



Product Sheet

# *Chlorella sp.* (ATCC®) 30562™)

## Please read this FIRST



## Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

## Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: *Chlorella sp.* (ATCC® 30562™)

American Type Culture Collection  
PO Box 1549  
Manassas, VA 20108 USA  
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## Description

**Strain Designation:** Syngen 2-3  
**Deposited Name:** *Chlorella sp.*  
**Depositor:** D Weis  
**Isolation:**  
Paramecium bursaria 2/III, Cleveland, OH, 1976

## Notes

Additional information on this culture is available on the ATCC web site at [www.atcc.org](http://www.atcc.org). While every effort is made to insure authenticity and reliability of strains on deposit, ATCC is not liable for damages arising from the misidentification or misrepresentation of cultures. ATCC recommends that individuals contemplating commercial use of any culture first contact the originating investigator to negotiate an agreement. Third party distribution of this culture is discouraged, since this practice has resulted in the unintentional spreading of contaminated cultures.

## Propagation

**Growth Conditions**  
**Temperature:** 25.0°C  
Duration: axenic

**Medium**  
ATCC® Medium 5: Sporulation agar

## Culture Maintenance

1. Screw the cap on tightly and vigorously agitate the culture.
2. Aseptically transfer a 0.1ml aliquot to 5 ml of fresh medium in a 16 x 125 mm screw-capped test tube.
3. Screw caps on loosely (loosened one-half turn) and incubate on a 15° horizontal slant at 25°C under a 14 hour light (~50 mEinsteins/m<sup>2</sup>/s irradiance)/10 hour dark cycle.
4. Subculture every 14-21 days.

## Cryopreservation

1. Harvest cells from a culture that is at or near peak density by centrifugation at 800 x g for 5 min.
2. Adjust the concentration of cells to 2 x 10<sup>6</sup> - 2 x 10<sup>7</sup>/ml in fresh medium.
3. While cells are centrifuging prepare a 10% (v/v) solution of sterile methanol in fresh medium.
4. Mix the cell preparation and the 10% methanol in equal portions. Thus, the final concentration will be 10<sup>6</sup> - 10<sup>7</sup> cells/ml and 5% (v/v) Methanol. The time from the mixing of the cell preparation and methanol stock solution to the beginning of the freezing process should be no less than 5 min and no greater than 15 min.
5. Dispense in 0.5 ml aliquots into 1.0 - 2.0 ml sterile plastic screw-capped cryovials (special plastic vials for cryopreservation).
6. Place the vials in a controlled rate freezing unit. From room temperature cool at -1°C/min to -40°C. If the freezing unit can compensate for the heat of fusion, maintain rate at -1°C/min through the heat of fusion. At -40°C plunge into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing apparatus. Place the apparatus at -80°C for 1.5 to 2 hours and then plunge ampules into liquid nitrogen. (The cooling rate in this apparatus is approximately -1°C/min.)
7. The frozen preparations should be stored in either the vapor or liquid phase of a nitrogen refrigerator. Frozen preparations stored below -130°C are stable indefinitely. Those stored at temperatures above -130°C are progressively less stable as the storage temperature is elevated. Vials should not be stored above -55°C.
8. To establish a culture from the frozen state place an ampule in a water bath set at 35°C. Immerse the vial just to a level just above the surface of the frozen material. Do not agitate the vial.
9. Immediately after thawing, do not leave in the water bath, aseptically remove the contents of the ampule and inoculate a 16 x 125 mm screw-capped test tube containing 5 ml of ATCC medium 847 broth or to the surface of ATCC medium 847 agar plate (20 x 100 mm Petri plate containing 20 ml of ATCC medium 847 agar).
10. Incubate the culture on a 15° horizontal slant at 25°C with the cap screwed on loosely (loosened one-half turn) and incubate under a 14 hour light (~50 mEinsteins/m<sup>2</sup>/s irradiance)/10 hour dark cycle. Alternatively, add the entire contents of the thawed ampule to the surface of a 20 x 100 mm Petri plate containing 20 ml of ATCC medium 847 agar. Spread the material evenly over the plate with a sterile cooled spread bar. Incubate in the same manner as the test tube culture.



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## **References**

References and other information relating to this product are available online at [www.atcc.org](http://www.atcc.org).

## **Biosafety Level: 1**

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

## **ATCC Warranty**

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## **Disclaimers**

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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at [www.atcc.org](http://www.atcc.org)

Additional information on this culture is available on the ATCC web site at [www.atcc.org](http://www.atcc.org).  
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