



Product Sheet

KG-1a (ATCC® CCL-246.1™)

Please read this FIRST



Storage Temp.
liquid nitrogen
vapor phase



Biosafety Level
1

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

The base medium for this cell line is ATCC-formulated Iscove's Modified Dulbecco's Medium, Catalog No. 30-2005. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 20%.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: KG-1a (ATCC® CCL-246.1™)

American Type Culture Collection
PO Box 1549
Manassas, VA 20108 USA
www.atcc.org

800.638.6597 or 703.365.2700
Fax: 703.365.2750
Email: Tech@atcc.org

Or contact your local distributor

Description

Organism: *Homo sapiens*, human
Tissue: bone marrow
Disease: acute myelogenous leukemia
Cell Type: promyeloblast, Macrophage
Age: 59 years
Gender: male
Morphology: myeloblast
Growth Properties: suspension
Isoenzymes:
AK-1, 0
ES-D, 1
G6PD, B
GLO-I, 2
Me-2, 1
PGM1, 1-2
PGM3, 0
DNA Profile:
Amelogenin: X,Y
CSF1PO: 7
D13S317: 11,12
D16S539: 10,11
D5S818: 13
D7S820: 8,10
THO1: 7,8
TPOX: 7,9
vWA: 14,19

Cytogenetic Analysis: The stemline chromosome number is 46 (pseudodiploid), with the 2S component occurring at 5.8%. Seven markers, including five ATCC CCL-246-specific markers, were found in most, if not all metaphases analyzed. Another marker ? del (7) was found only in about 50% of the metaphases. Normal chromosomes 5, 7, 8, 12, 17 and 22 were monosomic. The Y chromosome is detected in the Q-banded preparations.

Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 ml complete culture medium. and spin at approximately 125 x g for 5 to 7 minutes.
4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for



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the culture recommended dilution ratio). It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6). pH (7.0 to 7.6).

5. Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.



Handling Procedure for Flask Cultures

The flask was seeded with cells (see specific batch information), grown, and completely filled with medium at ATCC to prevent loss of cells during shipping.

1. Upon receipt visually examine the culture for macroscopic evidence of any microbial contamination. Using an inverted microscope (preferably equipped with phase-contrast optics), carefully check for any evidence of microbial contamination
2. Incubate the flask in an upright position for several hours at 37°C. After the temperature has equilibrated, aseptically remove the entire contents of the flask and centrifuge at 125 x g for 5 to 10 minutes. Remove shipping medium and save for reuse. Resuspend the cell pellet in 10 mL of this medium.
3. From this cell suspension remove a sample for a cell count and viability. Adjust the cell density of the suspension to 2 to 3 x 10⁵ viable cells/mL in the shipping medium.
4. Incubate the culture, horizontally, at 37°C in a 5% CO₂ in air atmosphere. Maintain the cell density of the culture as suggested under the subculture procedure.



Subculturing Procedure

Cultures can be maintained by the addition of fresh medium or replacement of medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at 2 x 10⁵ viable cells/mL.

Maintain cell density between 2 x 10⁵ and 1 x 10⁶ viable cells/mL.

Medium Renewal: Twice per week



Cryopreservation Medium

Complete growth medium described above supplemented with 5% (v/v) DMSO. Cell culture tested DMSO is available as ATCC Catalog No. 4-X.



Comments

After the tenth passage, the parental KG-1 cells were cultured in two separate laboratories within the same department under identical conditions.

After 35 passages the cells in one laboratory expressed morphological differences from the parent line. The variant KG-1a was composed of undifferentiated promyeloblasts.

The cells did not stain for ASD chloroacetate esterase, alpha-naphthyl butyrate esterase or peroxidase.

Both populations exhibit many common characteristics. They share a similar doubling time, are negative for EBNA and VCA, express no surface immunoglobulins and exhibit identical HLA and isoenzyme profiles.

In contrast to the parental KG-1 (ATCC CCL-246) the KG-1a population is unresponsive to colony-stimulating factor in soft-agar culture and does not express the Ia-like antigen.

KG-1a cells are resistant to phorbol diester induced macrophage differentiation and proliferation of the cells is unaffected by the presence of phorbol diesters

.The KG-1a cells are morphologically, cytochemically, and functionally less mature than the parental KG-1.

Although derived from, and almost identical to, KG-1 (ATCC CCL-246) these cells do not spontaneously differentiate to granulocyte and macrophage like cells, do not express DR and do not respond to colony stimulating factor (CSF).



References

References and other information relating to this product are available online at www.atcc.org.



Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S.

Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.


ATCC Warranty




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The viability of ATCC® products is warranted for 30 days from the date of shipment, and is valid only if the product is stored and cultured according to the information included on this product information sheet. ATCC lists the media formulation that has been found to be effective for this strain. While other, unspecified media may also produce satisfactory results, a change in media or the absence of an additive from the ATCC recommended media may affect recovery, growth and/or function of this strain. If an alternative medium formulation is used, the ATCC warranty for viability is no longer valid.

Disclaimers

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Additional information on this culture is available on the ATCC web site at www.atcc.org.
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