ATCC

Product Sheet

HCC1008 (ATCC® CRL-2320™)

Please read this FIRST

Storage Temp.
liquid nitrogen vapor phase

Biosafety Level
1

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

HITES medium supplemented with 10% fetal bovine serum.The base medium for this cell line is ATCC-formulated DMEM:F12 Medium Catalog No.30-2006. To make the complete growth medium,add the following components to the base medium:

- 0.005 mg/ml Insulin
-
- 0.01 mg/ml Transferrin
-
- 30nM Sodium selenite (final conc.)
-
- 10 nM Hydrocortisone (final conc.)
-
- 10 nM beta-estradiol (final conc.)
-
- extra 2mM L-glutamine (for final conc. of 4.5 mM)
-
- 10% fetal bovine serum (final conc.)

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: HCC1008 (ATCC® CRL-2320™)

American Type Culture Collection
PO Box 1549
Manassas, VA 20108 USA
www.atcc.org

800.638.6597 or 703.365.2700
Fax: 703.365.2750
Email: Tech@atcc.org

Or contact your local distributor

Description

Organism: *Homo sapiens*, human
Disease: TNM stage IIA, grade 3,ductal carcinoma
Cell Type: Epithelial
Age: 67 years adult
Gender: female
Morphology: epithelial
Growth Properties: adherent
DNA Profile:
Amelogenin: X
CSF1PO: 12,13
D13S317: 12
D16S539: 14,15
D5S818: 13
D7S820: 10
THO1: 7
TPOX: 11
vWA: 17,19
Cytogenetic Analysis: multiploid; cell population has several ploidy indices; double minute (DM) chromosomes were observed

SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

Handling Procedure for Frozen Cells
To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at
-70°C. Storage at -70°C will result in loss of viability.
SAFETY PRECAUTION: ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.
1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube with 9.0 ml culture medium. and spin at approximately 125 xg for 5 to7 minutes.
4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). and dispense into a 25 cm^2 culture flask. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
5. Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet. **This line recovers very slowly and will take approximately 2 weeks before the first subculture.**

Handling Procedure for Flask Cultures

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Handling Procedure for Flask Cultures

The flask was seeded with cells (see specific batch information) grown and completely filled with medium at ATCC to prevent loss of cells during shipping.

1. Upon receipt visually examine the culture for macroscopic evidence of any microbial contamination. Using an inverted microscope (preferably equipped with phase-contrast optics), carefully check for any evidence of microbial contamination. Also check to determine if the majority of cells are still attached to the bottom of the flask; during shipping the cultures are sometimes handled roughly and many of the cells often detach and become suspended in the culture medium (but are still viable).

2. If the cells are still attached, aseptically remove all but 5 to 10 ml of the shipping medium. The shipping medium can be saved for reuse. Incubate the cells at 37°C in a 5% CO_2 in air atmosphere until they are ready to be subcultured.

3. If the cells are not attached, aseptically remove the entire contents of the flask and centrifuge at 125 xg for 5 to 10 minutes. Remove shipping medium and save. Resuspend the pelleted cells in 10 ml of this medium and add to 25 cm^2 flask. Incubate at 37°C in a 5% CO_2 in air atmosphere until cells are ready to be subcultured.

Subculturing Procedure

Protocol:

1. **Remove and discard culture medium.**
2. **Add 3.0 to 4.0 ml Cell Dissociation Buffer (GIBCO #13150-016) to cell layer and incubate at room temperature or 37C.**
3. **Observe cells under an inverted microscope until cell layer is dispersed (usually with 5 to 10 minutes).**
   Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
4. **Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting.**
5. **To remove Cell Dissociation Buffer, transfer cell suspension to centrifuge tube and spin at approximately 125 xg for 5 to10 minutes.**
6. **Discard supernatant and resuspend cell pellet in fresh complete growth medium. Aspirate cells with a small bore pipette. Add appropriate aliquots of cell suspension to new culture vessels.**
7. **Place culture vessels in incubators at 37°C.**

Cells reattach slowly after subculture (about 50% attached and 50% in suspension). Allow flasks to remain undisturbed for at least a week for cultures to become reestablished. It can take as along as three to four weeks before cultures can be subcultured again.
Subcultivation Ratio: A subcultivation ratio of 1:2 is recommended
Medium Renewal: Every 3 to 4 days

Cryopreservation Medium

Cryoprotectant Medium
Growth medium supplemented with 7.5% (v/v) DMSO.
Cell culture tested DMSO is available as ATCC Catalog No. 4-X.

Comments

This cell line was initiated from an axillary lymph node on 6/7/94 and took 12.5 months to establish.
The tumor was classified as TNM stage IIA, grade 3, ductal carcinoma with 12 out of 12 lymph node metastasis.
The cells are positive for expressions of Her2-neu and p53 oncogenes.
HCC1008 is positive for the epithelial cell specific marker, Epithelial Glycoprotein 2 (EGP2) and cytokeratin 19.
The cells are negative for expression of estrogen receptor (ER) and for expression of progesterone receptor (PR).

References

References and other information relating to this product are available online at www.atcc.org.

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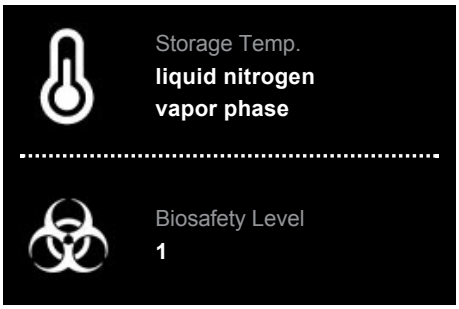

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Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

ATCC Warranty

The viability of ATCC® products is warranted for 30 days from the date of shipment, and is valid only if the product is stored and cultured according to the information included on this product information sheet. ATCC lists the media formulation that has been found to be effective for this strain. While other, unspecified media may also produce satisfactory results, a change in media or the absence of an additive from the ATCC recommended media may affect recovery, growth and/or function of this strain. If an alternative medium formulation is used, the ATCC warranty for viability is no longer valid.

Disclaimers

This product is intended for laboratory research purposes only. It is not intended for use in humans.

While ATCC uses reasonable efforts to include accurate and up-to-date information on this product sheet, ATCC makes no warranties or representations as to its accuracy. Citations from scientific literature and patents are provided for informational purposes only. ATCC does not warrant that such information has been confirmed to be accurate.

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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at www.atcc.org

Additional information on this culture is available on the ATCC web site at www.atcc.org.

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