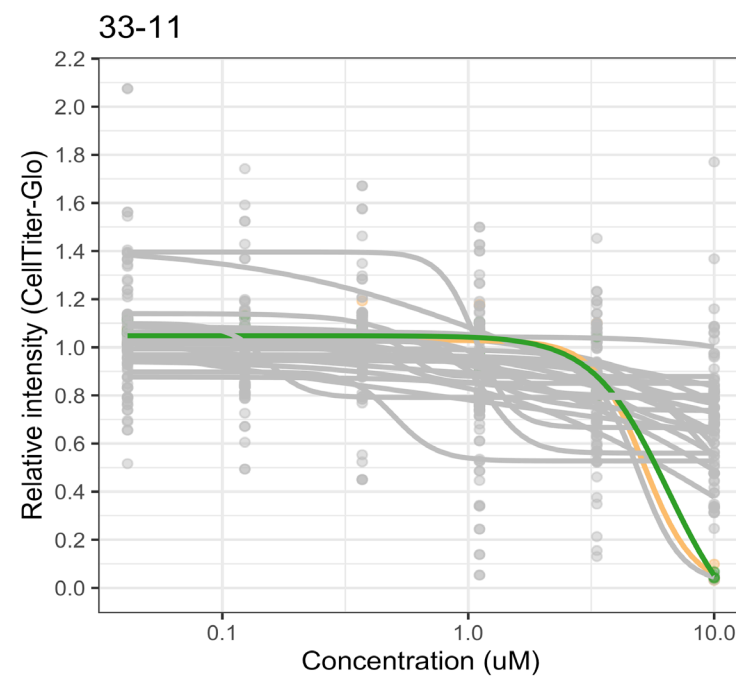
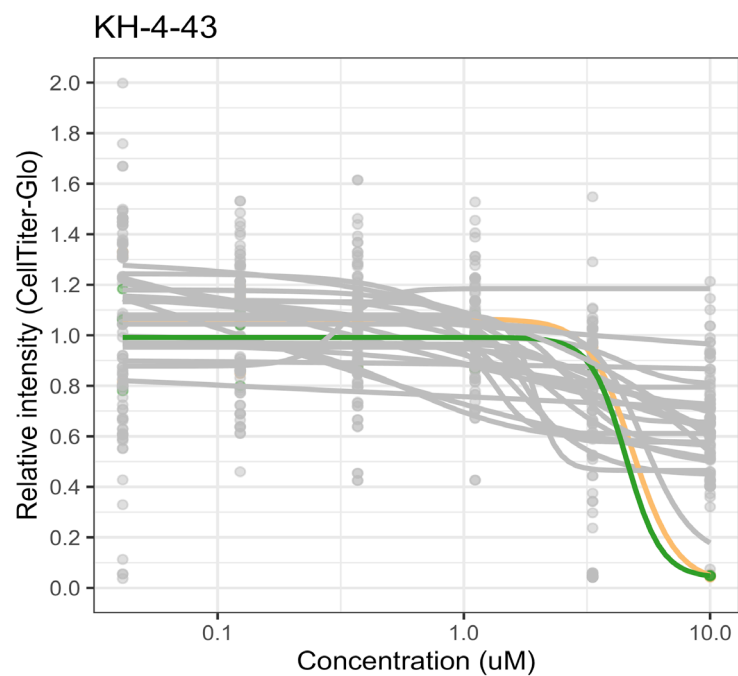
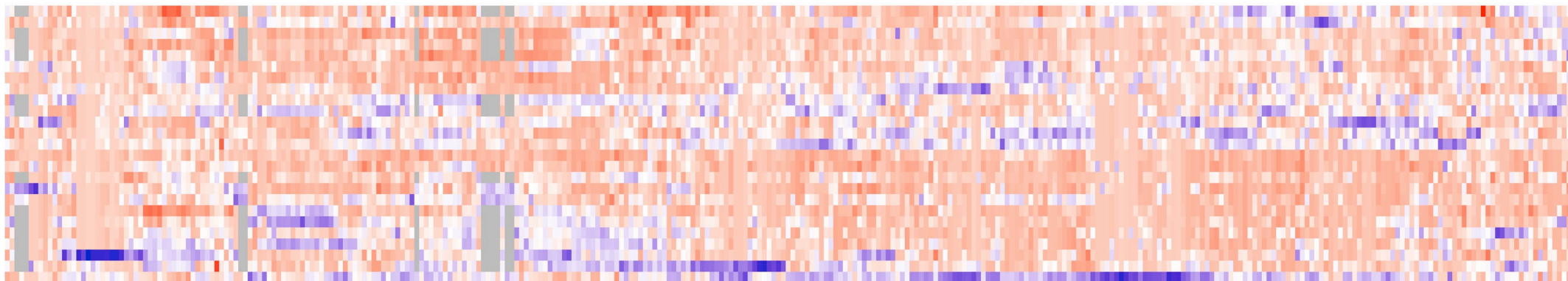


# Co-Clinical Modeling

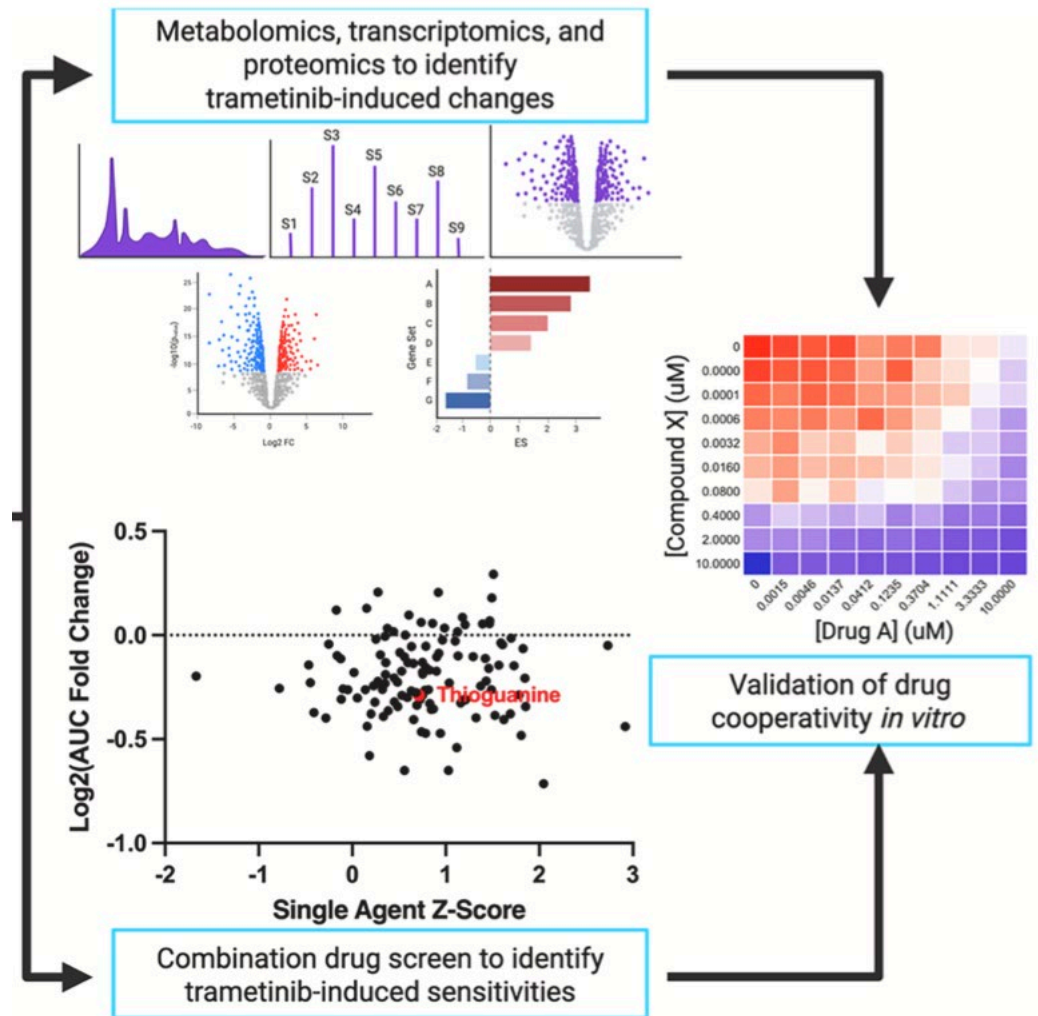


# Inverse Screening

Compounds



Organoids facilitate the integration of multi-omics with functional modeling providing platform to explore the effects of combinations in clinically representative models.





**Weill Cornell Medicine**  
**Englander Institute for Precision Medicine**

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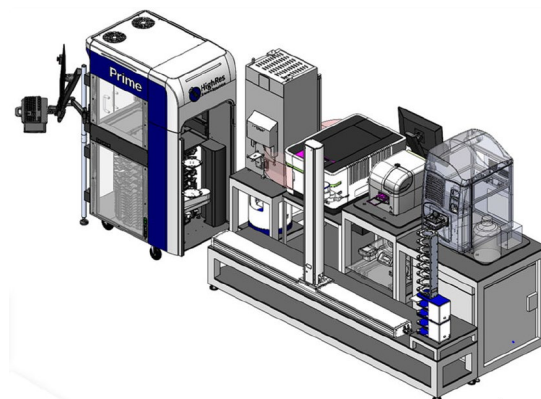
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Jesus Delgado De La Mora  
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Maame Esi Ackon  
Nile Rizvi  
Richard Farias  
Anneliese Baum  
Matthew Haeusgen

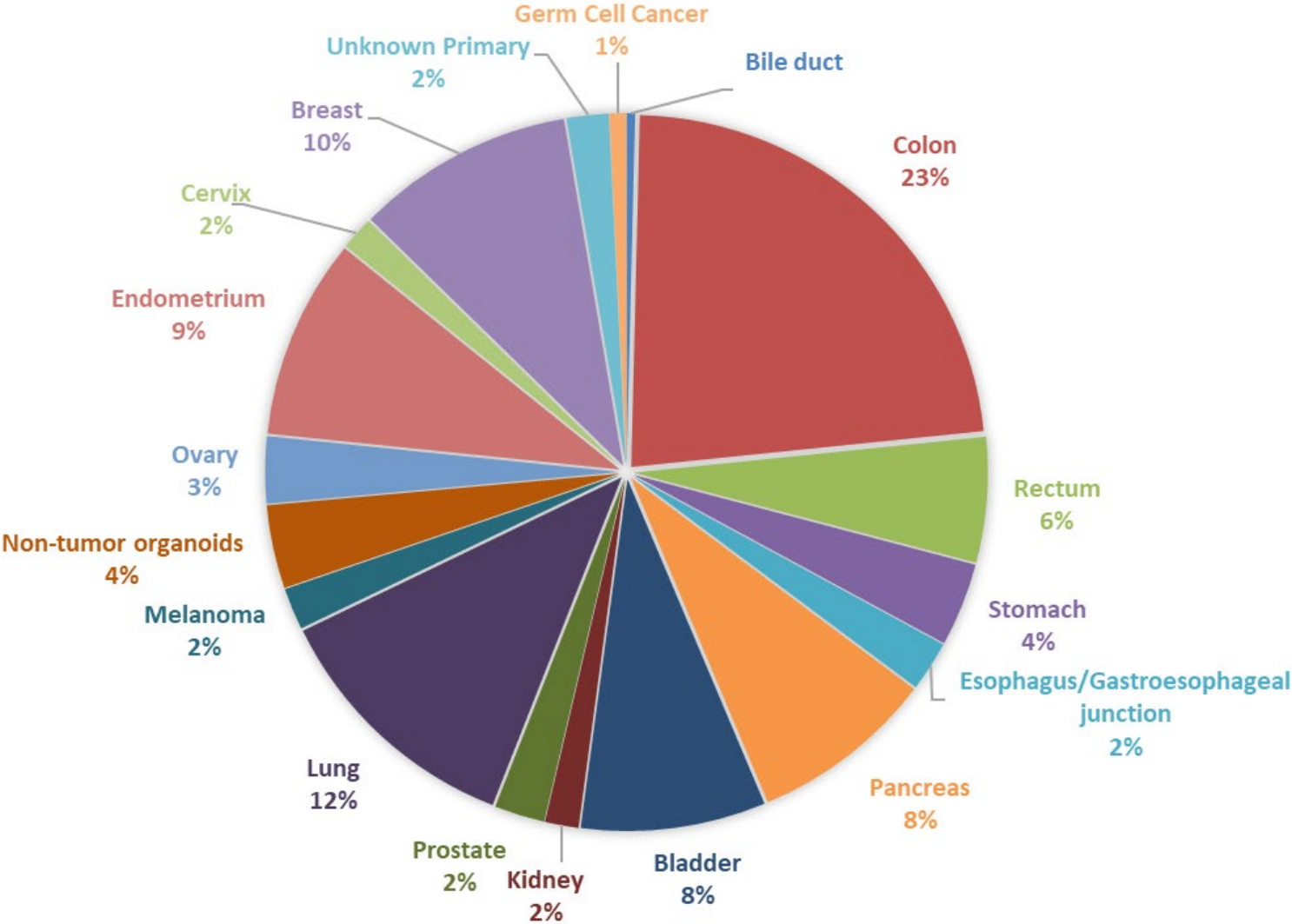


**Icahn School of Medicine**

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Willaim Zhao  
Daniela Sia  
Stephanie Blank  
Rachel Brody  
Robert Sebra  
Fred Hirsch



# Organoid biobank



# Model accessibility

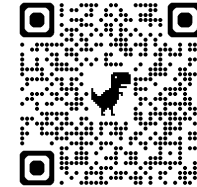
# Resources to learn more about ATCC and the HCMI



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HCMI Landing page  
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HCMI Searchable Catalog  
<https://hcmi-searchable.catalog.nci.nih.gov>

NCI Genomic Data Commons  
<https://portal.gdc.cancer.gov/projects/HCMI-CMDC>

# Posters



ATCC's patient-derived 2-D & 3-D cancer models make translational oncology a reality for the scientific community

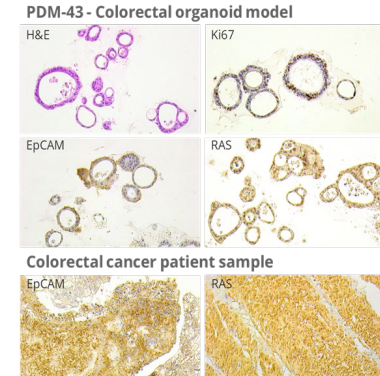
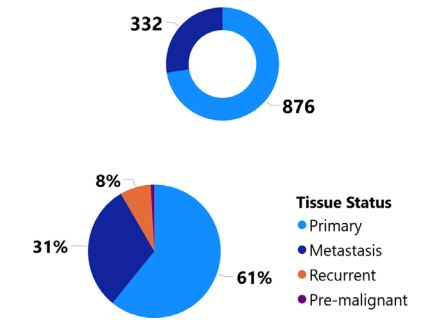
4/20/2026 2:00:00 PM

Location: Poster Section 28

Poster Board Number: 10

Presentation Number: 3405

● ATCC Production Pipeline ● Available for purchase at ATCC



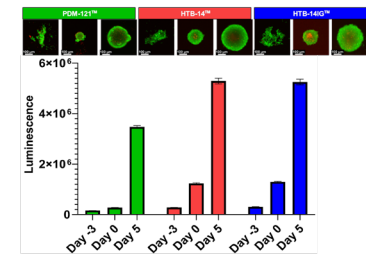
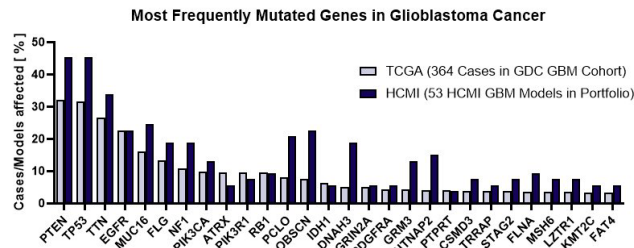
Patient-derived pediatric glioblastoma models provide key insights into IDH1-driven drug resistance

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Poster Board Number: 7

Presentation Number: 6171



Transcriptomic and therapeutic insights from patient-derived colorectal cancer organoids

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Location: Poster Section 1

Poster Board Number: 24

Presentation Number: 5457

