



# ***Nitrobacter winogradskyi*** **Winslow et al.**

**25391™**

## **Description**

*Nitrobacter winogradskyi* strain Nb-255 [1] is a whole-genome sequenced bacterial type strain.

**Strain designation:** Nb-255 [1]

**Deposited As:** *Nitrobacter winogradskyi* Winslow et al.

**Type strain:** Yes

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## **Storage Conditions**

**Product format:** Frozen

**Storage conditions:** -80°C or colder

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## **Intended Use**

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

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## **BSL 1**

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

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## Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at [www.atcc.org](http://www.atcc.org).

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## Growth Conditions

**Medium:**

ATCC Medium 480: *Nitrobacter* medium 203

**Temperature:** 26°C

**Atmosphere:** Aerobic

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## Handling Procedures

1. Allow vial to thaw. Transfer the entire contents into 5.0 mL of #480 broth contained in a large test tube (20 X 150 mm).
2. Incubate tube at 26°C, static, in the dark, and in a slanted position to increase surface area.
3. Monitor culture growth daily by assaying for the disappearance of NO<sub>2</sub> in the

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culture medium. Withdraw 0.1 mL from the culture and place in a small test tube. Add one drop of NO<sub>2</sub> Reagent A followed by one drop of NO<sub>2</sub> Reagent B, mixing after each addition. Pink color development occurs within 10 minutes and is directly related to amount of NO<sub>2</sub> present. Initial tests will probably be dark magenta reflecting the 3 mM NO<sub>2</sub> concentration of medium #480; but as culture grows and NO<sub>2</sub> is oxidized, color will become magenta, pink and finally clear. This process can take from 3 to 20 days depending on the viability of the stock.

4. When all NO<sub>2</sub> has been oxidized, inoculate a 250 mL flask containing 100 mL of medium #480 with the 5.0 mL tube culture. Wrap the flask in foil to protect from light and incubate at 26°C with gentle shaking. Monitor NO<sub>2</sub> concentration daily.
5. When NO<sub>2</sub> is again depleted in the culture medium, either transfer the culture to fresh media or feed the culture by adding additional NO<sub>2</sub>. A 10% inoculum is recommended for transfers. To feed cultures, aseptically add sterile NaNO<sub>2</sub> solution, returning NO<sub>2</sub> concentrations to approximately 10 mM (3.5 mL of 2% NaNO<sub>2</sub> per 100 mL culture). Cells should be harvested or sub-cultured to fresh media when cultures can utilize 10 mM NO<sub>2</sub> in less than 24 hours and cell numbers are greater than 10 cells per field at 1000X magnification.
6. Cells can be harvested by centrifugation. For long-term storage, 1.0 mL aliquots of concentrated cells can be frozen with glycerol as the cryoprotectant at a final concentration of 10%. Frozen vial should be stored at -70°C or below for best preservation.

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## Notes

This item does not grow on agar.

When a strain is received as a test tube culture, transfer to fresh media upon arrival.

(See step 4.)

NO<sub>2</sub> Reagent A:

Sulfanilic acid, 8 g

5N Acetic acid, 1.0 L

NO<sub>2</sub> Reagent B:

N,N-Dimethyl-1-naphthylamine, 6.0 ml

5N Acetic acid, 1.0 L

*5N acetic acid consists of 1 part glacial acetic acid to 2.5 parts distilled water.*

NO<sub>2</sub> Reagents A and B may also be purchased from Remel.

Additional information on this culture is available on the ATCC® web site at [www.atcc.org](http://www.atcc.org).

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## Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Nitrobacter winogradskyi* Winslow et al. (ATCC 25391)

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## References

References and other information relating to this material are available at [www.atcc.org](http://www.atcc.org).

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