



# Tera-1

HTB-105™

## Description

**Organism:** *Homo sapiens*, human

**Tissue:** Testis

**Age:** 47 years

**Gender:** Male

**Morphology:** Epithelial-like and/or rounded

**Growth properties:** Adherent

**Disease:** Carcinoma; Malignant embryonal

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## Storage Conditions

**Product format:** Frozen

**Storage conditions:** Vapor phase of liquid nitrogen

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## Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

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## BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

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## Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at [www.atcc.org](http://www.atcc.org).

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## Growth Conditions

**Temperature:** 37°C

**Atmosphere:** 95% Air, 5% CO<sub>2</sub>

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## Handling Procedures

### Unpacking and storage instructions:

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

**Complete medium:** The base medium for this cell line is ATCC-formulated McCoy's 5a Medium Modified , Catalog No. 30-2007. To make the complete growth medium, add

the following components to the base medium: fetal bovine serum to a final concentration of 15%.

**Handling Procedure:**

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a **37°C** water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. It is recommended that the cryoprotective agent be removed immediately. Transfer the freshly thawed cells to a sterile centrifuge tube containing 9 mL of complete growth medium. The DMSO should be removed by gentle centrifugation, approximately 125 x g, for 5 to 10 minutes. Discard the supernatant, and resuspend the cells in 1 or 2 mL of complete growth medium.
4. Transfer the cell suspension into the culture flask containing the pre-incubated complete growth medium and mix thoroughly by gentle rocking. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
5. Incubate the culture at 37°C in a suitable incubator. A 5% CO<sub>2</sub> in air atmosphere is recommended if using the medium described on this product sheet.

**Subculturing procedure:**

Subcultures are prepared by enzymatic dissociation using 0.25% Trypsin/0.53 mM EDTA (Trypsin-EDTA Solution, 1X, ATCC-30-2101™). Volumes are given for a 75 cm<sup>2</sup> flask. Corning® T-75 flasks (catalog # 430641) are recommended for subculturing this product. Increase or decrease the amount of dissociation medium needed proportionally for culture vessels of other sizes.

1. Remove and discard culture medium.

2. Briefly rinse the cell layer with dPBS solution to remove all traces of serum which contains trypsin inhibitor.
3. Add 3.0 to 5.0 mL of Trypsin-EDTA solution to flask and place the cells at 37°C to facilitate detachment (usually within 5 to 10 minutes).

**NOTE:** If debris is present the dissociation could take longer.

4. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting and transfer to an appropriate size sterile centrifuge tube.
5. Centrifuge at 150-400 g for 8-12 minutes and remove dissociation agent.
6. Resuspend resulting pellet in complete culture media and add appropriate aliquots of the cell suspension to new culture vessels with fresh medium.
7. Incubate cultures at 37°C.

**Subcultivation Ratio:** A subcultivation ratio of 1:2 to 1:3 is recommended.

**Medium Renewal:** It is imperative **NOT** to perform fluid changes in between subcultures, unless a great amount of debris is observed. **Note:** HTB-105 displays higher level of proliferation in an acidic environment.

Subculture every 7 to 10 days or before the cells reach full confluence.

**Reagents for cryopreservation:** Complete growth medium supplemented with 5% (v/v) DMSO (ATCC 4-X)

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## Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: Tera-1 (ATCC HTB-105)

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## References

References and other information relating to this material are available at [www.atcc.org](http://www.atcc.org).

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