

# Inside Job: Targeted Assembly of Obligate Intracellular Bacterial Genomes



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## Obligate Intracellular Bacteria

Obligate intracellular bacteria (OIB), unlike other bacterial genera, are entirely dependent upon their eukaryotic hosts for survival. Members of this group include species from the *Chlamydia*, *Anaplasma*, and the *Rickettsia* genera, many of which are pathogenic to humans. Due to their absolute dependence on their hosts, genome sequencing and assembly of OIB can be complicated by the inability to separate bacterial sequences from host DNA. Here, we present ATCC®’s OIB assembly pipeline.

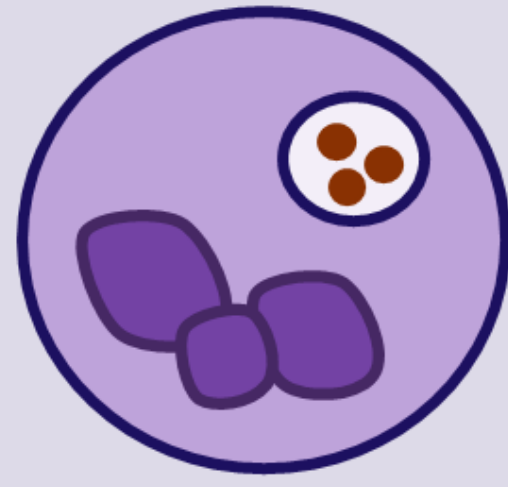
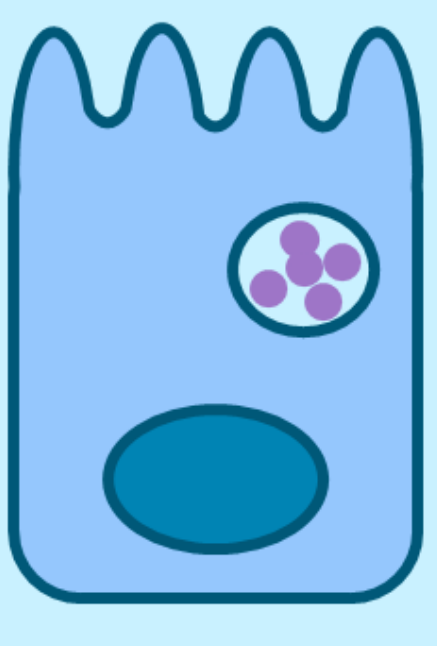
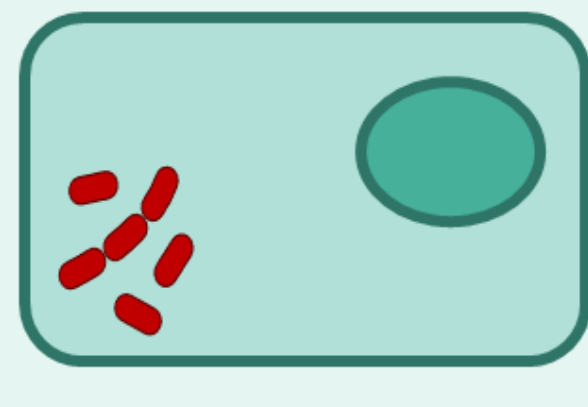
	<i>Anaplasma</i>	<i>Chlamydia</i>	<i>Rickettsia</i>
			
Genome Size	1-1.5 Mb	1-1.2 Mb	1.1-2 Mb
GC Percentage	41.5-49%	37.5-42%	29-32.5%
Transmission	vector	direct contact	vector
Transmission mode	arthropod-human	human-human	arthropod-human
Host cell tropism	neutrophils & erythrocytes	epithelial	endothelial
Replication niche	vacuole	vacuole	cytoplasm
Associated Diseases	Anaplasmosis	Chlamydia	Rocky Mountain Spotted Fever

Figure 1: Characteristics of OIB. Genomes of OIB are often quite small as compared to other bacteria, and their transmission mode and host cell tropism can vary widely across genera.

## OIB Pipeline Design

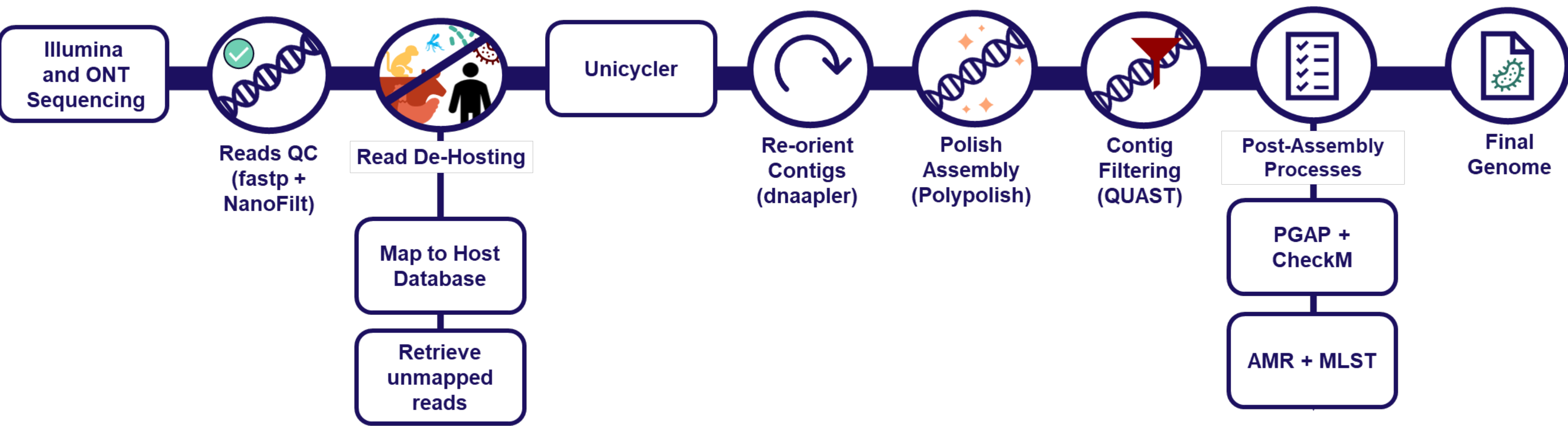


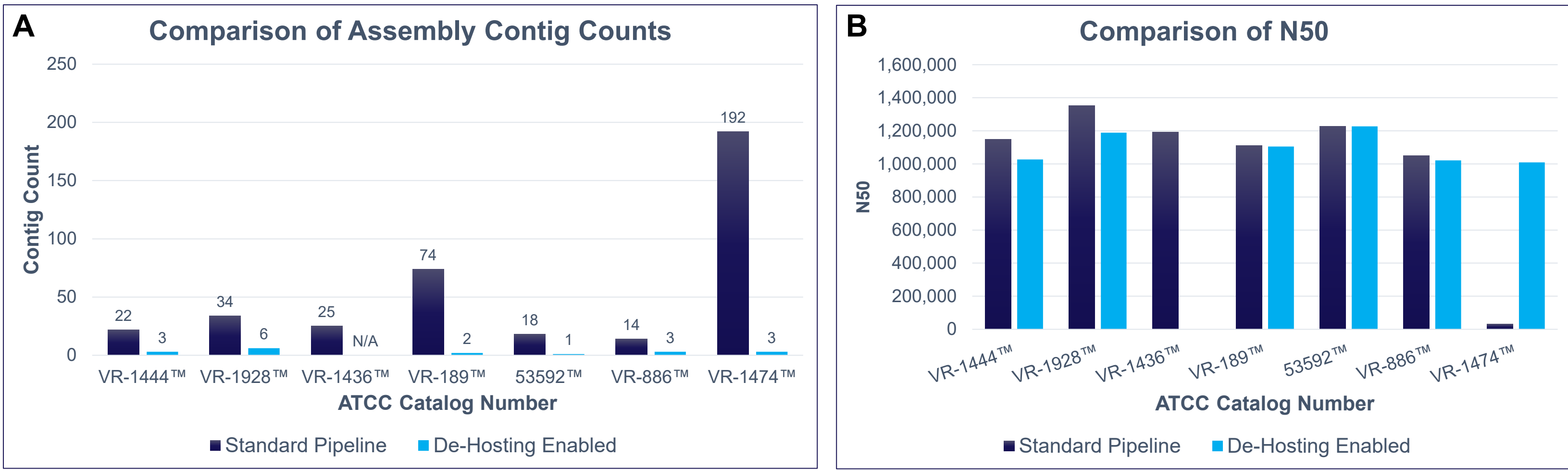
Figure 2: OIB pipeline. OIB go through ATCC®’s standard bacterial assembly pipeline with the addition of a read de-hosting step which involves mapping both short and long reads to a custom host database. Reads that do not map to the database are considered “non-host” and are used as input through the rest of the pipeline.

OIB Name	Additional Notes	Available Genomes in NCBI
<i>Rickettsia canadensis</i>	Unclear pathogenicity, infection observed in humans	2
<i>Rickettsia tamurae</i> subsp. <i>buchneri</i>	Tick symbiont	2
<i>Rickettsia monacensis</i>	Causes acute tickborne rickettsiosis in humans	1
<i>Anaplasma marginale</i>	Causes acute infection in cattle	24
<i>Chlamydia muridarum</i>	Causes chlamydia in mice	35
<i>Chlamydia trachomatis</i>	Causes chlamydia in humans	385
<i>Chlamydia pneumoniae</i>	Causes pneumonia in humans	16
<i>Chlamydia pecorum</i>	Causes acute polyarthritis in lambs	28
<i>Chlamydia abortus</i>	Causes abortion in mammals	74
<i>Chlamydia caviae</i>	Causes conjunctivitis and respiratory infections in guinea pigs	2

## Pipeline Comparisons

Table 2: Results summary. Assemblies generated with our standard bacterial pipeline have acceptable lengths when compared to the expected species length. However, when de-hosting was enabled, the generated bacterial assemblies are slightly closer in length.

Catalog Number	Name	Expected Length (NCBI)	Assembly Length	
			Standard Pipeline	De-Hosting Enabled
ATCC® VR-1444™	<i>Rickettsia canadensis</i>	1,150,228	1,344,391	1,160,365
ATCC® VR-1928™	<i>Rickettsia monacensis</i>	1,353,450	1,664,611	1,386,810
ATCC® VR-1436™	<i>Anaplasma marginale</i>	1,202,435	1,324,627	N/A
ATCC® VR-189™	<i>Chlamydia pecorum</i>	1,106,403	1,664,411	1,113,162
ATCC® 53592™	<i>Chlamydia pneumoniae</i>	1,225,935	1,371,451	1,227,376
ATCC® VR-886™	<i>Chlamydia trachomatis</i>	1,042,519	1,174,501	1,046,096
ATCC® VR-1474™	<i>Chlamydia suis</i>	1,094,002	1,508,837	1,033,562



Figures 4: Contig counts and completeness comparison. (A) The de-hosting enabled pipeline drastically reduced contig counts in OIBs compared to the standard pipeline. (B) However, the n50s for assemblies generated by each pipeline stayed about the same.

## OIB Methylation Data

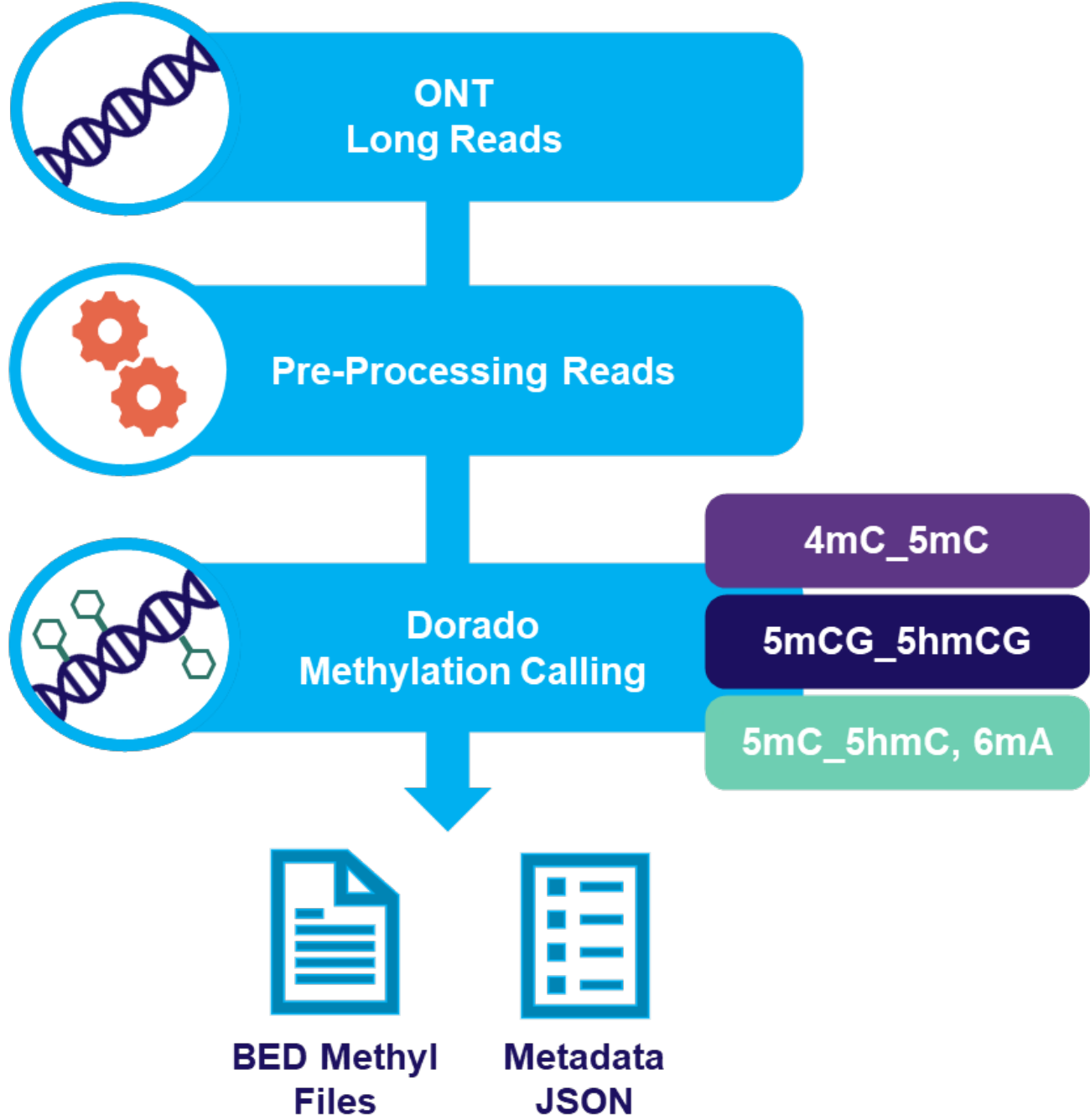


Figure 5: Methylation calling pipeline. All bacteria published to the ATCC® Genome Portal (AGP) go through long-read methylation calling. Methylation data can provide insight to the mechanisms which OIBs use to survive in host cells, which is still not quite understood. In this pipeline, several different methylation models are used.

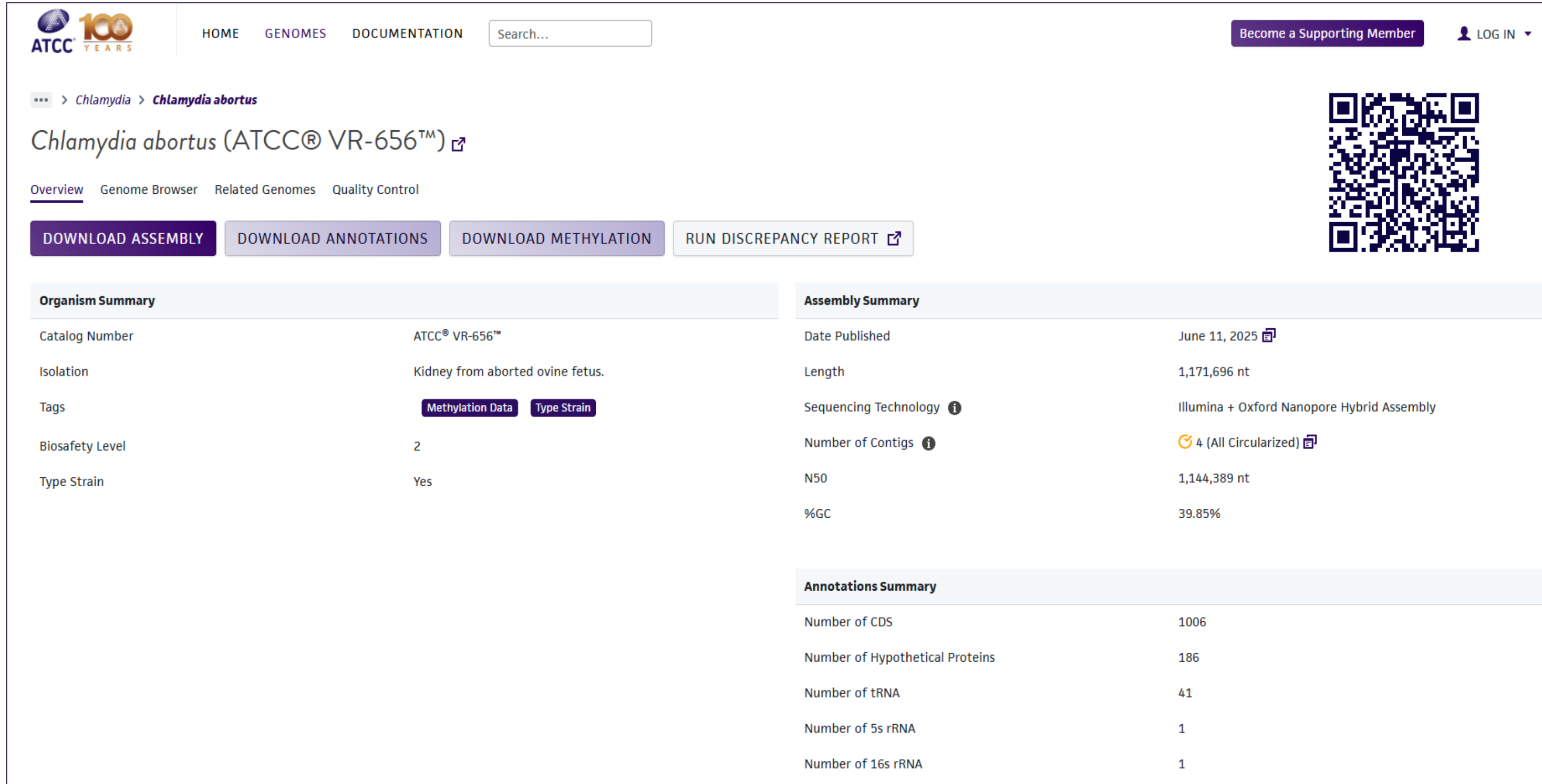


Figure 6: OIB on the AGP. An example page of VR-656™ is shown. All OIBs that met our publication criteria as a result of this analysis were or will be published to the AGP.

## Conclusion

The mechanisms that OIBs employ to survive within the host cell are not yet well understood, but through the assembly of their genomes, we can elucidate the unique host-pathogen interactions that allow OIBs to effectively replicate in their host cells. In this analysis, de-hosting reads prior to assembly led to viable, complete assemblies. Based on these results, this method will be used moving forward to publish OIBs to the AGP.