

**Product Sheet** 

## Ankistrodesmus braunii (ATCC® 12744™)

## Please read this FIRST



#### Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

### Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: *Ankistrodesmus braunii* (ATCC® 12744™)

American Type Culture Collection PO Box 1549 Manassas, VA 20108 USA www.atcc.org

800.638.6597 or 703.365.2700 Fax: 703.365.2750 Email: <u>Tech@atcc.org</u>

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## Description

Strain Designation: P202/78

Deposited Name: Ankistrodesmus braunii Brunnthaler

**Depositor:** RW Krauss

Isolation:



## Propagation

Growth Conditions Temperature: 25.0°C Duration: axenic

#### Medium

ATCC® Medium 5: Sporulation agar

#### Instructions for Complete Medium

ATCC Medium 5



#### **Protocols**

Frozen ampules packed in dry ice should either be thawed immediately or stored in liquid nitrogen. If liquid nitrogen storage facilities are not available, frozen ampules may be stored at or below -70°C for approximately one week. Do not under any circumstance store frozen ampules at refrigerator freezer temperatures (generally -20°C). Storage of frozen material at this temperature will result in the death of the culture

- 1. To thaw a frozen ampule, place in a 35°C water bath, until thawed (2-3 min). Immerse the ampule just sufficiently to cover the frozen material. Do not agitate the ampule.
- 2. Immediately after thawing, aseptically transfer the entire contents to a single 16 x 125 mm screw-capped test tube containing 5 ml of ATCC Medium 5 broth. Incubate the tube on a 15° horizontal slant with the cap screwed on loosely (loosened one half turn) at 25°C under a 14 hour light (~50 mEinsteins/m²/s irradiance)/10 hour dark cycle. Alternatively, add the entire thawed contents to the surface of a 20 x 100 mm Petri plate containing 20 ml of ATCC medium 5 agar. Wrap the plate culture with parafilm and incubate upright under the same light/dark cycle as specified for a test tube culture.

### **Culture Maintenance**

- 1. For a broth culture, inoculate a tube of fresh broth medium with 0.1 ml from a growing culture at or near peak density. For a plate culture, transfer cells with an inoculating loop to a plate of fresh agar medium from a growing culture at or near peak density.
- 2. Incubate at 25°C under a 14 hour light (~50 mEinsteins/m²/s irradiance)/10 hour dark cycle. In the case of a broth culture, screw tube cap on loosely (loosened one-half turn) and incubate on a 15° horizontal slant.
- 3. Subculture as necessary (i.e., every 14-21 days in broth media).



## Cryopreservation

- 1. Harvest cells from a culture that is at or near peak density by centrifugation at 700-800 x g for 5 min.
- 2. Adjust the concentration of cells to  $2 \times 10^6$   $2 \times 10^7$ /ml in fresh medium.
- 3. While cells are centrifuging prepare a 14% (v/v) solution of sterile DMSO in fresh medium.
- 4. Mix the cell preparation and the 14% DMSO solution in equal portions. Thus, the final concentration will be  $10^6 10^7$  cells/ml and 7% (v/v) DMSO. The time from the mixing of the cell preparation and DMSO stock solution to the beginning of the freezing process should be no less than 5 min and no greater than 15 min.
- 5. Dispense in 0.5 ml aliquots into 1.0 2.0 ml sterile plastic screw-capped cryules (special plastic vials for cryopreservation).
- 6. Place the vials in a controlled rate freezing unit. From room temperature cool at -1°C/min to -40°C. If the freezing unit can compensate for the heat of fusion, maintain rate at -1°C/min through the heat of fusion. At -40°C plunge into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing apparatus. Place the apparatus at -80°C for 1.5 to 2 hours and then plunge ampules into liquid nitrogen. (The cooling rate in this apparatus is approximately
  - -1°C/min.)
- 7. The frozen preparations should be stored in either the vapor or liquid phase of a nitrogen refrigerator. Frozen preparations stored below -130°C are stabile indefinitely. Those stored at temperatures above -130°C



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are progressively less stabile as the storage temperature is elevated. Vials should not be stored above -55°C.

- 8. To establish a culture from the frozen state place an ampule in a water bath set at 35°C. Immerse the vial just to a level just above the surface of the frozen material. Do not agitate the vial.
- 9. Immediately after thawing, do not leave in the water bath, aseptically remove the contents of the ampule and add to a 16 x 125 mm screw-capped test tube containing 5 ml of ATCC medium 5 broth or to the surface of an ATCC medium 5 agar plate (20 x 100 mm Petri plate containing 20 ml of ATCC medium 5 agar).
- 10. Incubate the culture at 50-100  $\mu$ Einsteins/m²/s irradiance at 25°C. Maintain under a 14/10h light-dark photoperiod.



## References

References and other information relating to this product are available online at www.atcc.org.



## Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

### **ATCC Warranty**

The viability of ATCC® products is warranted for 30 days from the date of shipment, and is valid only if the product is stored and cultured according to the information included on this product information sheet. ATCC lists the media formulation that has been found to be effective for this strain. While other, unspecified media may also produce satisfactory results, a change in media or the absence of an additive from the ATCC recommended media may affect recovery, growth and/or function of this strain. If an alternative medium formulation is used, the ATCC warranty for viability is no longer valid.

#### **Disclaimers**

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Additional information on this culture is available on the ATCC web site at www.atcc.org.

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