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Technical Data Sheet:

3 Strain Tagged Whole Cell Even Mix

| | |
|---------------------|---|
| ATCC® Number | MSA-2014™ |
| Components | 33.3% <i>Escherichia coli</i> with Tag 1 33.3% <i>Clostridium perfringens</i> with Tag 2 33.3% <i>Staphylococcus aureus</i> with Tag 3 |
| Product Description | We have engineered three bacterial genomes (<i>Escherichia coli</i> , <i>Staphylococcus aureus</i> , and <i>Clostridium perfringens</i>) to contain a unique synthetic DNA tag that can be detected via 16S rRNA profiling and whole genome sequencing assays. The unique tag comprises four artificial variable regions (corresponding to the V1 through V4 regions in the 16S rRNA gene) flanked by conserved regions for PCR amplification, thereby enabling the identification of spike-in reads during the analysis of next-generation sequencing data. Using these recombinant bacterial strains, we prepared an even mixture of three tagged strains for use as a spike-in control in microbiome research. |

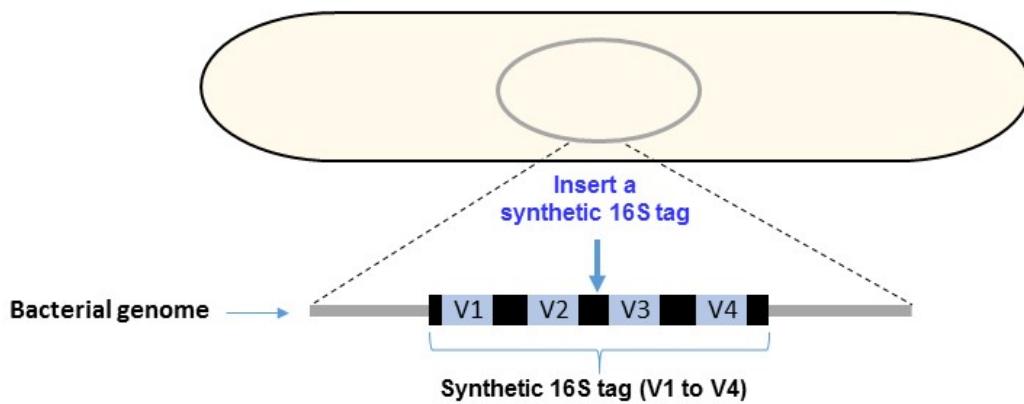


Figure 1. Production of tagged strains. ATCC created 3 unique synthetic 16S tag sequences mimicking the native 16S rRNA gene from 3 bacterial strains (*E. coli*, *C. perfringens*, and *S. aureus*). Each tag consists of 4 artificial variable regions (corresponding to V1 through V4 of the 16S rRNA gene) flanked by conserved regions for PCR amplification. Each tag sequence was integrated into the genome of their cognate strains to create three tagged strains. The tag sequences are provided below.

Escherichia coli 16S Tag 1

AAATTGAAGAGTTGATCATGGCTCAGATTACATCGGGATTGAGGGGCCACAGGAGGCATCACTGACATGCCCTATCGTGAT
AGGGGCTAGCTACAGCAGGTGGCGACGGGTGAGTAATGTGAGTAGCGAAAAGTCATGGCTAAAGGTACTGTTCGTCAT
CCGATAAGATTGACGGAAATTGATTCTCACACGTCCCAGTGTGGGAGCCGCGACCGTCACAGGTGAGAAATCTCTCTAAAAA
GATTTATGCCATAGTAGATTCACTACAAATCCAGACACACGGGTAGTCGACTCGATTCTAATATCTTATGGAT
CCTAATCTAGACTCCTACGGGAGGCAGCAGTGGTCACTGCATGATCAACCAAAGGTGTTCCGGCTACGTTCAATTGAGA
ACGGCGGTCTGGAGCATGAAAGGACGAGACGGCATTAGGACTTGCCAGGCGATGTATGCTGATGGGAAGTAGGGAAACAT
GAGAGGCGCTCTAAATCCTCCGTGCCAGCAGCGCGTAATACGACGTGATCATGGTAGACGTCCACTTTACCGT
GTGTGGCGATGAGGGATGCAAGAGGATCATTGGTAGCGTATTGTGACTCTGAAACTAAGGCAGGAGACCAGGTTAGGT
AACCGGTGCGTAGATAATCCGAGACCCGCGTGCCTAGCTAAGGAAAGGGTATGCCGACGTTGAAACAGATAAC
CTAAACCAGGTGACGCCTATAGAGAACGACGAAGCTAATCTATCCGAAACAGGATTAGATAACCTGGTAGTCCACGCCGTA
AACGATGTCGAC

Clostridium perfringens 16S Tag 2

AAATTGAGAGTTGATCCTGGCTCAGGATGTACGACGAGGATTAGGTGGGAGGGACTGGCACGAGTAGTATACGGTTT
AAAAAGTATTGGAGCGGGCGACGGGTGAGTAACACCTTAAGGCGGGTTGGGCGTCCGAAACATACGATCCGCTGGCAA
GGTGCAGTGGCAGACCTGGCGGGAGTACCGGAGCATAAAGGATTGCAAGCACGTTAGCGGTTAGGGAGGCTGGCT
GCAAGCGCGAAGGCCAGCGCTTACCGTGCATGGTAGCAAATGAGTCCCTGACCGGACCCACATAATCGTACGTCGTAT
CCTCTCTACAGACTCCTACGGGAGGCAGCAGTGGTAGGCTCCATAATGCTAGTCGACCTCGTGCCTGGTCGCTCACC
GTTCACAAGAACTCCTGCCAACGTTAACGGCTAGCGCTAACGTTAACGGCGTAAACGGTTGCGCTGCGGAAATGGCGT
GCCAGCAGCCGCGTAATACGTGAAAGTTGAACTGAAATCAAGAAAGTTAACAGGGTTGCGTGCAGGAAATGGCGT
AAAACAAATTGAGCGGGTGGGAACAAACGAAGATGGTAGTTCTAGGTTGAGATAACTCCGTAACCTAGCTGCGGTA
AGGAAGTGTGCGCTACGGGATAGCGATATGACTGGCTAACAGAGCTCGGATTCTAACGCTGCTGGTCGAATGTAAGCAC
GACACATCTAATCCGAAACAGGATTAGATAACCTGGTAGTCCACGCCGAAACGATGAATAC

Staphylococcus aureus 16S Tag 3

TTTATGGAGAGTTGATCCTGGCTCAGGATCATTACCTCGATTGAGATAAGCAAACAAGTCGCCTAGTGAAGGCACGTCT
GATCGTCACACCTGGAAGCGCGGACGGGTGAGTAACACCTACAGTATGGTATCCTGGATATTCACTTAGTATTCACTGGT
CCTCCATAGTAACGGCGTCTCGATAAGCGGTCCCAGGATTAGATAAAATCGACTGGCCTATGAATGCCCTAGGCACACGAGTG
GGATACGATCAAGAGTGACACAGAGCGAACCCGTCAAATGTCGGAAATATGGGTGTATGACGTACCCAGAGTGGTATA
GAAAATCCTGTGAAAGACTCCTACGGGAGGCAGCAGTAGAGGGTGGTGCAGGGCACAGTTAGGTTCATGATCATGGCGACAG
AGAAGCTGGGACAGCGAACGGTCAAACCAACACAGTAGCAGTCACACTAACGCTGGGAGGTGATGGTTCTGCGCATAGGG
TATTAAACACACTGTAATTGATCCTGATACGTGCCAGCAGCGCGTAATACGAAGTCTATTATCTGGCATGCTCGTGG
AGCTCAGACCGCTGAGGTGAAAGTATAAAGTGTGCGAGGATCGAGATAAACGGCTATATATGTGATGGGACCAGTTAAA
ATACCGGATATGCACTGACGGACCAGGAGGGACGGAGAGGGACCTCTTACTGCAATGTTCAATGGGAGGTCACTGAC
CAGAGAGTAGGTAATACTGTGAGACGAAGAGAAAGAGATTGTGAATCCTCAAACAGGATTAGATAACCTGGTAGTCCACGC
CGTAAACGATGAGTGC

Table 1. Composition of the 3 Strain Tagged Whole Cell Even Mix (ATCC® MSA-2014™)

| Species | Gram Stain | Genome size (Mb) | Tag size (bp) | G/C Content (%) | Number of 16S Copies | Number of Tag Copies | Cells per vial* |
|-------------------------------------|------------|------------------|---------------|-----------------|----------------------|----------------------|-----------------|
| <i>Escherichia coli</i> Tag1 | Negative | 4.59 | 829 | 50.8 | 7 | 1 | 2×10^7 |
| <i>Clostridium perfringens</i> Tag2 | Positive | 3.25 | 799 | 29.0 | 10 | 1 | 2×10^7 |
| <i>Staphylococcus aureus</i> Tag3 | Positive | 2.70 | 833 | 32.8 | 6 | 1 | 2×10^7 |

*Specification Range: Total of 6.0×10^7 cells/vial ± 1 log. Whole cell concentrations indicate ATCC® manufacturing specifications and are provided as a reference only.

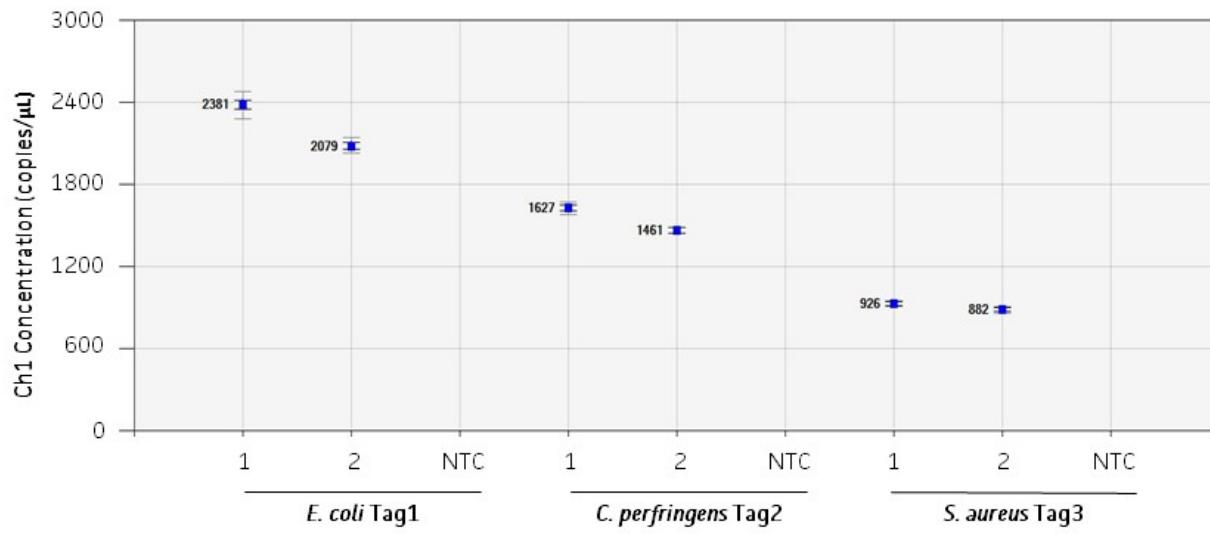


Figure 2. Genome copy number of individual bacteria determined by Droplet Digital™ PCR (ddPCR™; Bio-Rad) assay after extraction of DNA from ATCC® MSA-2014™. Total DNA was extracted from two vials of ATCC® MSA-2014™ by using a DNeasy® Powelyzer® Microbial Kit (QIAGEN) and eluted in a final volume of 50 μL according to the manufacturer's instruction. The concentration of extracted DNA was assayed by PicoGreen® dsDNA quantitation assay (Thermo Fisher Scientific); Vial 1 (1) = 9.7 ng/μL and Vial 2 (2) = 8.6 ng/μL. Both vials were diluted 100-fold and assayed in triplicate (1, 2) along with a no template control (NTC). The primer and probe set used for the three tag amplicons were the following: *E. coli* Tag1 (Forward: 5'-GAACGGGTGAGTAATGTGAGTAG -3'; Reverse: 5'-CGGGACGTGTGAGAATCAAT -3'; Probe: 5'-TCGGATGACGAAACAGTACCTTAGCC -3'), *C. perfringens* Tag2: (Forward: 5'-TAACCATGCACGGTAAAGCG -3'; Reverse: 5'-AAGGATTGCAAGCACGTT -3'; Probe: 5'-TTCGCGCTTGAGCCCAGGCTCCCT -3'), and *S. aureus* Tag3: (Forward: 5'-AATACGCGGATATGCAGTGC 3'; Reverse: 5'-CTGACCTCCATTGAACGATTGC -3'; Probe: 5'-AGAGGTCCCTCTCCGTCCTCCTGGTCCGT -3'). The concentration (genome copies/μL) represents the average of three wells in triplicate assays. The total genome copy number per organism per vial was calculated based on the average of triplicate reactions per specific ddPCR™ assay (Table 2).

Table 2. Genome copy number of individual bacteria in the 3 Strain Tagged Whole Cell Even Mix

| Species | Genome copy number per vial | |
|-------------------------------------|-----------------------------|-----------------------------|
| | Vial 1 | Vial 2 |
| <i>Escherichia coli</i> Tag1 | $4.75 \pm 0.18 \times 10^7$ | $4.16 \pm 0.10 \times 10^7$ |
| <i>Clostridium perfringens</i> Tag2 | $3.25 \pm 0.08 \times 10^7$ | $2.92 \pm 0.04 \times 10^7$ |
| <i>Staphylococcus aureus</i> Tag3 | $1.85 \pm 0.03 \times 10^7$ | $1.76 \pm 0.04 \times 10^7$ |

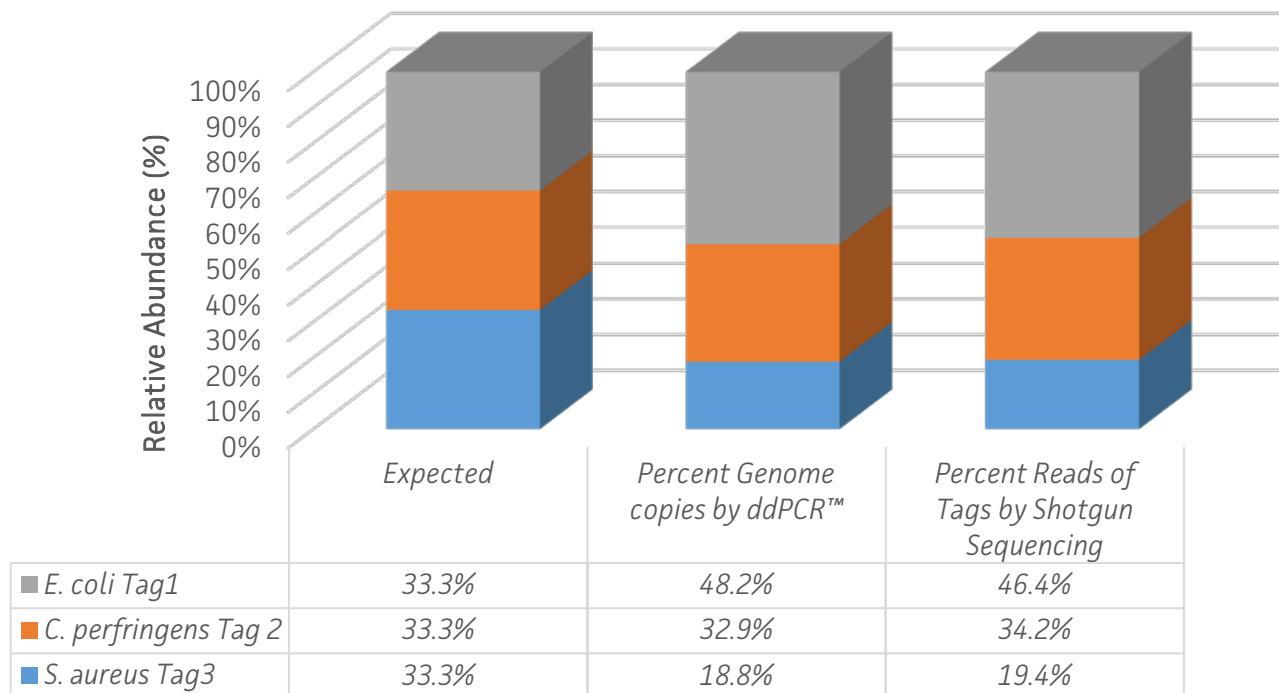


Figure 3. Relative abundance of three genomes extracted from ATCC® MSA-2014™. Shotgun metagenomic sequencing was performed on the Illumina® platform (Nextera XT DNA Library Preparation Kit and MiniSeq 2x150). Total shotgun reads (7.27 million) were mapped to unique tag sequences (total = 1856 reads), and the percent of reads mapped to individual tags were calculated. Reads mapping to tag sequences was performed via Bowtie2 tool in Geneious 11.1.4 Software. Note: The discrepancy between the expected and observed ratios may be attributed to cell counting and extraction efficiency, which is related to the diverse physical properties of the bacterial species.

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