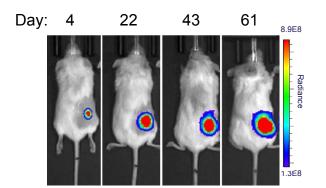


Technical Data Sheet: NRAS Mutant-A375 Isogenic-Luc2

ATCC® Number	CRL-1619IG-2-LUC2™
Organism	Homo sapiens
Tissue/Disease Source	Malignant melanoma
Product Description	This luciferase expressing cell line was derived from NRAS Mutant-A375 isogenic line by transduction with lentiviral vector encoding firefly luciferase gene (luc2) and subsequently through single cell cloning • Signal noise ratio: ≥ 1,000 • Bioluminescence: ≥ 100,000 photons/cell/sec (subject to imaging and culture condition) • Confirmed to be murine pathogen free
Application	BRAF drug resistant melanoma model. Excellent signal/background ratio and stable Luciferase expression make this cell line ideal for in vivo bioluminescence imaging of xenograft animal model to study human cancer and monitor activity of anti-cancer drug. It also can be used in cell-based assays for cancer research.

In vivo Bioluminescent Imaging



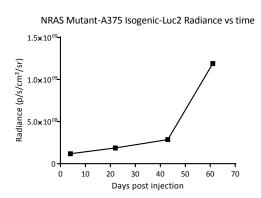


Figure 1: *In vivo* **detection of luciferase activity of NRAS mutant-A375 Isogenic-Luc2.** NRAS Mutant-A375 Isogenic-Luc2 cells (3 x 10⁶) were injected subcutaneously into the dorsal region near the thigh of female NSG mice. Tumor growth was monitored weekly using a Xenogen IVIS Spectrum. *In vivo* bioluminescence imaging demonstrated the progression of tumors.

Cell Morphology

NRAS Mutant-A375 Isogenic
(CRL-1619IG-2-™)

Isogenic-Luc2
(CRL-1619IG-2-LUC2™)

Doubling time = 18.8

NRAS Mutant-A375
Isogenic-Luc2
(CRL-1619IG-2-LUC2™)

Doubling time = 17.0

Figure 2: Cell morphology of NRAS Mutant-A375 Isogenic parental and NRAS Mutant-A375 Isogenic-Luc2. Cells were maintained in ATCC recommended culture conditions. Cell morphology was observed under microscopy and images were captured by digital camera.

hours

Luciferase Expression

hours

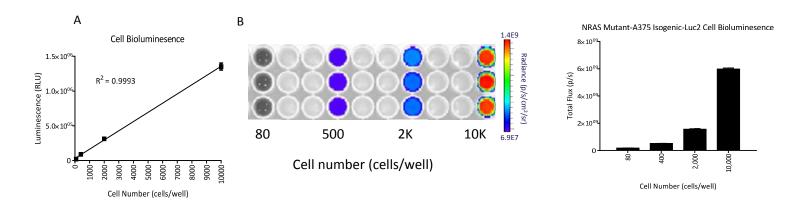


Figure 3: Linearity of luminescence and of in vitro quantification of luciferase activity of NRAS Mutant-A375 Isogenic-Luc2. Cells were seeded in a 96-well plate at indicated cell numbers per well, and Bright-Glo (Promega) was added to the indicated wells. The luminescence of the plate was read within 10 minutes using a luminescence plate reader (A) and determined to have a linear correlation of bioluminescence intensity with cell numbers. (B) The plate was imaged using a Xenogen IVIS Spectrum to quantify photons emitted per cell.

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