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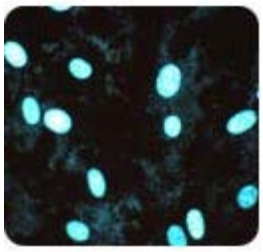
microscoop



Titered Mycoplasma Panel – *Now Available!*

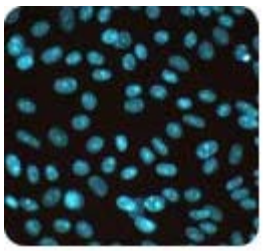
Mycoplasma are frequent contaminants of cell cultures, often causing a wide variety of adverse effects on cellular function and growth. To reduce the risk of contamination, cell lines should be routinely screened for infection. To aid in the detection of Mycoplasma, ATCC offers the ATCC® Titered Mycoplasma Reference Strains Panel (ATCC® MP-7™). This panel includes 10 different species of Mollicutes, including the four species most commonly associated with cell culture contamination[1].

Generally, the preparation of mycoplasma reference strains with low ratios (≤10:1) of genome copy number to colony forming units (CFU) is the most reliable way to evaluate the detection limit of nucleic acid-based mycoplasma testing methods[2]. Therefore, each ATCC mycoplasma reference strain is evaluated for genome copy number, and cultures are quantified by CFU. The ratio between copy number and CFU are calculated and expected to fall below 10:1.

Learn more ▸

[1]McGarrity GJ and Kontani H., Cell culture mycoplasma. In: The Mycoplasma, Vol. IV. Razin S and Barile MF, eds. New York: Academic Press; 1985: pp. 353-390.

[2]Dabrazhynetskaya A, et al. Preparation of reference strains for validation and comparison of mycoplasma testing methods. J Appl Microbiol. (2011) 111(4): 904-914.



VERO Cells Adapted for Serum-Free Media – New Prospects for Virus Production

The development of cell culture media that does not require supplementation with sera or animal-derived components is of critical importance in increasing the safety and efficacy of reagents produced for viral therapy and vaccination. A primary concern among vaccine developers is that sera and animal-derived components may serve as a potential source of cryptic bacterial, fungal, or viral contamination. Additionally, serum-based media is often physiologically inconsistent due to batch-to-batch variations in

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Microbial Panels – *Available Now!*

Six panels of ATCC Genuine Cultures grouped together based on their utility in infectious disease research applications

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ATCC Publications

ATCC® Bacterial Culture Guide – *Coming this fall in BioTechniques!*

A manual featuring tips and techniques for culturing bacteria and bacteriophages

View from the Petri Dish

An ATCC blog highlighting the intersection between microbiology and society

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composition. This latter concern can stem from disparities in protein content, which may hinder product purification, or the presence of varying amounts of inhibitors such as endotoxins and hemoglobin.

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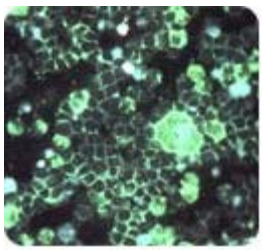
To combat these concerns, ATCC has developed a serum-free system for *in vitro* viral or protein production. The system includes a serum-free, animal component-free Vero derivative, Vero-SF-ACF (ATCC® CCL-81.5™), and associated culture medium, VeroPlus SFM (ATCC® ACS-4001™).

Together, these products provide a complete solution to serum-free, animal component-free growth of virus, or as a platform for protein production.

Learn more ▸



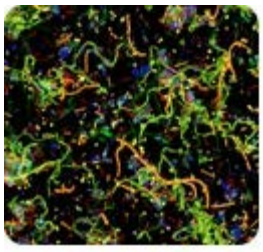
High-titer, Purified Virus – *NEW!*

Respiratory Syncytial Virus (RSV) is the most common cause of bronchiolitis and pneumonia in infants less than 1 year of age, and has been recognized as an important cause of respiratory illness in adults[1]. There is no vaccine for RSV, and previous exposure does not provide complete immunity against re-exposure.

ATCC now offers a high-titer, sucrose-cushion purified RSV A-2 strain (ATCC® VR-1540P™) for use in RSV research. Each ampoule contains 1.0 mL of purified virus at a concentration >2 x 10^8 PFU/mL, and is useful in a variety of applications where high titer material is required, such as challenge studies and viral clearance assays.

Learn more ▸

[1]Respiratory Syncytial Virus (RSV): Incidence and Infection. CDC. Available on the web at http://www.cdc.gov/rsv/about/infection.html.
Photo courtesy Dr. Craig Lyerla, CDC



ATCC Photo Contest –Congratulations to Alex Valm for Best in Show!

Congratulations to Alex Valm for his photograph illustrating the use of Combinatorial Labeling and Spectral Imaging Fluorescence In situ Hybridization (CLASI-FISH)! This innovative technique, developed at the Marine Biological Laboratory (MBL), allows for a systems-level analysis of microbial community spatial arrangements via combinatorial labeling and spectral imaging[1]. The organisms portrayed in this spectral fluorescence image are a representative mixture of 15 different laboratory-propagated species characteristic of the human oral microbiome. Both Gram-positive and Gram-negative microbial cells were labeled with a combination of taxon-specific probes, and then examined using a novel method of computational image analysis to distinguish fluorophores with highly overlapping emission spectra. This unique image reveals previously unknown interspecies associations and demonstrates that multiple taxa in the plaque community can be imaged simultaneously. This fresh approach to analyzing biological organization holds potential for future applications in systems biology.

Learn more ▸

[1]Valm AM, et al., Systems-level analysis of microbial community organization through combinatorial labeling and spectral imaging. PNAS (2011) 108(10): 4152-4157.

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