

GAS-Luc2 Reporter Cell Lines for Immune Checkpoint Drug Screening in Solid Tumors

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Abstract

Cancer immunotherapies that target immune checkpoints, such as immune checkpoint inhibitors (ICIs), antibody-dependent cellular cytotoxicity (ADCC), and antibody-drug conjugates (ADCs), have shown tremendous success in the treatment of solid tumors, including skin, lung, breast, renal, and liver cancers. However, the built-in complexity of immunological models and the variable drug responses among different cancer types have challenged the development and application of these novel immunotherapies. To facilitate large-scale drug discovery for this growing class of immunomodulators, we conducted a comprehensive cell surface protein profiling of ATCC®'s vast portfolio of human tumor and immune cell lines for established and novel immune checkpoint molecules as well as their binding ligands. Based on this protein profiling data, we generated the three immune checkpoint reporter cell lines HCC827-GAS-Luc2 (ATCC® CRL-2868-GAS-LUC2™), MG-63-GAS-Luc2 (ATCC® CRL-1427-GAS-LUC2™), and NCI-H1650-GAS-Luc2 (ATCC® CRL-5883-GAS-LUC2™), which endogenously express high levels of programmed death-ligand 1 (PD-L1), cluster of differentiation 155 (CD155), and B7 homolog 3 protein (B7-H3/CD276), respectively. These reporter cell lines were engineered to contain a gamma interferon activation site (GAS)-response element upstream of a luciferase gene. The luciferase expression is suppressed when the relevant immune checkpoint marker on the cancer cells binds to the corresponding checkpoint protein on T cells. In the presence of a relevant immune checkpoint inhibitor, the GAS-Luc2 reporter cell senses the IFN γ from the activated T cells to produce a luciferase expression-based bioluminescent signal. This signal can be readily detected and quantified to evaluate the efficacy, potency, and dynamics of the checkpoint inhibitor. In addition to drug screening for immune checkpoint inhibitors, these GAS-Luc2 reporter tumor cell lines have also been demonstrated to be effective in detecting paracrine IFN γ signaling for immune checkpoint-targeted ADCC drug development.

Background

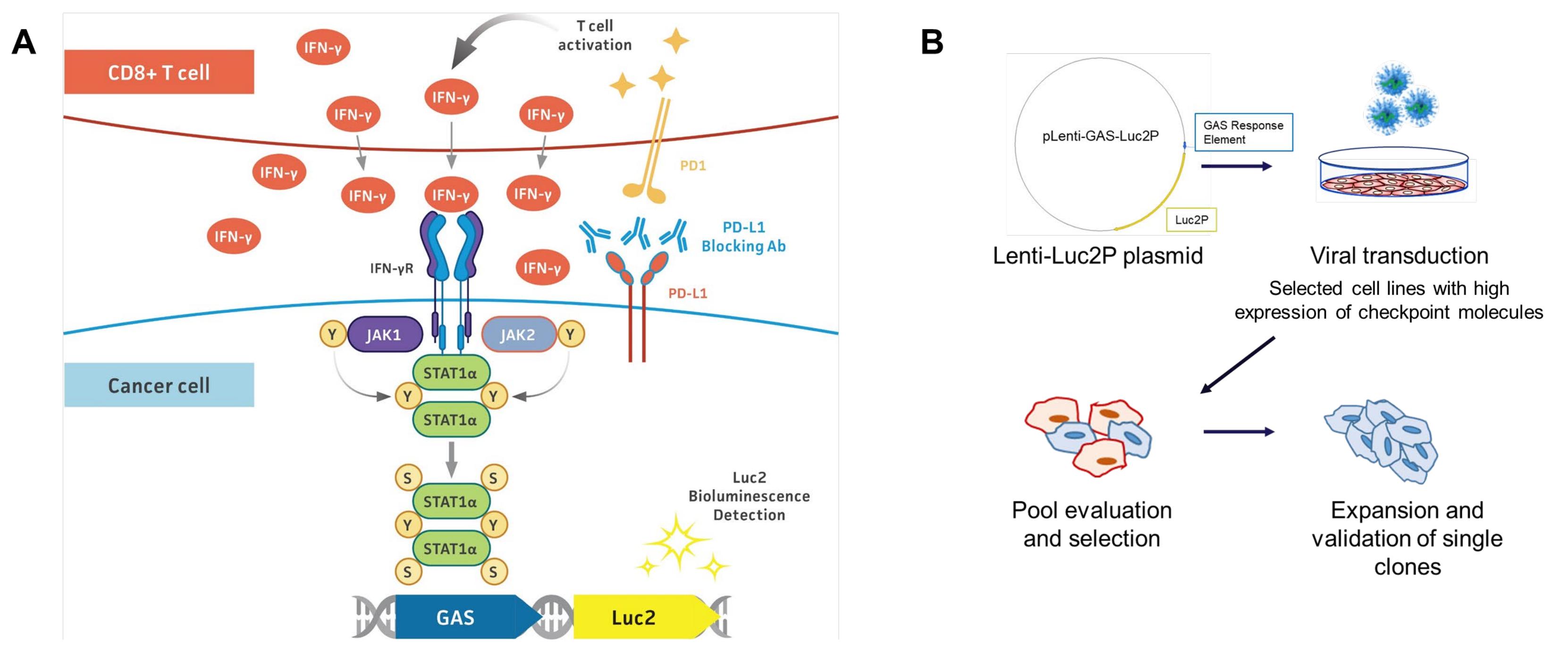


Figure 1: Schematics of the immune checkpoint molecule-expressing GAS-Luc2 reporter system. (A) Disruption of immune checkpoint binding, such as PD-1/PD-L1 recognition, by a blocking antibody activates CD8+ T cells, which then release IFN- γ . IFN- γ activates JAK-STAT signaling in cancer reporter cells, promoting GAS-induced transcription of the luciferase gene, producing an easily detectable bioluminescence signal. Created with BioRender.com. (B) Selected cell lines with high endogenous expression of PD-L1, CD155, or B7-H3 were transduced with lentiviral-GAS-Luc2 plasmids in the presence of 50 μ g/mL protamine sulfate (Sigma) for 24 hours. The cells were then enriched by puromycin selection and single cells were isolated by automatic cell sorting (Sony SH800). Expanded single cell clones were evaluated by IFN- γ stimulation. The clones that yielded the highest luciferase signal upon IFN- γ stimulation were selected for future experiments.

Results

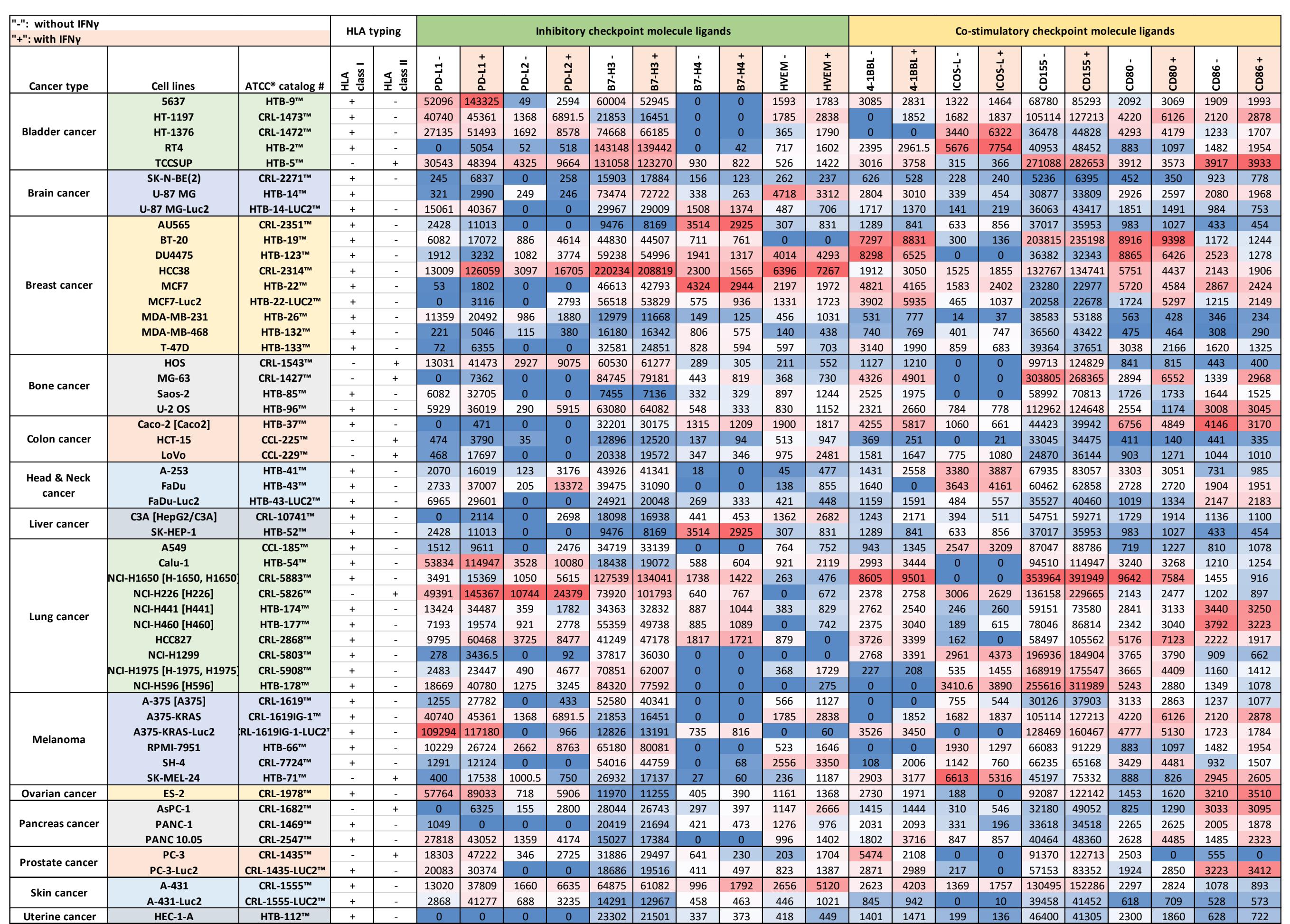


Figure 2: Heat map based on protein profiling data of selected cancer cell lines for immune checkpoint molecule expression by flow cytometry. Immune checkpoint molecule ligand expression levels in cancer cell lines under basal (-) and 100 ng/mL IFN- γ stimulated (+) conditions were profiled. HLA class was defined by either low expression (-) or high expression (+). Table values represent median fluorescence intensity sample values subtracted by isotype control MFI. Each column was color-coded separately to avoid cross comparison.

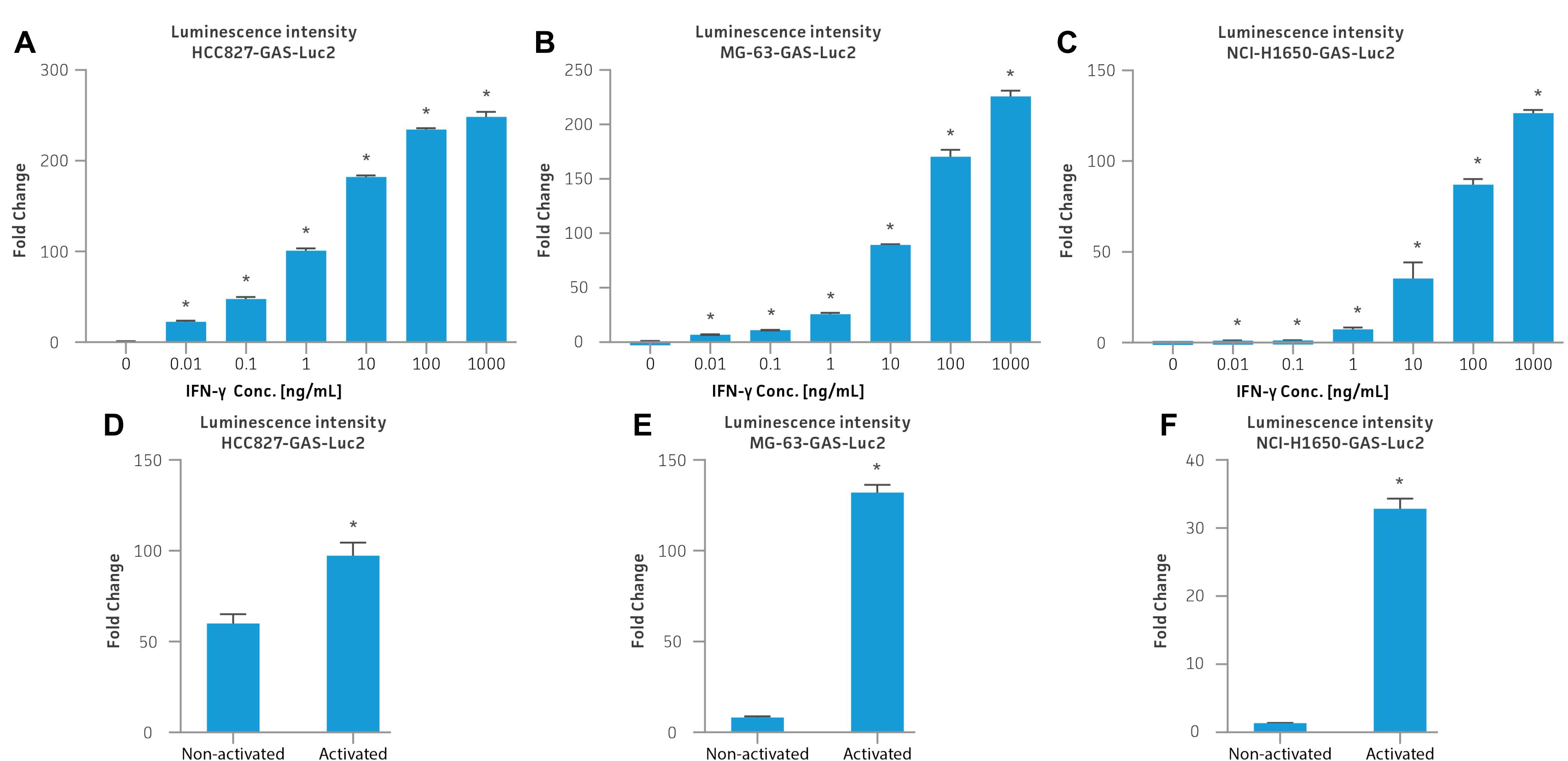


Figure 3: Cytokine stimulation of monoclonal GAS-Luc2 cell lines with IFNy or CD8+ cytotoxic T cell-conditioned media. (A-C) The luminescence intensity of the (A) HCC827-GAS-Luc2, (B) MG-63-GAS-Luc2, or (C) NCI-H1650-GAS-Luc2 cell line upon IFNy stimulation (0.01-1,000 ng/mL). (D-F) The luminescence intensity of the (D) HCC827-GAS-Luc2, (E) MG-63-GAS-Luc2, or (F) NCI-H1650-GAS-Luc2 cell line upon stimulation with conditioned media collected from non-activated or activated CD8+ cytotoxic T cells.

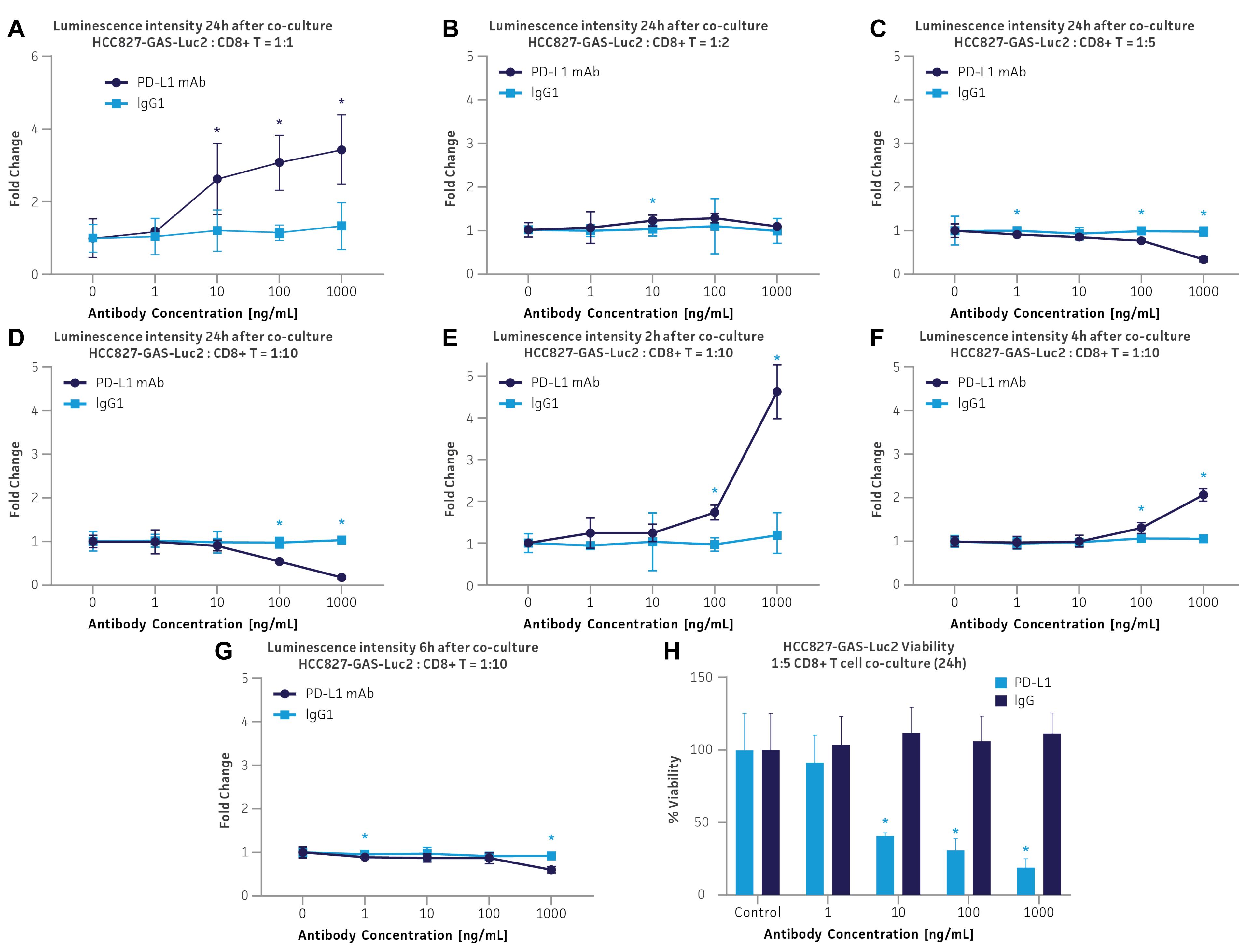


Figure 4: Co-culture of monoclonal HCC827-GAS-Luc2 with primary human CD8+ cytotoxic T cells at varying cell ratios and co-culture durations in the presence of a PD-L1 blocking antibody. (A-D) The luminescence intensity from HCC827-GAS-Luc2 cells after 24-hour co-culture with CD8+ cytotoxic T cells at a (A) 1:1, (B) 1:2, (C) 1:5, and (D) 1:10 ratio of target cells to effector cells. (E-G) The luminescence intensity from HCC827-GAS-Luc2 cells after co-culture at a 1:10 ratio with CD8+ cytotoxic T cells for periods of (E) 2 hours, (F) 4 hours, and (G) 6 hours. (H) The viability of HCC827-GAS-Luc2 after 24 hours of co-culture with a 1:5 ratio with CD8+ cytotoxic T cells for 24 hours. During the co-culture, the cells were administered with either PD-L1 mAb or isotype control IgG (1-1,000 ng/mL). N=3 in all experiments. *P < 0.05.

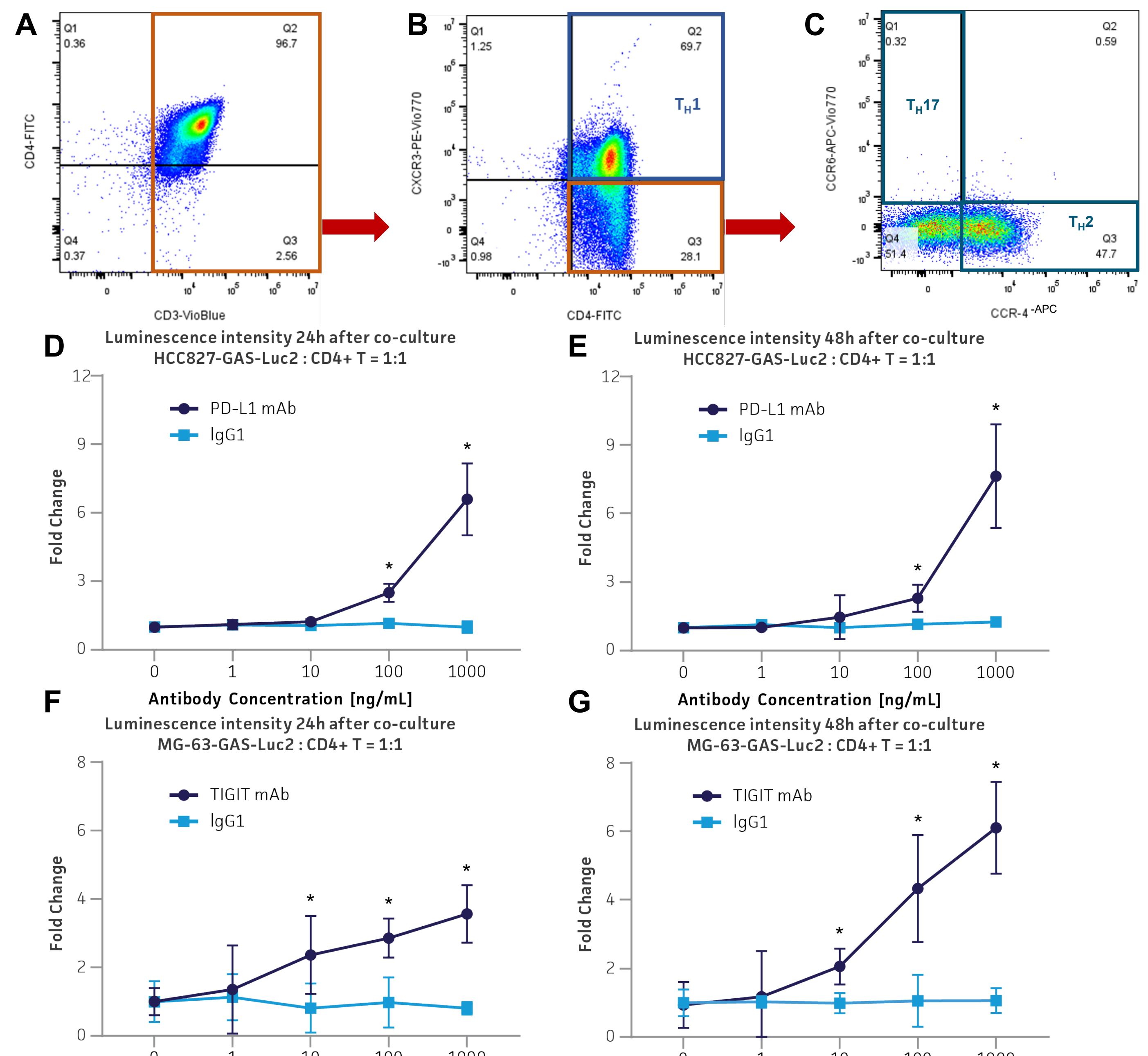


Figure 5: Co-culture of monoclonal HCC827-GAS-Luc2 or MG-63-GAS-Luc2 cells with primary human CD4+ helper T cells in the presence of a respective blocking antibody after CD4+ helper T cell subset phenotyping. (A-C) The flow cytometry analysis of helper T cell subsets T_H1 , T_H17 , and T_H2 . (D-E) The luminescence intensity from HCC827-GAS-Luc2 cells after co-culture at a 1:1 ratio with CD4+ helper T cells for (D) 24 hours or (E) 48 hours in the presence of a PD-L1 mAb or isotype control IgG1 (1-1,000 ng/mL). (F-G) The luminescence intensity from MG-63-GAS-Luc2 cells after co-culture at a 1:1 ratio with CD4+ helper T cells for (F) 24 hours or (G) 48 hours in the presence of a TIGIT mAb or isotype control IgG (1-1,000 ng/mL). N=3 in all experiments. * $P < 0.05$

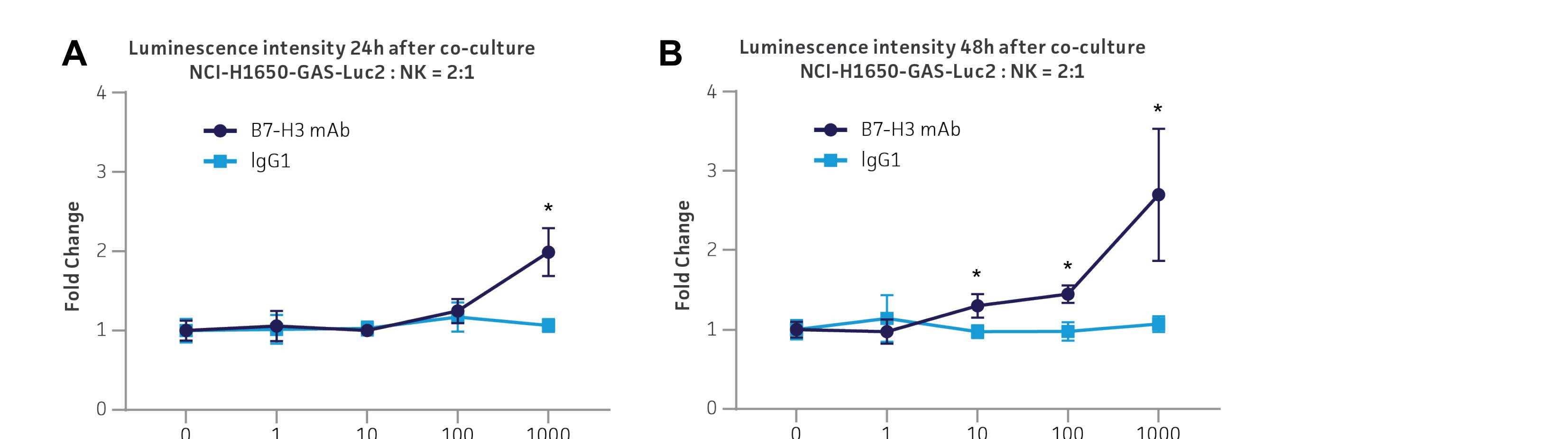


Figure 6: Co-culture of monoclonal NCI-H1650-GAS-Luc2 with primary human CD56+ NK cells in the presence of a B7-H3 ADCC antibody. (A-B) The luminescence intensity from NCI-H1650-GAS-Luc2 after co-culture with CD56+ NK cells at a 2:1 ratio of target to effector cells for (A) 24 hours or (B) 48 hours. During the co-culture, the cells

Conclusion

- The expansive protein profiling of cancer cell lines for numerous immune checkpoint molecules and their ligands provides crucial information that facilitates immune checkpoint molecule interaction studies, checkpoint assay development, and cancer immunotherapy screening.
 - Based on the protein profiling data, we developed three cancer reporter cell lines with a high endogenous expression of PD-L1, CD155, or B7-H3. The naturally high expression of the immune checkpoint molecules on the surface of the reporter cells facilitates the immune checkpoint inhibitor drug candidate binding, promoting the immune cell activation and subsequent release of IFN- γ by the immune cells, resulting in the intracellular IFN- γ -IFN- γ R JAK-STAT GAS signaling activation and luciferase expression by the reporter cells.
 - The reporter cell lines present robust, responsive, and reproducible luciferase expression upon signaling activation that enables reliable measurement of the potency and stability of the relevant immune checkpoint inhibitors that trigger immune/tumor microenvironmental cell-mediated immune responses.
 - By maintaining physiological relevance and stable expression of the checkpoint ligand owing to the endogenous expression, these reporter