

## **ATCC Medium: 5608 PY Medium (M2-VPI) With D-Glucose w/ Durham Tube**

D-Glucose.....	10.0g
Peptone.....	5.0 g
Trypticase.....	5.0 g
Yeast Extract.....	10.0 g
0.025% Resazurin.....	4.0 ml
Salt Solution (see below).....	40.0 ml
Hemin Solution (see below).....	10.0 ml
Vitamin K <sub>1</sub> Solution (see below).....	0.2 ml
Cysteine HCl x H <sub>2</sub> O.....	0.5 g
DI Water.....	950.0 ml

Adjust medium for final pH 7.0 +/- 0.2. Prepare medium anaerobically using a gas phase of 80% N<sub>2</sub>, 10% CO<sub>2</sub> and 10% H<sub>2</sub>.

Add durham tubes and purge air by autoclaving at 110°C for 1-3 minutes.

\*\*\*\*\* For stabs, do not use durham tubes.

### **Hemin Solution**

Hemin.....	50.0 mg
1N NaOH.....	1.0 ml
DI Water.....	100.0 ml

Dissolve hemin in NaOH. Add DI Water. Sterilize by autoclaving at 121°C.

### **Vitamin K<sub>1</sub> Solution**

Vitamin K <sub>1</sub> (2-Methyl-3-phytl-1,4-naphthoquinine)....	0.15 ml
95% Ethanol.....	30.0 ml

Refrigerate Vitamin K<sub>1</sub> in a brown bottle. Discard after one month.

### **Salt Solution**

CaCl <sub>2</sub> .....	0.2 g
MgSO <sub>4</sub> .....	0.2 g
K <sub>2</sub> HPO <sub>4</sub> .....	1.0 g
KH <sub>2</sub> PO <sub>4</sub> .....	1.0 g
NaHCO <sub>3</sub> .....	10.0 g
NaCl.....	2.0 g
DI Water.....	1000 ml

Mix calcium chloride, magnesium sulfate and sodium chloride in 300 ml of distilled water. Mix di- and monopotassium phosphates in 200 ml of distilled water. Dissolve sodium bicarbonate in 500 ml of distilled water. Combine the 3 solutions. Store at 4°C.